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**TAXONOMY OF SELECTED SPECIES OF *MYXOBOLUS* (MYXOZOA) FROM
FRESHWATER FISHES IN CANADA; APPLICATIONS OF LIGHT
MICROSCOPY AND MOLECULAR METHODS**

A Thesis

Submitted to the Graduate Faculty

In Partial Fulfillment of the Requirements

for the Degree of

Master of Science

in the Department of Biology

Faculty of Science

St. Mary's University

Halifax, Nova Scotia

Russell H. Easy

Halifax, Nova Scotia

Date: April 22, 2004

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LIGHT MICROSCOPY AND MOLECULAR METHODS

Name of Author: Russell H. Easy

Department: Biology

Degree: Master of Applied Science

Year: 2004

Signature

Date: April 22, 2004

St. Mary's University

Faculty of Science

Halifax

Abstract

Russell H. Easy

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Species of *Myxobolus* (Myxozoa) are compared on the basis of spore morphology, site of development, host species and sequence data of the 18S rDNA. The first situation compares the morphology and molecular features of *Myxobolus procerus* from large intercellular and small intracellular cysts in trout-perch (*Percopsis omiscomaycus*) muscle. The data suggests that spores from each cyst type represent a single species of *Myxobolus* with variable cyst phenotypes. Interestingly, spores from the small cysts were shorter than those from the large cysts implying ecophenotypic variation. The fact that the two cyst types did not occur independently of each other suggests that they are a single species. The observed differences in the morphological and molecular data suggest species divergence. The second situation provides a comparison between *Myxobolus neurophilus* from the optic tectum of yellow perch (*Perca flavescens*) and specimens that were tentatively identified as *M. neurophilus* from the tectum of Johnny darter (*Etheostoma nigrum*). Although the morphology of the spores of these samples is very similar, molecular information identifies the latter parasite as a possible new species of *Myxobolus*. The morphological and molecular similarities between these three species are examined on both an individual basis and between other species in the genus *Myxobolus*. Phylogenies are drawn from the molecular data and a phylogenetic tree is constructed that includes sequences presented in the current work which have not been analysed at this level. The validity of traditional taxonomic methods and their use in classification of the myxobolids is discussed. Alternately modern techniques of molecular biology and their role in the taxonomy of these parasites are evaluated.

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LIST OF ABBREVIATIONS

BLAST	Basic Local Alignment Search Tool
CISTI	Canadian Institute for Scientific and Technical Information
DNA	Deoxyribonucleic Acid
IMB	Institute for Marine Biosciences
ITS	Inter Transcribed Spacer region
NCBI	National Center for Biotechnology Information
NRC	National Research Council
DNTP	deoxyribonucleotide triphosphate
PAUP	Phylogenetic Analysis Using Parsimony
PCR	Polymerase Chain Reaction
PFGE	Pulsed Field Gel Electrophoresis
RAPD	Randomly Amplified Polymorphic DNA
RFLP	Restriction Fragment Length Polymorphism
RNA	Ribonucleic Acid
SSU	Small Subunit
SNPs	Single Nucleotide Polymorphisms
<i>Taq</i>	<i>Thermus aquaticus</i>
X-gal	5-bromo-4-chloro-3-indolyl- β -D-galactoside

1.0 Introduction

The phylum Myxozoa is a diverse group of approximately 1350 described species all of which are parasites (Kent et al. 2001). Their hosts include annelids, bony fishes, amphibians and reptiles (Andree et al. 1999 a). They can be considered ubiquitous within a wide range of environments and occupy an even wider range of hosts.

Myxozoans are considered tissue specific in their site of development (Molnár 1994), but as a group they can be found in a variety of organs within individual fish hosts. Numerous species can be found in the lumen of the gall bladder, kidney tubules, and digestive tract (coelozoic species), whereas others are known from sites within tissues of the digestive, reproductive, muscular, circulatory, and skeletal systems including the skin (histozoic species). Although myxozoans are generally not pathogenic there are some major exceptions. For example, Whirling disease in trout is caused by *Myxobolus cerebralis* and enzymatic lysis or soft tissue disease of salmon is caused by *Kudoa* and *Hexacapsula* species.

Until the early 1990's the Myxozoa were usually grouped with the protists (Kent et al. 2001). Pioneering studies into the morphology of myxozoans, however, identified characteristics typical of multicellular animals including structural and developmental patterns similar to those of the phylum Cnidaria (Siddall et al. 1995). Cnidarians are those animals armed with stingers or nematocysts and include the jellyfish and corals. Siddall et al. (1995) identified

myxozoan “polar capsules” within “capsulogenic cells” that bore a distinct homology to cnidarian nematocysts. The results appeared to group the Myxozoa within a clade of highly derived parasitic cnidarians (Siddall et al. 1995). In phylogenetic analyses, Siddall et al. (1995) included the cnidarian narcomedusan *Polypodium hydriforme* and considered it essential to the resolution of the myxozoans as cnidarians. However, Hanelt et al. (1996) argued that the small subunit (SSU) 18S ribosomal DNA (rDNA) sequence of *P. hydriforme* is too different to be included in a proper phylogenetic comparison. It was concluded by Hanelt et al. (1996) that data collected through BLAST searches using *Polypodium* 18S rDNA sequences differed from existing 18S sequences to such an extent that its grouping within the myxozoans must be due to “long branch abstraction” (Siddall and Whiting 1999). Conflicts in the molecular data may arise from the long-branch nature of the Myxozoa as well as the low “taxon density” common to most molecular analyses (Zrzavý et al. 1998). Siddall and Whiting (1999), subsequently, concluded that there is no support for the suggestion that myxozoans are unrelated to cnidarians, stating that homology in ontogenetic stages and intercellular parasitism common to myxozoans and cnidarians cannot be due to long-branch abstraction (Siddall and Whiting 1999). Figure 1 illustrates parsimony analysis of SSU 18S rDNA sequences for the phylum Myxozoa as constructed by Siddall and Whiting (1999). Since these molecular lineages have been defined using a single data set, this cannot be the final word on myxozoan origins (Kent et al. 2001).

It has recently been suggested that the Myxozoa originated from a bilaterian ancestor such as *Buddenbrockia*, a nematode-like parasite of freshwater bryozoans (Okamura et al. 2002). Phylogenetic analyses based on SSU 18S rDNA sequences and one on *Hox* gene sequences further supported this hypothesis (Canning et al. 2002). Zrzavý and Hypša (2003) support this relationship of the Myxozoa while allowing at the same time for a homology between polar capsules of the myxozoan and cnidarian nematocysts. Successive clades of nematocyst bearing animals including the Myxozoa, *Polypodium* and Cnidaria, appear to bridge the gap between the animals like *Buddenbrockia* and more primitive animals (Zrzavý and Hypša 2003).

Life cycles are known for only about 25 myxozoans (Oumouna et al. 2002) as summarized in Table 1. Wolf and Markiw (1984) elucidated the biphasic life cycle of *M. cerebralis* including development into an actinosporean (triacinomyxon) within an annelid host prior to infecting the fish host (Wolf and Markiw 1984) (Fig. 2). Spores from salmonid hosts having Whirling disease are not infective to healthy fish. It is only after the spores have been ingested by annelid worms and have gone through a series of complex stages of development does an infective triacinomyxon, or TAM, emerge. Spores are liberated when dead fish decay. Or, if a predator eats a fish the spores are passed out with the predator's feces. The tubificid worms then ingest the spores and the binucleate sporoplasm that emerges from the spores invades the gut epithelium of the host worm (Brinkhurst 1996).

In the majority of cases species have a biphasic cycle involving myxosporean and actinosporean phases. Issues concerning nomenclature of the myxozoans have been raised in response to the respective life stage the organism is presently undergoing. Kent et al. (2001) proposed that based on their interpretation of the International Code of Zoological Nomenclature that the generic names of all actinosporeans should be treated as collective groups (Kent et al. 2001). This would eliminate the discrepancies arising from alternate myxosporean generic names. There is concern over this proposed change, however, because it is uncertain that all species of Actinosporea produce corresponding myxosporean stages in their life cycles (Morris et al. 2002). Those species of *Myxobolus* in which separate life stages have been identified should not be described based solely on their actinosporean forms. Major discrepancies in nomenclature arise from interpretations of the significance of new and unusual actinosporean forms and not simply on the interpretations as defined by the International Code of Zoological Nomenclature (Kent et al. 2001).

Conventional light microscopy provides information that allows significant functional resolution at the species level of the Myxozoa (Oumouna et al. 2002). However, the use of these classical zoological methods is sometimes very difficult to accurately characterize morphologically similar myxozoans inhabiting taxonomically related host species (Molnár et al. 2002). The morphology of myxosporeans is thought to vary significantly with host, environment within which they are found and tissue. However, traditional diagnostic procedures are based solely on these characteristics (Yokoyama et al. 1997).

Species concepts based on traditional morphological variance are continually being modified to include data obtained through analysis of an organism's genotype. There are few homologous morphological characters that can be compared among all living organisms whereas a number of genes with fundamental biochemical functions are found within all species (Hillis and Dixon 1991). The use of rDNA sequences is a well-accepted method to resolve taxonomic relationships at the intrageneric level as well as at higher taxonomic levels (Kent et al. 2001). The SSU 18S rDNA has become the genetic region of choice by researchers and as such was used in this work as well. Sequence data from regions distributed along the ribosomal RNA provide a molecular template that can be used as a tool for identifying relationships among organisms (See Appendix A). This is possible because the rRNA is universally distributed, is functionally equivalent, and occurs in large enough sizes that interspersed patterns of conserved and non-conserved regions allow for statistical analyses that provide valid interpretations of genetic similarities between species (Frasca et al. 1999). The SSU 18S rDNA gene is also the most slowly evolving of the rDNA genes thus it is suitable for addressing questions concerning ancient evolution (Hillis and Dixon 1991). However, a drawback of being the most evolutionarily conserved region of the ribosomal genes is sometimes a concern in that closely related species do not often show significant differences (Hervio et al. 1997). This is a concern when looking for variation within closely related species. The lack of significant variation in the 18S rDNA however, has led researchers to explore other molecular regions within the genome of the

Myxozoa. Such regions include the internal transcribed spacer regions (ITS1 and ITS2). The ITS regions are under fewer constraints of selective pressure and have shown significant variability which is useful in discriminating both between closely related species and among populations of a chosen species (Andree et al. 1999 b). Generally, highly conserved genes, including the functional ribosomal RNA genes as well as enzymes or structural proteins, are useful for defining ancestral relationships within eukaryotic lineages, whereas rapidly evolving sequences are best suited for more recent phylogenetic groupings (Barta 2001). Further, molecular analysis of the SSU 18S rDNA has been attempted using a series of restriction enzymes to digest the nucleic acid into readily distinguishable profiles. This has proved a quick and easy method to identify species of *Myxobolus* that are otherwise difficult to distinguish by morphology and tissue tropism alone (Eszterbauer et al. 2001). Information acquired through the use of molecular methods provides an independent data set separate from morphological information. Phylogenetic trees generated using molecular data are always open to interpretation; however, phylogenies have shed some light on the branching patterns of descent within many phyla (Collins and Valentine 2001).

The present thesis attempts to address important areas of concern that confront taxonomy of the myxozoans based solely on spore morphology. Spore taxonomy works well when the species being studied have spores with distinct morphological characteristics. However, the system breaks down when species with very similar spores are being compared. My work assesses the reliability of

the taxonomy in these situations through the application of modern molecular tools.

The first section of my thesis involves the taxonomy of *Myxobolus procerus* (Kudo 1934), a common parasite of trout-perch (*Percopsis omiscomaycus*) in the Great Lakes region of North America (Kudo 1934; Cone et al. 1997). *Myxobolus procerus* is unusual in that it forms two morphologically distinct cyst types within its host, with each form containing what is reported to be similarly shaped and sized spores (Cone et al. 1997). One cyst is large, oval and intercellular between muscle fibres. The other is small, cigar shaped and intracellular within muscle cells. The questions addressed are 1) is this two sibling species that produce different cyst types but similar spores? or 2) is it a single species with variable spore morphology depending on the cyst type the spore is in?

The second area investigated involves the taxonomy of *Myxobolus neurophilus* (Guilford 1963), a common parasite of the brain of yellow perch (*Perca flavescens*) from central and eastern North America (Dzulinsky et al. 1994). In this case the type host was yellow perch; however, Guilford (1963) reported the same species within the brain of etheostomid darters as well. The question addressed is: based on morphological and molecular data are these the same or different species of parasite? Spores of both *M. procerus* and *M. neurophilus* are themselves very similar in morphology being typically oval in shape. In this thesis I used molecular and morphological data to determine the relationship between the species noted above and other species of *Myxobolus*

for which there are sequence data available in GenBank (National Center for Biotechnology Information).

2.0 Materials And Methods

2.1 Collection of Parasites

Trout-perch (*Percopsis omiscomaycus*) (Fig. 3) heavily infected with *Myxobolus procerus* were collected by seine net fishing in the shallows of the St. Lawrence River at Île Dorval (45° 26'N, 72° 44'W) on June 3, 2002 by Dr. David Marcogliese, a fellow collaborator. Fish were transported live to a laboratory at the St. Lawrence Center, Environment Canada, Montreal. Yellow perch (*Perca flavescens*) (Fig. 4) infected with *Myxobolus neurophilus* were collected May 2003 by trap net fishing in Vinegar Lake, Nova Scotia (44° 40' N, 64° 02' W). In both cases, fish were fixed either in 10% formalin (for light microscopy) or 95% ethanol (for molecular studies). Samples of Johnny darter (*Etheostoma nigrum*) (Fig. 5) were collected May 2003 from Mykiss Lake, Algonquin Park, Ontario (45° 40' 05 N, 78° 10' 20 W) and taken to the Harkness Fisheries Research Station on Lake Opeongo. There the fish were necropsied, the spores were photographed live, and the samples were preserved in 95% ethanol.

2.2 Light Microscopy

Formalin-fixed fish were examined for cysts containing mature spores. Cysts were excised and transferred to microscope slides where they were ruptured in 2-3 drops of water. A solution of 1% molten agar on a cover slip was placed over cyst contents, and the resulting preparation used to examine

immobilized spores. Digital images of spores of *M. procerus* were obtained using a Zeiss photomicroscope camera and Zeiss morphometric software. Length and width of the spore and of the polar capsules (Fig. 6) were determined to the nearest 0.1 μM from these images. The number of polar filament coils within the capsules were counted where possible and notes made on the presence and form of intercapsular appendices, vacuoles, and sutural ridge folds.

Intense infections of *M. procerus* in trout-perch allowed detailed comparison of spore size within cysts in individual fish, between fish, and between small and large cyst types. Spore length and width, and polar capsule length and width of 25 spores from each of 3 small intracellular and 3 large intercellular cysts from 5 individual fish were compared to assess variation between cysts and among separate hosts.

2.3 Statistical analysis

Morphological data were analysed using SYSTAT. One variable was non-transformable and therefore a Kruskal-Wallis test was completed in lieu of a one-way MANOVA. The statistical analyses are therefore distribution free tests. In order to reduce the chance of a Type 1 error a Bonferroni test was completed.

Note that terms used in this thesis are defined in the glossary in Appendix B.

2.4 Molecular Biology

2.4.1 DNA Extraction

Spores were lysed in 10 mM Tris-Cl pH 8.0, 1 mM EDTA (TE), 1% SDS with proteinase K (200 µg/ml) for 2 hours in a 37°C water bath. Suspensions were extracted twice with phenol: chloroform: isoamyl alcohol (25:24:1) and the DNA was precipitated with cold 100% ethanol and 3 M sodium acetate (pH 7.0) followed by centrifugation at 9,300 g for 10 minutes. The pellet was washed once with 70% ethanol and air dried at room temperature. Genomic DNA was resuspended in 50 µl 10 mM Tris-Cl pH 8.0, 1 mM EDTA (TE) and stored in TE at 4°C. Quantification of DNA was completed using a Beckman Spectrophotometer. An aliquot of each extraction was visualized on a 1% agarose gel against known standards (Fig. 7).

2.4.2 Polymerase Chain Reaction

Small subunit (SSU) 18S rDNA was amplified using myxozoan general and specific primers (Table 2). Primers were re-suspended in ddH₂O to a stock concentration of 10 µM. PCR was performed in 50 µl reaction volumes containing 1.5 mM MgCl₂, 0.2 mM dNTP, 1.25 Units *Taq* polymerase, 1 µM of each primer and 300 ng DNA. Amplifications were performed on a Perkin-Elmer Gene Amp 9700 Thermocycler. Cycling parameters were as follows: initial denaturation at 95° C for 5 minutes followed by 35 cycles of 94° C for 1 minute, 55° C for 1 minute, 72° C for 1.5 minutes and a final extension at 72° C for 10 minutes. The

PCR products were excised from the agarose gel and purified. A list of reagents used in this work is given in Appendix C.

2.4.3 Cloning

The purified PCR products were analysed on a 1% agarose gel and 4 μ l was used for cloning reactions. PCR products were cloned using the TOPO TA Cloning Kit (see Appendix F). Prior to cloning 4 μ l of the “cleaned” PCR product was incubated at 72°C for 15 minutes in a sterile PCR tube containing 1 μ l dATP (at 1 μ M), 0.25 U *Taq* and 0.5 μ l 10X *Taq* buffer. This was to ensure the addition of the poly A tail necessary for a successful TOPO reaction. Chemically competent *Escherichia coli* cells were transformed with the TOPO vector containing the PCR generated inserts. Approximately 150 μ l of each transformation was streaked onto LB plates containing carbenicillin to a final concentration of 25 μ g/ml + X-gal and grown overnight at 37°C/300 RPM. A representative number of “white” colonies indicating positive clones were transferred into 1 ml LB broth + carbenicillin (25 μ g/ml) and grown at 37°C /300 rpm overnight. Glycerol stocks of each colony were prepared by mixing 100 μ l of culture with 100 μ l of a sterile 50% glycerol solution. The stock cultures were then stored at –80°C.

2.4.4 Screening

2.4.4.a Protoplasting

Protoplasts were generated using a modified chemical treatment assay.

Approximately 3 ml of LB containing carbenicillin (25 μ g /ml) were inoculated with a single colony. The tubes were placed at 37°C/300 rpm for 18 hours. After incubation 50 μ l of culture was transferred into a sterile 0.6 ml Eppendorf tube and centrifuged at 9,300g for 10 minutes. The supernatant was discarded and the pellet re-suspended in 30 μ l sucrose buffer. A further 10 μ l of phenol:chloroform:isoamyl alcohol (25:24:1) was added and the tube was vortexed for 1 minute at high speed. The tubes were then centrifuged for 20 minutes at 9,300g.

2.4.4.b Plasmid Preparation

Plasmid DNA was isolated from clones using the Qia-Prep Mini prep isolation procedure or a simple protoplasting technique and transformants were analysed on a 1% agarose gel (Fig. 8). Sizes of the inserts were determined by comparison against a standard of known molecular weight.

2.4.4.c Screening by M13 PCR

To determine the presence and sizes of inserts purified plasmid DNA was amplified using primers M13 F and M13 R (Fig. 9). Reaction conditions are as follows;

PCR Reaction	Cycling conditions	
1 X PCR buffer	1. 95°C	5'
dNTP 0.2 mM	2. 95°C	1'
<i>Taq</i> polymerase 1.25 U	3. 55°C	1'
M13F 100 ng	4. 72°C	1'
M13R 100 ng	5. Go to 2 for 30 X	
ddH ₂ O to 20 μ l	6. 4°C α	
DNA to 300ng		

2.4.5 DNA Sequencing

Sequencing reactions were performed using those primers that generated PCR products. Sequence data was also obtained using M13 F and M13 R primers. The following clones were sequenced: three clones each of *M. procerus* spores from intercellular cysts with 18e and MX3 primers, three clones of *M. procerus* spores from intracellular cysts using MX3 primer and two clones using Myxgen2r (18r) primer, and three clones from yellow perch using M13 F and M13 R primers with cloned 18e-Myxgen2r (18r) PCR product, three clones each from Johnny darter using M13F and M13 R primers with cloned 18e-Myxgen2r PCR product and cloned Act3f-MX3 PCR product. In order to obtain more SSU 18S rDNA from *M. procerus*, “walking” primers were designed from those regions containing unambiguous sequence data. Sequencing reactions were performed using ET terminator chemistry and sequencing was done on a MegaBACE 1000 capillary sequencer.

Sequences were aligned against the vector to eliminate any contaminating vector DNA. To further eliminate the presence of any vector sequence a restriction map of the SSU 18S rDNA isolated in this work was constructed. Recognition sites common to the TOPO plasmid were not identified, thereby further confirming the lack of vector contamination.

2.4.6 Sequence and Phylogenetic Analysis

Sequencher (Version 4.0.5) was used to edit sequence data and produce contiguous alignments. Sequences were subjected to BLAST (Altschul et al.

1990) searches against sequences in GenBank (NCBI). Bit scores are listed in the tables along with the corresponding E values. The Bit score indicates the relatedness of the sequence to those against which it aligns. It is also a link to the NCBI database from which the sequence originates. The E value illustrates the number of matches to the current non-redundant database that are expected by chance alone. The lower the E value the more “real” the alignment in question. The higher the bit score the greater the similarity.

Alignment of SSU 18S rDNA sequences is completed using DNA analysis software, termed Clustal X (Thompson et al. 1997) with optimal parameters defined in Hall (2001). Clustal X uses an algorithm that aligns most closely related sequences first then these groups are gradually aligned together retaining the previous alignments (Thompson et al. 1997). The output of the Clustal X alignment is read by the phylogenetic tree-generating program PAUP (Swofford et al. 2001). Algorithms used in the PAUP analysis included parsimony, maximum likelihood, and distance methods and bootstrap analysis with 1000 replicates. Sequences used were; *Ceratomyxa shasta* GenBank AF001579, *Myxidium truttae* GenBank AF201374, *Myxidium* spp. GenBank U13829, *Myxidium lieberkuehni* GenBank X766390, *Henneguya lesteri* GenBank AF306794, *Henneguya* spp. GenBank U13826, *Henneguya doori* GenBank U37549, *Henneguya ictaluri* GenBank AF195510, *Henneguya exilis* GenBank AF021881, *Myxobolus osburni* GenBank AF378338, *Myxobolus spinacurvatura* GenBank F378341, *Myxobolus ichkeulensis* GenBank AF378337, *Myxobolus lentisuturalis* GenBank AY119688, *Myxobolus* KAB-2001B GenBank AF378343,

Myxobolus xiaoi GenBank AF186843, *Myxobolus portucalensis* GenBank F085182, *Myxobolus cerebralis* GenBank U96492, *Myxobolus insidiosus* GenBank U96494, *Myxobolus neurobius* GenBank AF085180, *Myxobolus elipsoides* GenBank AF085178, *Myxobolus djragini* GenBank AF085179, *Myxobolus arcticus* GenBank AF085176, *Myxobolus bramae* GenBank AF085177, *Myxobolus sandrae* GenBank AF085181, *Myxobolus* spp. KAB2001A GenBank AF378342, *Henneguya zschokkei* GenBank AF378344, *Henneguya salminicola* GenBank AF031411, *Myxobolus pseudodispar* GenBank AF380143, *Myxobolus musculi* GenBank AF380141, *Myxobolus cyprini* GenBank AF380140, *Myxobolus hungaricus* GenBank AF448444, *Myxobolus pendula* GenBank AF378340, *Myxobolus pellicides* GenBank AF378339, *Myxobolus bibullatus* GenBank AF378336, *Myxobolus elegans* GenBank AF448445, *Myxobolus algonquinensis* GenBank AF378335.

3. Situation 1 – Cyst Types of *Myxobolus procerus*.

3.1 Light Microscopy

Microscopic examination of trout-perch during necropsy revealed the presence of both intercellular and intracellular cysts. Cysts exist intercellularly within connective tissue associated with muscle as well as within the outer epidermis between dermal scales. The intercellular cysts were white, oval and approximately 1.5-2 mm in size and concentrated around the head and caudal peduncle regions (Fig. 10). Intracellular cysts inside striated muscle cells were dark grey and cigar shaped structures (up to 1mm long) running the length of the muscle cell (Fig. 11). Intracellular cysts were less prone to rupture and could be handled with greater pressure than the intercellular cysts that were more fragile and had to be handled more delicately. Intercellular and intracellular cysts were treated separately so as to avoid crossover contamination.

Spores of both cyst types were typically pyriform in shape, tapering anteriorly to a knob-like end. The polar capsules were a narrow pyriform shape, with a pore-like intercapsular appendix (Fig. 12). The sutural ridge was thin, with some spores having 3-4 posteriorly located sutural ridge folds.

Table 4 summarizes the statistical comparison of morphometrics obtained from intercellular and intracellular cysts. The histogram in Figure 13 provides an illustration of the distribution of spore length in both cyst types. There were no statistically significant differences in spore morphometrics, and the number of polar filament coils in the spores, from intracellular cysts within individual fish or between the five fish sampled. Similarly, the same applied to spores from

intercellular cysts. However, spores from intercellular cysts were significantly longer than spores from intracellular cysts.

3.2 Molecular analysis

Polymerase Chain Reactions using myxozoan general primers produced products of approximately 1.6 Kb (Fig. 14). Primer set 18e-Myxgen2r (18r) was more consistent and therefore PCR products from this primer set were used for further molecular analysis. Subsequent primer sets were used to “walk” the 18S region of *Myxobolus procerus* isolated from both cyst types (Table 2). PCR products were used in TOPO cloning reactions and white colonies indicating possible transformants were screened using the methods discussed above. Positive transformants were sequenced and those clones with “reliable” sequence data were aligned and analysed. Analysing individual clones for sequence heterogeneity provided an indication of variability within different cyst types. Individual clones from each reaction were analysed and regions with poor sequence data removed to provide a reliable region to identify sequence differences. Table 3 shows the number of base pairs of each clone that was compared. As indicated, there were no differences in SSU 18S rDNA sequence between those clones of *Myxobolus procerus* analysed.

3.3 Basic Local Alignment Search (BLAST) Results

Comparison of the consensus sequence (Figs. 15 and 16) for intracellular and intercellular forms of *M. procerus* with myxozoan sequences submitted to GenBank showed that they are indeed myxobolids with a high degree of similarity to *Myxobolus exiguus* (93%) Accession # AF021878.1, *M. muelleri* (93%) Accession # AY129317.1, *Aurantiactinomyxon mississippiensis* (90%) Accession # AF021878.1, *Henneguya exilis* (89%) Accession # AF021881.1 and *H. ictaluri* (89%) Accession # AF195510.1 (Table 5). The remaining 8 sequences producing significant alignments with the consensus sequences of *M. procerus* from both intercellular and intracellular cysts types are listed in Figures 17 and 18.

3.4 Clustal X Alignment of SSU 18S rDNA

A total of 989 base pairs from consensus sequences for both intercellular and intracellular cysts types were compared using Clustal X software (Fig. 19). There were 20 base pairs difference (2.1% of total) in SSU18S rDNA sequence between *M. procerus* within the intracellular cysts and *M. procerus* within intercellular cysts. Of these 20 base pair differences 10 were due to base substitutions and the remaining 10 due to the presence or absence of a nucleotide. Differences in sequence could be attributed to *Taq* polymerase error; however, according to the manufacturer the enzyme is responsible for very little error. Error rates for thermal resistant DNA polymerases have been calculated at

1.1 X 10⁻⁵ base substitutions/bp (Tindall and Kunkel 1988). This indicates only one error in every 110 Kilobases, a value that is in question in the current work.

4.0 Situation 2 – *Myxobolus neurophilus*

4.1 Light Microscopy

Formalin fixed spores of *M. neurophilus* from the optic tectum of yellow perch (*Perca flavescens*) from Vinegar Lake, Nova Scotia, were pyriform in shape with a bluntly pointed anterior extremity and broadly rounded posterior (Fig. 20). Spores measured $14.40\ \mu\text{m} \pm 0.77\ \mu\text{m}$ long, $6.96\ \mu\text{m} \pm 0.24$ wide (N=13), with two similarly sized polar capsules $6.50\ \mu\text{m} \pm 0.24$ long and $2.10\ \mu\text{m} \pm 0.21$ wide containing 7 – 9 filament coils arranged perpendicular to the capsule length. There was no evidence of an intercapsular appendix, sutural ridge folds, or a mucous coat.

Material collected from the optic tectum of Johnny darter (*Etheostoma nigrum*) from Mykiss Lake, Algonquin Park was tentatively identified by Guilford (1963) as *Myxobolus neurophilus* on the basis of host, developmental site, and morphology of the spores examined and photographed live in the field. Spores were pyriform but with a subtle narrowing at the anterior extremity (Fig. 21). Fresh spores measured $14.40\ \mu\text{m} \pm 0.72$ long and $7.40\ \mu\text{m} \pm 1.20$ wide, with two similarly sized narrowly pyriform polar capsules $7.50\ \mu\text{m} \pm 0.40$ long and $2\ \mu\text{m}$ wide. Some spores had what appeared to be a round intercapsular appendix anteriorly as well as 3 – 4 posteriorly located sutural ridge folds. A large

iodinophilous vacuole was present in the sporoplasm. There was no evidence of a mucous coat.

4.2 Molecular Analysis

4.2.1 *Myxobolus neurophilus* from yellow perch (*Perca flavescens*)

Using primer sets Myxgp2f-Act1R and Act3F-MX3 both specific for the Myxozoa, I was able to amplify products of approximately 920bp and 750bp respectively from the neural tissue of yellow perch. Three clones were generated from the 18e-Myxgen2r PCR product only. Each clone was successfully sequenced using M13 forward and reverse primers. After removing unreliable sequence data a total of 723 bp from each of the three clones were aligned and differences between the clones noted (Fig. 22). There were no differences in the consensus sequence from each clone. The resulting 723 bp sequence used for the subsequent BLAST search is illustrated in Figure 23.

4.2.2 *Myxobolus neurophilus* from the Johnny darter (*Etheostoma nigrum*)

Using the same primers as noted above PCR products were generated from the neural tissue of Johnny darter. Products from both primer sets were successfully cloned and sequences from each of three clones from each primer set were compared. Lining up clones 1-3 from the Myxgp2f-Act1r primer set gives 844 bp of sequence with no differences. Lining up clones 1-3 for the Act3F-MX3 primer set gives 750 bp sequence data with two differences in clone 3 sequenced in both forward and reverse directions. Differences were noted at

position 429 with C for a T (questionable) and position 575 with G for an A (uncontested). Lining up all clones gives me 1570 bp with the two differences from clone 3 AM (Fig. 24). The resulting consensus sequence Johnny darter (Fig. 25) was used in a Clustal X alignment against the consensus sequence from yellow perch and trout-perch.

4.2.3 Basic Local Alignment Search (BLAST) Results

BLAST results for each consensus sequence from Johnny darter and yellow perch are illustrated in Figures 26 and 27. Sequences obtained from PCR reactions with DNA isolated from yellow perch were most similar to *Henneguya zschokkei* (Accession # AF378344), *Henneguya weishanensis* (Accession # AY165182), *Henneguya ictaluri* (Accession # AF195510) and *Neoactinomyxum* sp. (Accession # AF378353). Sequences obtained from Johnny darter BLAST'ed with a high degree of similarity against *Henneguya weishanensis* (Accession # AY165182), *Henneguya ictaluri* (Accession # AF195510) and *Neoactinomyxum* sp. (Accession # AF378353) and *Tetraspora discoidea* (Accession # AF306793).

4.2.4 Clustal X Alignment of SSU 18S rDNA

Sequences generated by amplification of the SSU 18S rDNA from all of the species of *Myxobolus* studied were aligned using Clustal X. Ambiguous regions were removed and sequences aligned using Clustal X. The alignment between *Myxobolus* species isolated from Johnny darter and the yellow perch resulted in an 8% difference noted (54/690) (Fig. 28). The differences noted were confirmed using the accompanying chromatogram from data generated by the MegaBace

sequencer. The results suggest that the species of *Myxobolus* isolated from Johnny darter is a different species from *M. neurophilus* isolated from yellow perch. The consensus sequence from Johnny darter was also compared to the consensus sequences generated from the SSU 18S rDNA from spores within each cyst type of *M. procerus*. A total of 989 base pairs of the SSU 18S rDNA from both the darter and trout-perch were aligned with a 53 base pair difference noted (or 5.3 % sequence difference) between darter and *M. procerus* intracellular cysts and a 56 base pair difference (5.6%) between darter and *M. procerus* intercellular cyst types (Fig. 29). What is interesting is that the sequence variation between the myxozoans from the darter and the trout-perch is less than that between the darter and the yellow perch. The consensus sequence from the yellow perch was aligned against consensus sequences from the trout-perch and the differences were significant at > 25% sequence difference (Fig. 30) and clearly indicates a separate species of *Myxobolus*.

5.0 Phylogenetic tree of species of *Myxobolus* identified in this thesis

In my thesis I have provided a molecular comparison between myxozoan SSU 18S rDNA isolated from yellow perch, trout-perch and Johnny darter. Using these three species their position within the phylogeny of the Myxozoa was determined. The phylogenetic tree confirms the presence of three separate species that have been identified in the present work (Fig. 31).

Using three algorithms of analysis, neighbour-joining, parsimony and maximum likelihood the topology of the trees remained unchanged (See

Appendix E and F). The phylogenetic tree places *Myxobolus* sp. from Johnny darter within a clade that includes *Henneguya lesteri* with strong bootstrap support. *Myxobolus procerus* isolates from intercellular and intracellular cyst types are also closely aligned with *Myxobolus* sp. from Johnny darter, once again with high bootstrap support. However, sequence data from *M. procerus* of trout-perch and *Myxobolus* sp. of Johnny darter remain significantly different suggesting that they are in fact separate species. *Myxobolus neurophilus* from yellow perch occupies an independent position from those species sequenced to date. This sequence is more closely related to *Myxobolus* species KAB 2001A and *Myxidium* species. The position of isolates from Johnny darter and yellow perch in the phylogenetic tree suggests that they are in fact different species of *Myxobolus*.

Further information on *Myxobolus* species used in the phylogenetic tree is given in Appendix G. The Clustal X alignment of all the species of *Myxobolus* used in creation of the phylogenetic tree is illustrated in Appendix H.

6.0 Discussion

Of the recent advances in biological research few have created a greater impact than those involving molecular biology. This “so-called” cutting edge technology has created, arguably, a “new” taxonomy based upon information acquired through analysis of the genotype of an organism. The meeting of morphological and molecular data has not come without controversy. On one side are the traditional taxonomists who feel that studies must remain focused on

morphology to address issues of diversity, especially in areas in which molecular analysis comes with too great a financial commitment. Indeed funding for molecular based projects is a concern leading researchers to speculate as to where this funding will come from (Mallet and Willmott 2003). Seberg et al. (2003) asks the question “what has the molecular revolution really achieved for taxonomy?” A significant quantity of data has become available for phylogenetic analyses however the practice of using a single specimen as representative of the taxon can create havoc in taxonomy (Seberg et al. 2003). Opponents however, concede that the information to be gained is a welcome and at times essential component to the construction of real taxonomic schemes. A final word on this side of the controversy; questions have been raised concerning the trend towards reducing taxonomy of all organism to this impoverished state by replacing existing methods with a system that uses only a tiny fraction of an organisms genome to classify and identify the organism in question (Lipscomb et al. 2003). On the alternate side there are those more adamant who feel that it is not only advantageous but also compulsory to add molecular information to current taxonomy. Proponents of molecular taxonomy are convinced that all hopes for a reliable classification lie in the incorporation of molecular data to in effect generate a taxon “barcode ” (Hebert et al. 2002). The present work is very much a participant in this debate for taxonomic questions are addressed that are explained using both techniques of traditional taxonomy and molecular biology.

The first situation addresses the significance of two cyst types of *M. procerus* in trout-perch. In the original species description of this parasite, Kudo

(1934) described only large intercellular cysts from trout-perch in Illinois. Cone et al. (1997) described what was considered to be a variant of *M. procerus* from Lake Superior involving the majority of the cysts being intracellular in muscle cells with relatively few large intercellular cysts being produced. The present study of host tissue from the St. Lawrence River, Quebec, similarly found both cyst morphotypes co-existing within the single host species. The fact that the two cyst types do not exist independently of each other further suggests polymorphism in cyst development for *M. procerus*.

In contrast to what was reported by Cone et al. (1997) the present morphological comparison of spores from the two cyst types observed that spores from the small, intracellular cysts were shorter than those from the large, intercellular cysts. It appears that site of development has an effect on the morphology of the spores being produced. How does this come about? The life cycle of *M. procerus* is unknown. However, it is assumed that fish acquire infections from invasive actinosporeans released from oligochaetes (Kent et al. 2001). The migrating infective sporoplasm enters individual striated muscle cells and develops or lodges between cells and develops, forming the two cyst types. Implications are that there may be a physical limitation on spore size as a direct effect of the environment within which the spores are located. As might be expected spores contained within the smaller intracellular cysts are indeed shorter than those within the larger intercellular cysts. What the present work suggests is that during development there may be certain physical or chemical limitations in the way of nutrients available to the developing spore that would

affect growth. Previous work has suggested that it is possible that spore dimensions could vary due to developmental temperature, site of infection, host fish parameters and spore age (Yokoyama et al. 1997). Morphological examinations of spores of *Myxobolus koi* in the gills of common carp identified large and small cysts correlated with several distinct differences in their seasonality and mode of development (Yokoyama et al. 1997). It is tempting to suggest that the material studied from the two cyst types of *M. procerus* reflects the process of speciation.

The molecular data acquired in the present work supports the conclusion of ongoing ecophenotypic variation in spores of *M. procerus* depending on cyst type. The sequence data suggests a high degree of homology within the region of the genome that was sequenced. As indicated there is only a 2.1% difference in the SSU 18S rDNA between spores contained within each cyst type. The conclusion is that this is a single species with phenotypic variation. The first reported example of two morphologically distinct spore types that were indistinguishable genetically was described for the aurantiactinomyxons by Hallett et al. (2002). The authors examined a small region of the parasite genome near the 5' end and contend that only a 358-bp region was sufficient to provide information on the variability inherent in different species (Hallett et al. 2002.) Their findings indicate a shared homology between morphological distinct aurantiactinomyxons of only 42-44% of their bases (Hallett et al. 2002).

The second hypothesis examined in the present thesis involves a presumed single species of *Myxobolus* occupying the same specific tissue but in

separate hosts. Guilford (1967) described *M. neurophilus* in the optic tectum of its parent host the yellow perch. This description is now considered inadequate by present standards because of the lack of accurate photomicrographs of spores in this early work. Guilford (1963) also reported the parasite from the tectum of Johnny darter (*Etheostoma nigrum*) though this was a peremptory conclusion for he failed to provide any morphological information to support the conclusion. Assuming, as Guilford (1963) did, that the species of *Myxobolus* within the neural tissue of Johnny darter was the same as that described for yellow perch, I attempted a detailed analysis of specific genetic regions of the isolates. On a morphological basis the spores of both myxobolids are very similar, however, the SSU 18S rDNA sequence information differs significantly. Using this single genetic region as a taxon criteria places doubt on classifying this particular parasite from Johnny darter as *M. neurophilus*. In fact, it implies that the material collected from Johnny darter is a new species of *Myxobolus*.

Unfortunately we did not have sufficient material for a reliable morphological comparison between spores of *M. neurophilus* from yellow perch and those isolated from Johnny darter. The available material reveals that the spores differ slightly in shape; those of *M. neurophilus* from yellow perch do not have the indent in the shell as it tapers toward the anterior extremity as seen with the presumed new species in Johnny darter. What is interesting is that in this specific feature, spores of *Myxobolus* sp. from Johnny darter are very similar to *M. procerus*. A detailed analysis of the pyriform shape of these myxobolids needs to be completed to reveal the importance of these observations. Recent

information from Dr. Mark Ridgeway in Ontario suggests that there may presently not be any yellow perch in Mykiss Lake, thus supporting the conclusion of a separate species of *Myxobolus* in Johnny darter.

DNA extractions were performed on pooled spores and therefore the possibility of variation within individual samples becomes an issue. To address this a representative number of clones generated from PCR reactions using specific and general myxozoan primers were sequenced and aligned to identify any variations that may exist. Table 3 indicates the clones of *M. procerus* that were aligned to identify sample variation. As indicated, within each cyst phenotype there were no sequence variations suggesting there was no variability within samples. Alignment of clones from those species isolated from yellow perch and Johnny darter are illustrated in Figures 68 and 72 respectively. There are no differences within the three clones of SSU 18S rDNA isolated and sequenced from the yellow perch. There was however two differences noted in clone #3 with primers Act3F-MX3 amplified from Johnny darter. Sequence data from this clone was ambiguous and not as “clean” as that obtained from clones 1 and 2 with the same primer. Thus the consensus sequence from clones 1 and 2 of Johnny darter were used for the alignment against the yellow perch sequences. Differences in clone # 3 from Johnny darter can be attributed to mispriming by the *Taq* polymerase or may be sequencing error resulting in base substitutions however infrequently they do occur. Errors due to *Taq* polymerase is statistically low and not considered a reliable source of sequence variation however this manufacturers claim is not always substantiated.

The phylogenetic tree illustrates the taxonomic placement of a series of myxobolids based upon analysis of SSU 18S rDNA. The tree in Figure 31 is very similar to and defines topologies resembling those generated by Dyková et al. (2002) using the same SSU 18S rDNA sequences that I used in my analysis.

Using the phylogenetic tree as a method of classifying the myxobolids raises many interesting questions. Phylogenetic analyses using the 18S rDNA have grouped morphologically distinct organisms into clades of similar species bringing into question the division of parasites into genera based solely on polar capsule numbers (Whipps et al. 2003). The two morphotypes of *Myxobolus procerus* spores from trout-perch are grouped identically with high bootstrap support. The isolate from Johnny darter falls in with *M. procerus* suggesting a genetic link thus supporting the morphological similarities previously noted. This clade is assigned high bootstrap support, which enhances the reliability of the grouping.

It must be noted that BLAST results do not provide the same information as the phylogenetic analysis program. The BLAST program identifies those sequences that are sufficiently similar to the sequence of interest as compared against the international database at NCBI. The phylogenetic tree program PAUP then provides an illustrated look at the relationships of each of these sequences using a chosen algorithm. This indicates the variability in sequence analysis results that are dependant upon the chosen algorithm.

Myxobolus neurophilus from yellow perch appears independently and more closely aligned to *Myxobolus* sp. in GenBank than to the isolates from

Johnny darter. In fact the yellow perch isolate is not far removed from other brain dwellers including *Myxobolus insidiosus*, *M. arcticus* and *M. neurobius*, the latter two species isolated from the neural tissue of salmon. *Myxobolus neurobius* is also found in the spinal cord of trout and salmon distributed throughout British Columbia and Newfoundland (Lom and Dyková 1992).

The species examined in this thesis all have classic pyriform shaped spores, a characteristic that traditional taxonomy would use to group them together. The molecular data however draws a different picture by examining the SSU 18S rDNA of those species studied. In a less conserved region this would not appear significant but within highly conserved genetic regions differences are significant.

The present work questions the feasibility of clarifying species based on spore morphology or molecular data alone. In my thesis I have identified what appears to be three separate species of *Myxobolus* based upon comparison of morphological and molecular data. What provides the defining criterion when molecular and morphological data disagree? Research by Whipps et al. (2003) identifies a variant of *Kudoa permulticapsula* that is significantly different in morphology than any species of *Kudoa* previously described. However based on the SSU 18S rDNA sequence the authors contend that assigning the myxozoan to a new species (or family) would be unacceptable (Whipps et al. 2003). This thesis suggests that conclusions based solely on the analysis of a single genetic region place doubt on an accurate classification of species of *Myxobolus*.

A reliable taxonomic division cannot be defined unless data from both methods are thoroughly examined. In cases when the molecular data from a single genetic region seriously contradicts morphological information it is suggested that other regions of the genome be explored. Molecular biology has provided a sensitive tool with which to examine an organism at a level separate from traditional techniques. Targeting either conserved or variable genes allow examinations into the similarities between species at precise levels of molecular differentiation. However molecular techniques like traditional methods are simply tools to be used in taxonomy. If the data is sound both morphological and molecular data should be used synergistically and not competitively.

Certainly more information will become available as new techniques are discovered and existing techniques improved however the present state of taxonomy can only be enriched by inclusion of the methods discussed in the present work.

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8. FIGURES

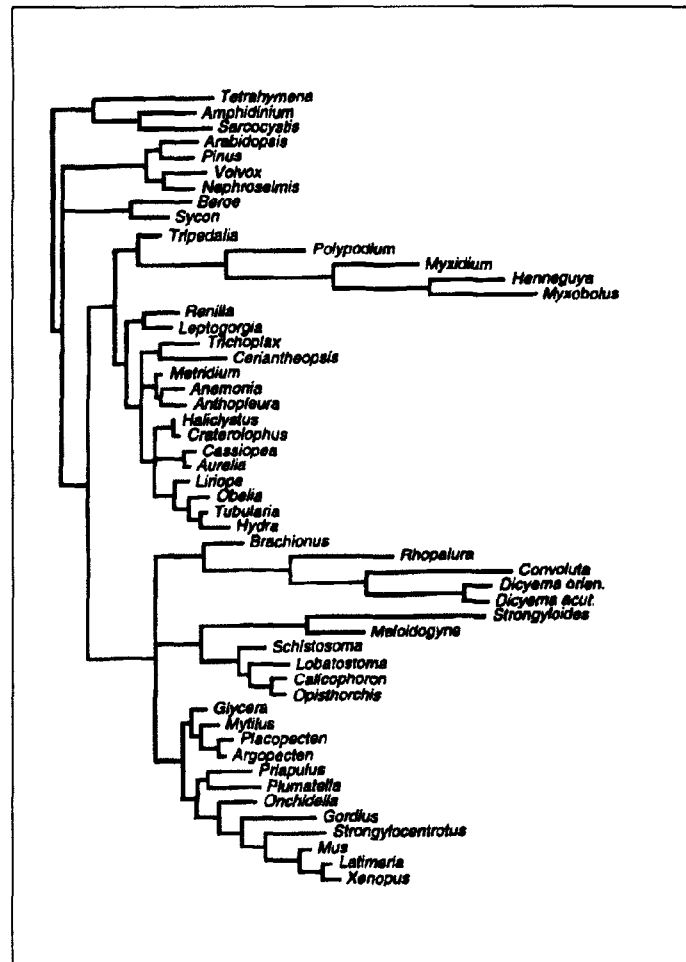


Figure 1. Parsimony analysis of a taxonomically more inclusive molecular data set results in support for the monophyly of the myxozoans with *Polypodium hydriforme* as well as for the phylum Mesozoa (from Siddall and Whiting, 1999).

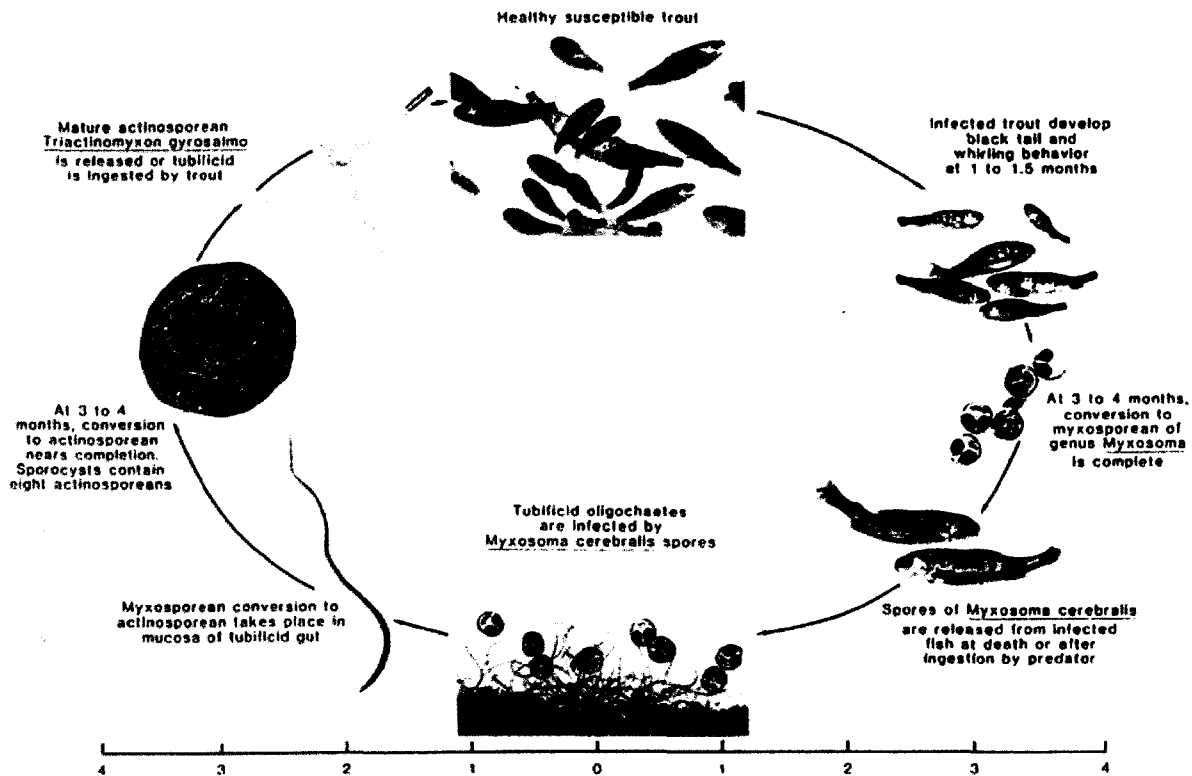


Figure 2. Biphasic life cycle of a myxozoan. From Wolf and Markiw, Science, 1984.



Figure 3. Trout-perch (*Percopsis omiscomaycus*)



Figure 4. Yellow perch (*Perca flavescens*)



Figure 5. Johnny darter (*Etheostoma nigrum*)

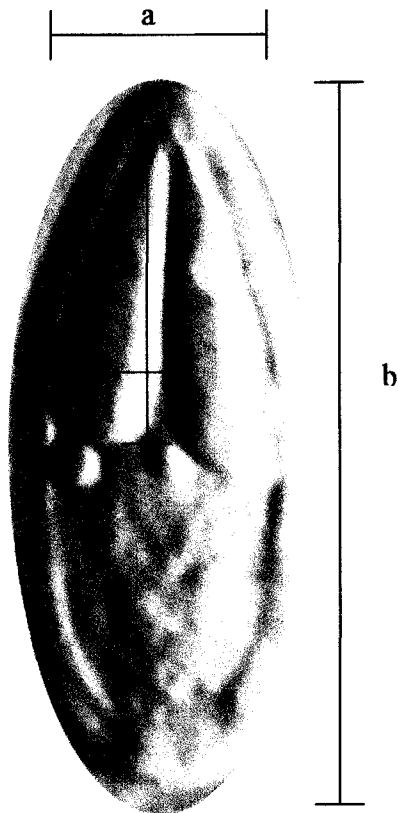


Illustration of spore measurements. a and b indicate width and length respectively. Width of polar capsules inside spores were measured in the same way.

Figure 6. Spore measurement schematic



Figure 7. DNA extraction of species of *Myxobolus* using a traditional phenol:chloroform:isoamyl alcohol method. Lanes at right contain pGEM DNA at known concentrations of 10 – 100 nanograms DNA. High molecular weight DNA is evident at the top of the gel with some degradation throughout the lanes. Marker is supercoiled DNA ladder.



Figure 8 a and b. Screening of transformants using Qiagen mini prep kit on left and simple protoplasting method on right. High molecular weight DNA is indicated by comparison with Mass ladder. Inserts are calculated by size of PCR product + Vector. Example: 1.6 kb PCR product + 3.9 kb vector = 5.5 Kb fragment.

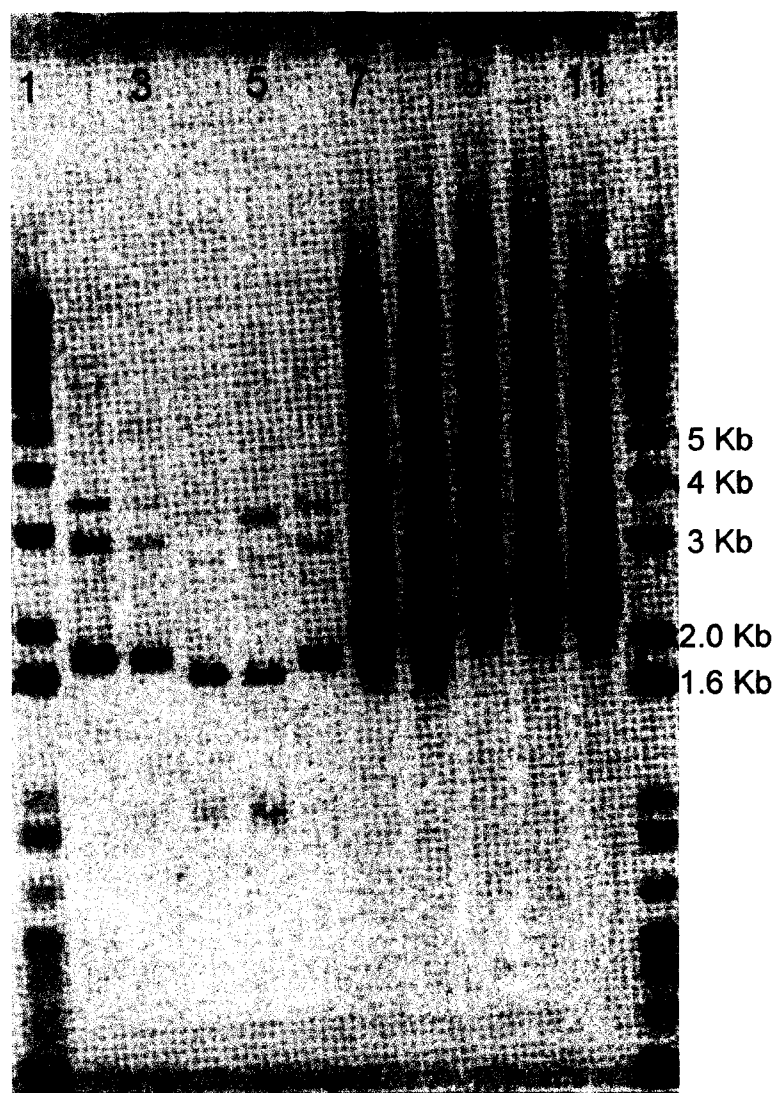


Figure 9. M13 PCR for screening of transformants. Lanes 1 and 12 contain 1 kb+ ladder. The primers amplify regions between M13 forward and reverse and presence of insert is determined by the size of the PCR product + intervening primer sequence. Positive products at between 1.6kb and 2kb are indicated.

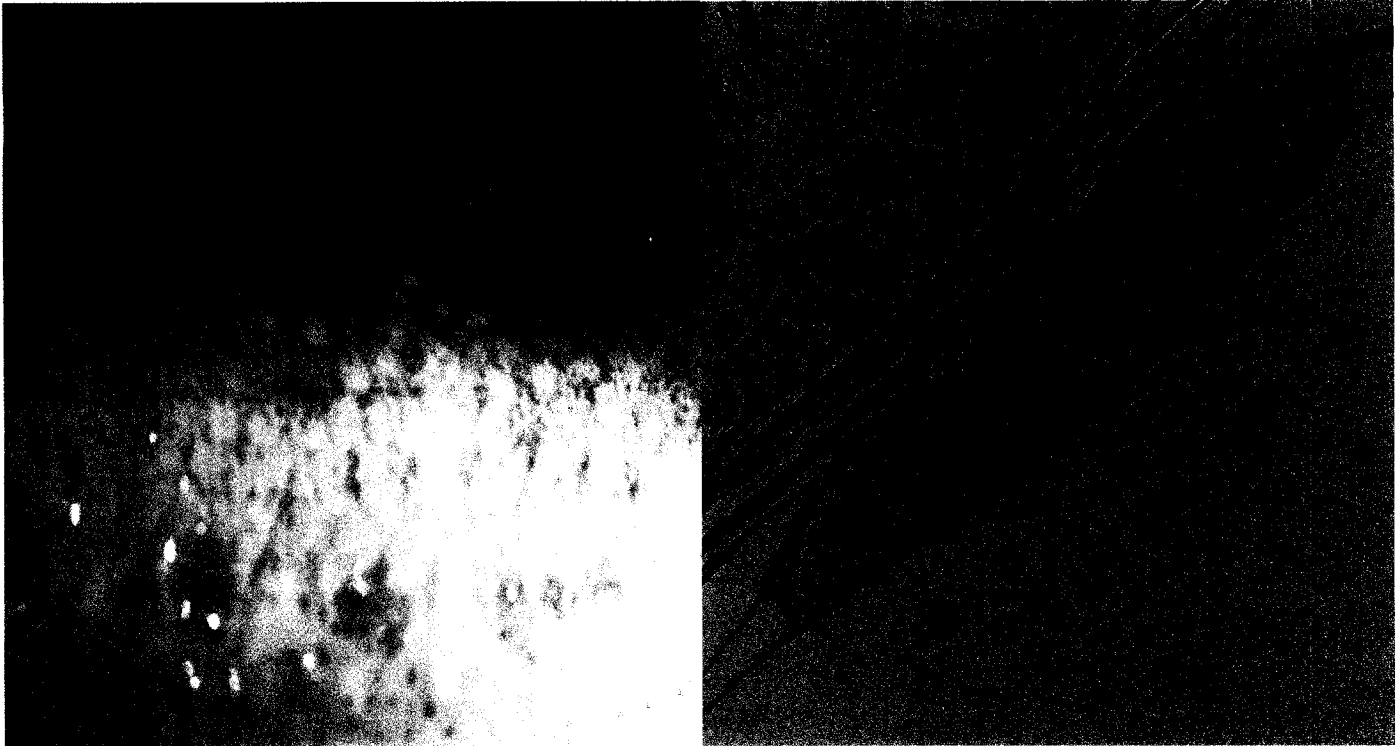


Figure 10. Intercellular cysts of *Myxobolus procerus*. The large cysts containing the parasitic spores in the left illustration are photographed on the surface of the trout-perch alongside the dorsal fin. The picture on the right illustrates a cross-sectional view of the intercellular cysts lying just beneath the epithelium of the host.

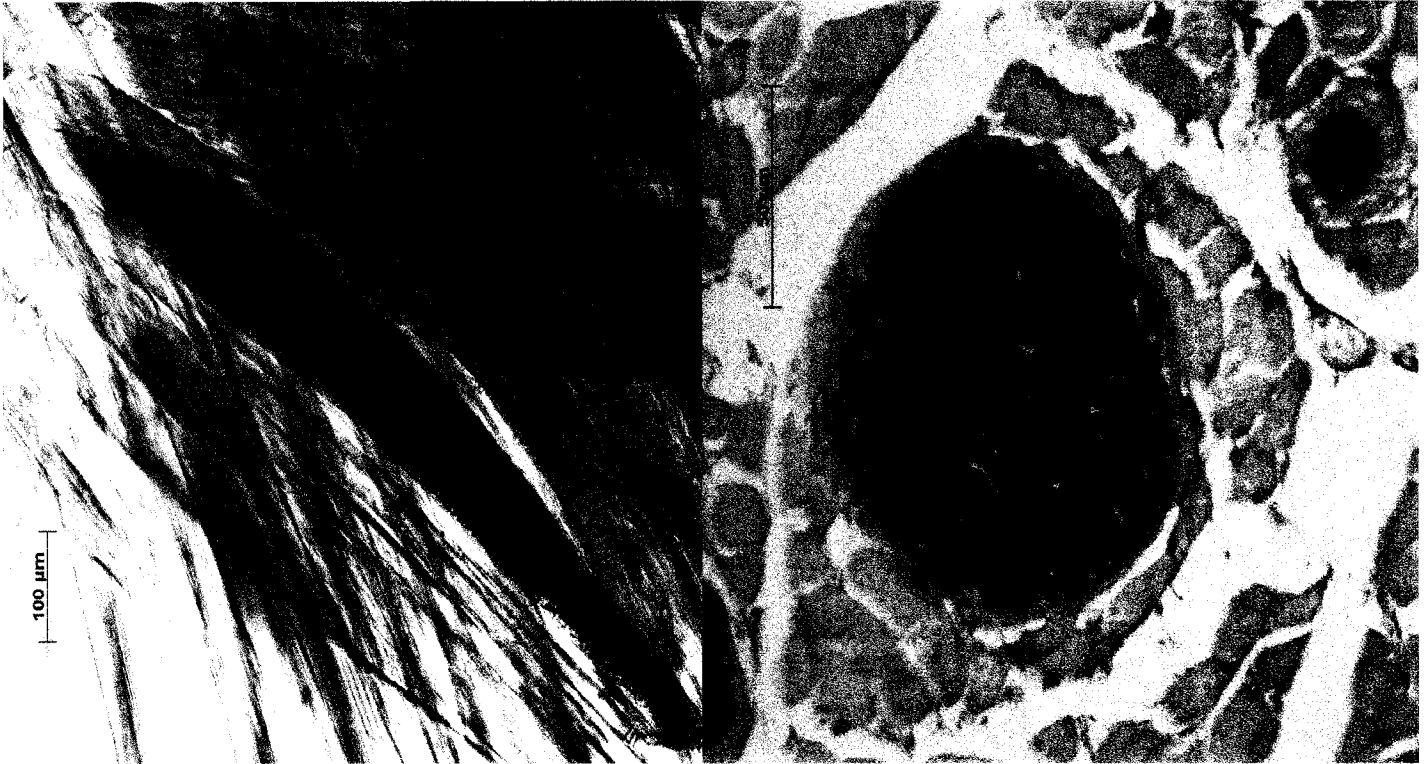


Figure 11. Intracellular cysts of *Myxobolus procerus*. The cyst lies inside of the cell of its host and is clearly fusiform in shape. The picture on the right is a cross sectional view of the developing cyst. Spores are visible as dark oval bodies inside of the cyst.

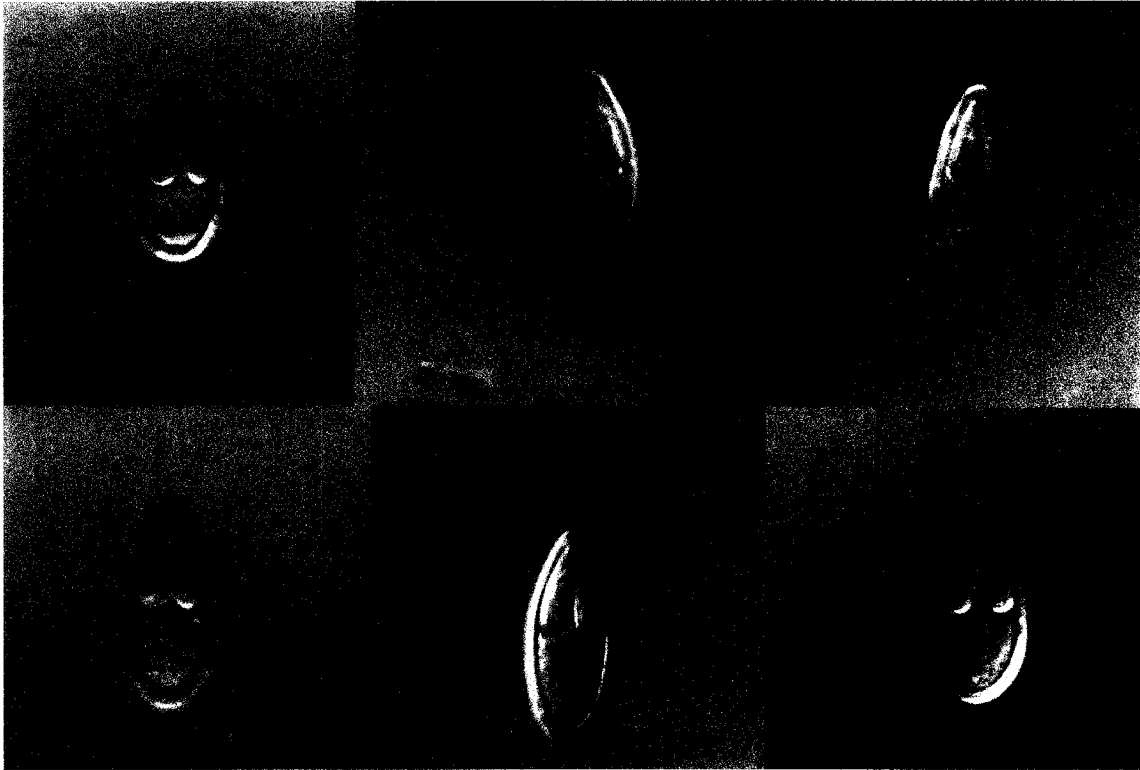


Figure 12. Spores of *Myxobolus procerus*. The top three spores are from the small intracellular cyst. The bottom three from the large intercellular cysts. Note the elongated spore shapes of those from the larger cysts. Scale bar at left is 10 μ m.

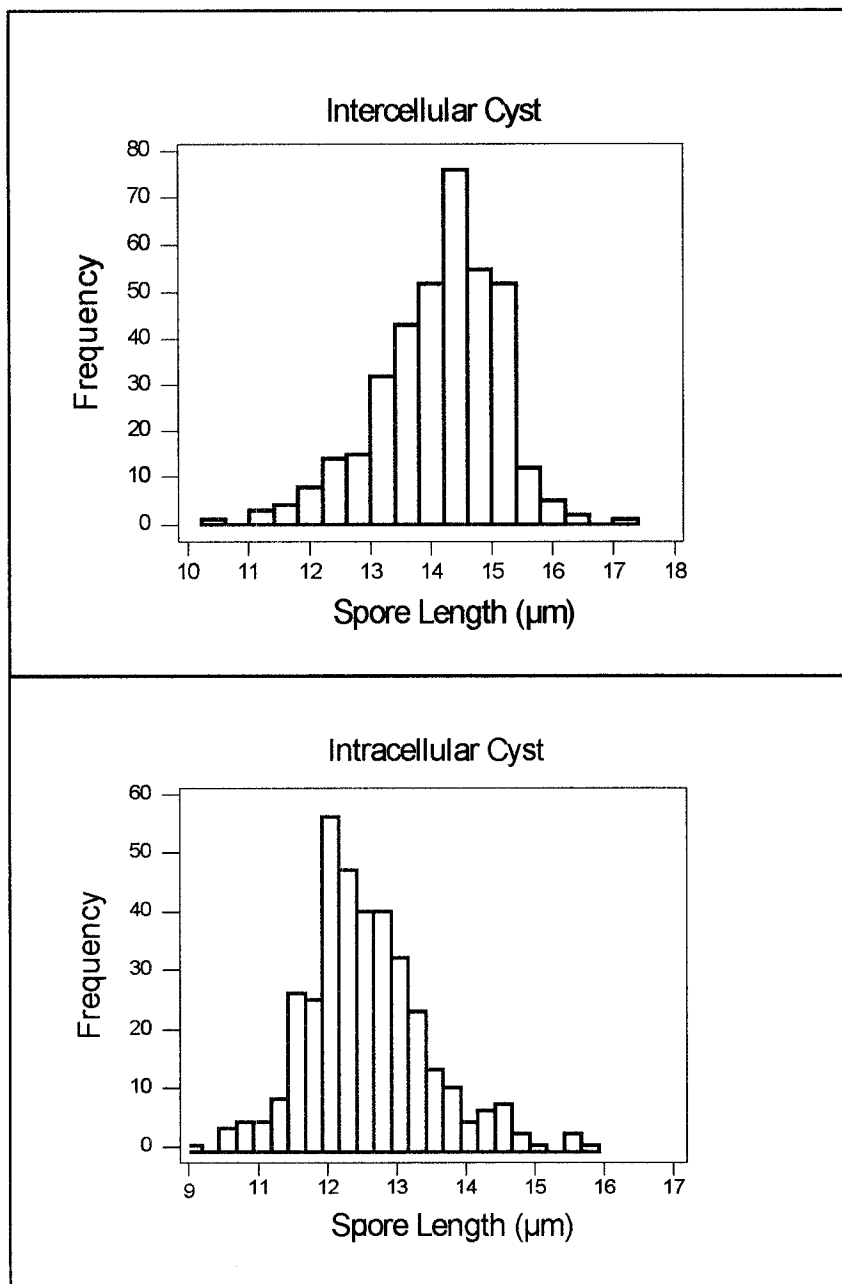


Figure 13. Frequency distributions of spore length within two cyst types of *Myxobolus procerus*. Spores in the larger intercellular cysts have a mean length of 14.5 μm while the smaller intracellular spores have a mean length of 12.0 μm.

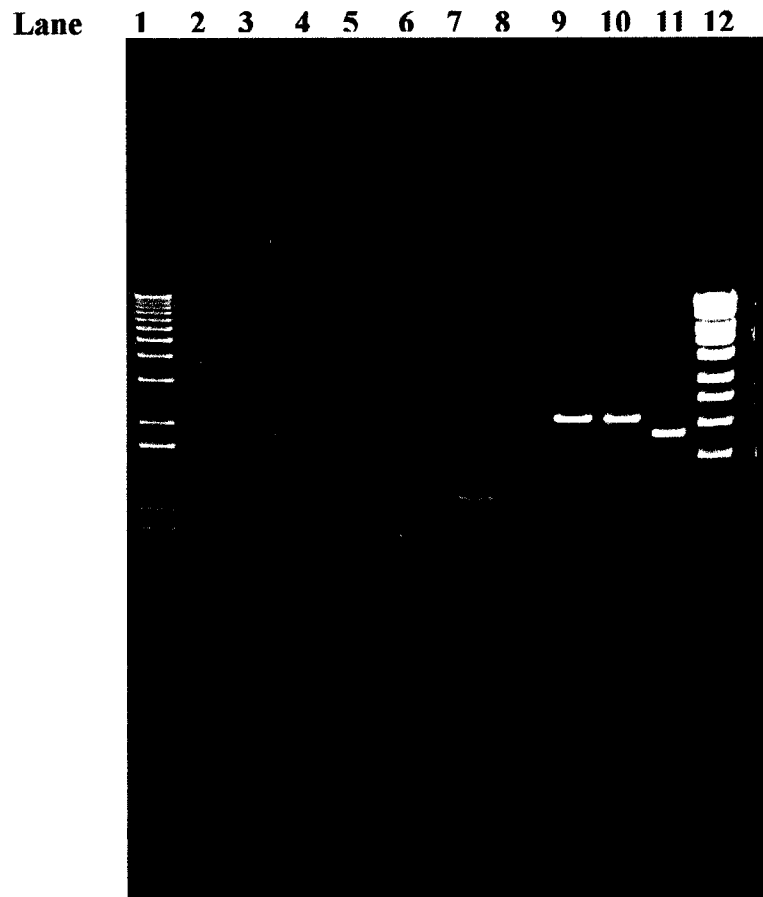


Figure 14. PCR products following amplification of SSU 18S rDNA from species of *Myxobolus*. PCR product using MX3-MX5 primer sets are illustrated in lanes 2-5. Lanes 6-8 with 18e-18m primer set. Lanes 9-11 with primers 18e-Myxgen2r. Lane 1 – 1kb + ladder. Lane - 12 Mass Ladder.

CTGCGGACGGCTCAGTATATCAGTTATAATCTGCTCGATTGTTATGGTTATTGGATAACCGTG
 GGAAATCTAGAGCTAATACATGCTATAATGTTGGGTGTGGTTTCGCTGCATTTCAGCCGCATTT
 ATTAGAGAATACCAATCGCATGAGCAATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGC
 ATGGCCTTTGAGCCGGCGACATTTTCGATTGAGTTTCTGCCCTATCACCTTGATGCTAGTGTA
 TTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGGTTTCGATCCCGGAGAGGGAGCC
 TTAGAAACGGCTACCAATCCAAGGAAGGCAGCAGGCGCGCAAATTACCCAATCCAGACAA
 TGGGAGGTGGTGACGAAGAGTACCGGATGACTGATATATTATTAGTCAGTCTGGAATGAAAT
 CAATTTAAGCAATTGAGTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGT
 AATTCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAACGCTCGTAGTTGGATGAG
 AATCTATATATCGGATAGTTATCTACTAGATTTAAGGCTAATTATTACTTATTTCCGTGATGCT
 TATGGTATCCGACAATATTGATATTGGTTCAGCTAGTGATAACATTAACTGGTATGTAGTA
 ATTGCGTATGAGGATGACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATGGGATGTGC
 TTGAGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAATGTTGATAGCATGGAACGAAC
 AATAGTGTAGATGTGTATTGAAGATGGTAGTGTATTATTACAATGTAATAGTATACTACTACAA
 TATGCAACACCGACCGCCAATTACGGATGTTGGTTCGCTATTGGGGTACTGATTAAGAGGAG
 CGGTTGGGGGCATTGGTATTTGGCAGCGAGAGGTGAAATTCTTGGACCTGCACAAG

Figure 15. *Myxobolus procerus* intercellular SSU 18S rDNA consensus sequence.

CTGCGGACGGCTCAGTAAATCAGTTATAATCTGCTCGATTGTTATGGTTATTGGATAACCGT
 GGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTGTGGTTCGCTGCATTCAGCCGCATT
 TATTAGAGAATACCAATCGCATGAGCAATCATGTGTGGTGAATCTAGATAACTTTGCTGATCG
 CATGGCCTTTGAGCCGGCGACATTTGATTGAGTTTCTGCCCTATCACCTTGATGCTAGTGT
 ATTGAACTAGCATGGGAGTAACGGGTAAACGGAGGATCAGGGTTCGATCCCGGAGAGGGAG
 CCTTAGAAACGGCTACCACATCCAAGGAAGGCAGCAGGCGCGCAAATTACCCAATCCAGAC
 AATGGGAGGTGGTGACGAAGAGTACCGGATGACTGATATATTATTAGTCAGTCTGGAATGAA
 ATCAATTTAAGCAATTGAGTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCG
 GTAATTCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAAACGCTCGTAGTTGGATG
 AGAAACTACATATCGGATAGTTATTTACTAGATTTAGGACTAATTATTACATATTTCCGTGATG
 CTTATGGTATCCGGATAATAGTGATATTGGTTCAACTAGTGATAACATTAACTGGTATGTAG
 TAATTGCGTATGAGGATGACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATGGGATGT
 GCCTTGAGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAATGTTGATAGCATGGAAC
 GAACAATAGTGTAGATGTGTATTGAAGATGGTAGTGTATTATTACAATGTAATAGTATACTACT
 ACAATATGCAACACCGACCGCCAATTACGGATGTTGGTTCCGTATTGGGGTACTGATTAAGA
 GGAGCGGTTGGGGGCATTGTATTGCAGCGAGAGTGAATTCTGACCTGCCAAG

Figure 16. *Myxobolus procerus* intracellular small subunit 18S rDNA consensus sequence.

Sequences producing significant alignments		Bit Score	E Value
gi 4103271 gb AF021878.1 	<i>Aurantiactinomyxon mississippiens...</i>	420	e-114
gi 4103274 gb AF021881.1 	<i>Henneguya exilis</i> 18S small subuni...	414	e-112
gi 11066099 gb AF195510.1 	<i>Henneguya ictaluri</i> small subunit...	414	e-112
gi 23429474 gb AY129317.1 	<i>Myxobolus exiguus</i> 18S ribosomal ...	394	e-106
gi 23429470 gb AY129314.1 	<i>Myxobolus muelleri</i> 18S ribosomal...	394	e-106
gi 23429476 gb AY129318.1 	<i>Myxobolus bizerti</i> 18S ribosomal ...	391	e-105
gi 23429467 gb AY129312.1 	<i>Myxobolus episquamalis</i> 18S ribos...	375	e-100
gi 21729516 gb AF378341.2 	<i>Myxobolus spinacurvatura</i> 18S rib...	359	6e-96
gi 13172451 gb AF306794.1 	AF306794 <i>Henneguya lesteri</i> 18S ri...	351	1e-93
gi 13172453 gb AF306791.1 	AF306791 <i>Endocapsa rosulata</i> 18S r...	323	3e-85

Figure 17. BLAST results for SSU 18S rDNA from spores of *Myxobolus procerus* - Intercellular cyst types of trout-perch.

Sequences producing significant alignments		Bit Score	E Value
gi 4103271 gb AF021878.1 	<i>Aurantiactinomyxon mississippiensis...</i>	<u>436</u>	e-119
gi 4103274 gb AF021881.1 	<i>Henneguya exilis</i> 18S small subuni...	<u>430</u>	e-117
gi 11066099 gb AF195510.1 	<i>Henneguya ictaluri</i> small subunit...	<u>430</u>	e-117
gi 23429474 gb AY129317.1 	<i>Myxobolus exiguus</i> 18S ribosomal ...	<u>412</u>	e-112
gi 23429470 gb AY129314.1 	<i>Myxobolus muelleri</i> 18S ribosomal...	<u>412</u>	e-112
gi 23429476 gb AY129318.1 	<i>Myxobolus bizerti</i> 18S ribosomal ...	<u>406</u>	e-110
gi 23429467 gb AY129312.1 	<i>Myxobolus episquamalis</i> 18S ribos...	<u>392</u>	e-106
gi 21729516 gb AF378341.2 	<i>Myxobolus spinacurvatura</i> 18S rib...	<u>377</u>	e-101
gi 13172451 gb AF306794.1 AF306794	<i>Henneguya lesteri</i> 18S ri...	<u>367</u>	2e-98
gi 13172453 gb AF306791.1 AF306791	<i>Endocapsa rosulata</i> 18S r...	<u>339</u>	6e-90

Figure 18. BLAST results for SSU 18S rDNA from spores of *Myxobolus procerus* - Intracellular cyst types of trout-perch.

Intercellular CTGCGGACGGCTCAGTATATCAGTTATAATCTGCTCGATTGTTATGGTTA
 Intracellular CTGCGGACGGCTCAGTAAATCAGTTATAATCTGCTCGATTGTTATGGTTA

Intercellular TTGGATAACCGTGGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTG
 Intracellular TTGGATAACCGTGGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTG

Intercellular TGGTTCGCTGCATTAGCCGCATTTATTAGAGAATACCAATCGCATGAGC
 Intracellular TGGTTCGCTGCATTAGCCGCATTTATTAGAGAATACCAATCGCATGAGC

Intercellular AATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGCATGGCCTTTGAG
 Intracellular AATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGCATGGCCTTTGAG

Intercellular CCGGCGACATTTGATTGAGTTTCTGCCCTATCACCTTGATGCTAGTGTA
 Intracellular CCGGCGACATTTGATTGAGTTTCTGCCCTATCACCTTGATGCTAGTGTA

Intercellular TTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGG -TTGATCCCCG
 Intracellular TTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGGTTTCGATCCCCG

Intercellular GAGAGGGAGCCTTAGAAACGGCTACCACATCCAAGGAAGGCAGCAGGCGC
 Intracellular GAGAGGGAGCCTTAGAAACGGCTACCACATCCAAGGAAGGCAGCAGGCGC

Intercellular GCAAATTACCAATCCAGACAATGGGAGGTGGTGACGAAGAGTACCGGAT
 Intracellular GCAAATTACCAATCCAGACAATGGGAGGTGGTGACGAAGAGTACCGGAT

Intercellular GACTGATATATTATTAGTCAGTCTGGAATGAAATCAATTTAAGCAATTGA
 Intracellular GACTGATATATTATTAGTCAGTCTGGAATGAAATCAATTTAAGCAATTGA

Intercellular GTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAATTC
 Intracellular GTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAATTC

Intercellular CAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAACGCTCGTAGTT
 Intracellular CAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAACGCTCGTAGTT

Intercellular GGATGAGAATCTATATATCGGATAGTTATCTACTAGATTTAAGGCTAATT
 Intracellular GGATGAGAACTACATATCGGATAGTTATTTACTAGATTTAGGACTAATT

Intercellular ATTACTTATTTCCGTGATGCTTATGGTATCCGGACAATAATTGATATTGGT
 Intracellular ATTACATATTTCCGTGATGCTTATGGTATCCGGATAATAGTGATATTGGT

Intercellular TCAGCTAGTGATAACATTAACCTGGTATGTAGTAATTGCGTATGAGGATG
 Intracellular TCAACTAGTGATAACATTAACCTGGTATGTAGTAATTGCGTATGAGGATG

Intercellular ACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATGGGATGTGC -TTG
 Intracellular ACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATGGGATGTGCCTTG

Intercellular AGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAATGTTGATAGCAT
 Intracellular AGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAATGTTGATAGCAT

Intercellular GGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTAGTGTATTATTAC
 Intracellular GGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTAGTGTATTATTAC

Intercellular AATGTAATAGTATACTACTACAATATGCAACACCGACCGCCAATTACGGA
 Intracellular AATGTAATAGTATACTACTACAATATGCAACACCGACCGCCAATTACGGA

Intercellular TGTTGGTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGGGGGCATTG
 Intracellular TGTTGGTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGGGGGCATTG

Intercellular GTATTTGGCAGCGAGAGGTGAAATCTTGGACCTGCACAAG
 Intracellular - TATT- - GCAGCGAGAG -TGAA -TTCT - - GACCTGC - CAAG

Figure 19. Clustal X alignment of *Myxobolus procerus* small subunit 18S rDNA obtained from spores contained within intercellular and intracellular cysts in trout-perch. Differences are noted in red.

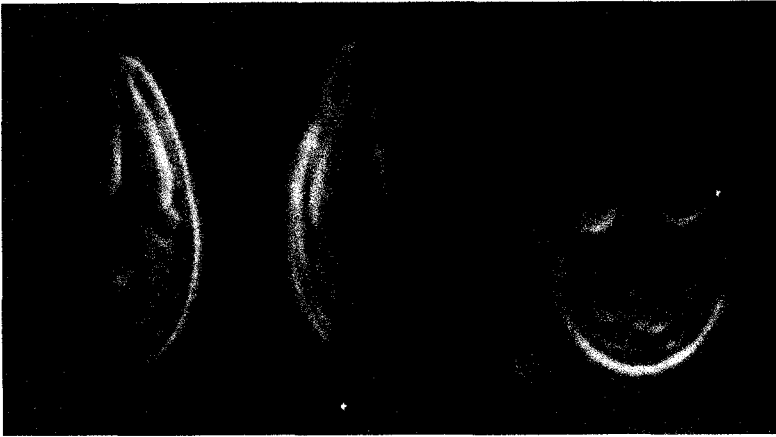


Figure 20. Spores of *Myxobolus neurophilus* from yellow perch. Polar capsules appear as raised oval shaped structures uneven in size. Scale bar at right is 10 μm .



Figure 21. Spores of *Myxobolus* sp. from Johnny darter. Scale bar at right is 10 μm .

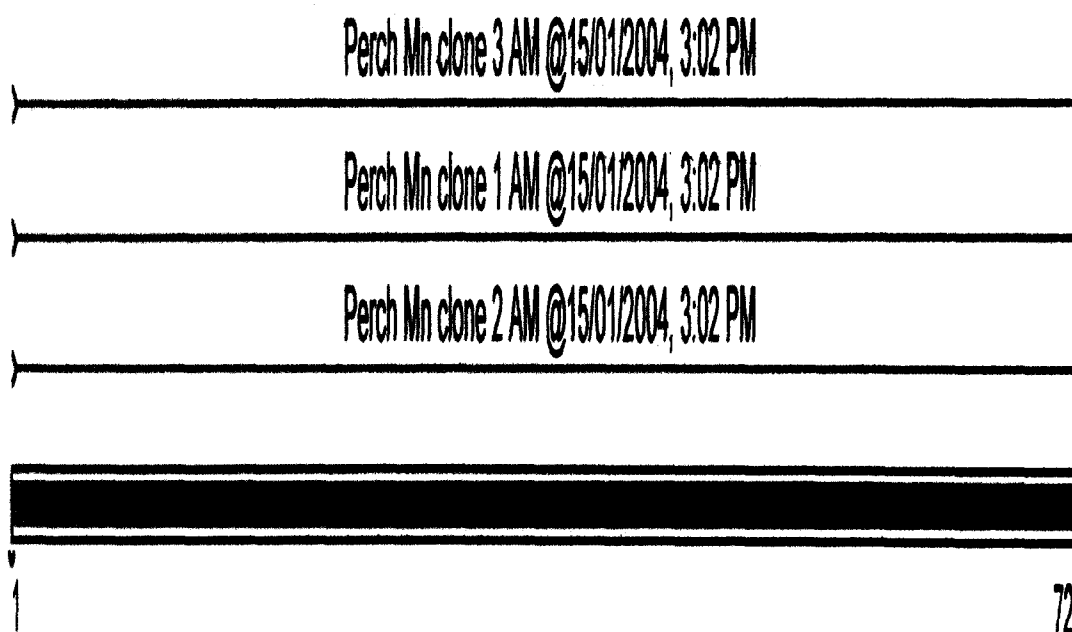


Figure 22. Clones of *Myxobolus neurophilus* from yellow perch. Three clones from a single primer set with ambiguous regions removed. There were no differences.

TGCAATGTTTGACTGGAGGGAAAGTGCAACTTCAACACCCATCCACACCAAGCATGTCATAA
ACACACCCAATGCTTCAGAGTATCTCTTTCACACCTTACCAACAATTCAAATCATATCAATACA
GACCTGTTACAGTCTATAAAGAAGATTTGAATTTACTCCAGCTCGTCAAATGGAGATTGTGAT
CTCGTCCGTTACCGGAATAAACCTGACAAATCACTTCACGAACCTAAGAACGGCCATGCACCA
CCAATCACCAGATCATGTAAGATCTTCGGTCTGACAATCCTTTTGATGTCCTGATCCGGTAAG
TTTTCCCGTGTTGAGTCAAATTAAGCCGCAGGCTCCACCCCTGGTGGTGCCCTTCCGTCAAT
TCCTTTAAGTTTCAGCCTTGCGACCATACTCCCCCGCAACCGAAAACTTTAGG:TTTCCCA
GGGGGACCAACCCAAGCCGTAATAATAAGGCATAGGACTAATCCCGGGTCGGCATAGTTTA
CAGTGAGAACTAGGACGGTATCTGATCGTCTTCGAACCTCTCACTATCGTTCTTGATTAATG
GATACGGTCTGGACAAATGCCTTCGCATTTGTTAGTCCTTGGCAGGTCCAAGAATTTACCT
CTCGCTGCCAAATACCAATGCCCCCAACCGCTCCTCTTAATCAGTTACCCAATACGGAACC
AACATCCGTAATTGGCGGTTCGGTGTTCATATTGAAG

Figure 23. DNA sequence of small subunit 18S rDNA from yellow perch isolates.

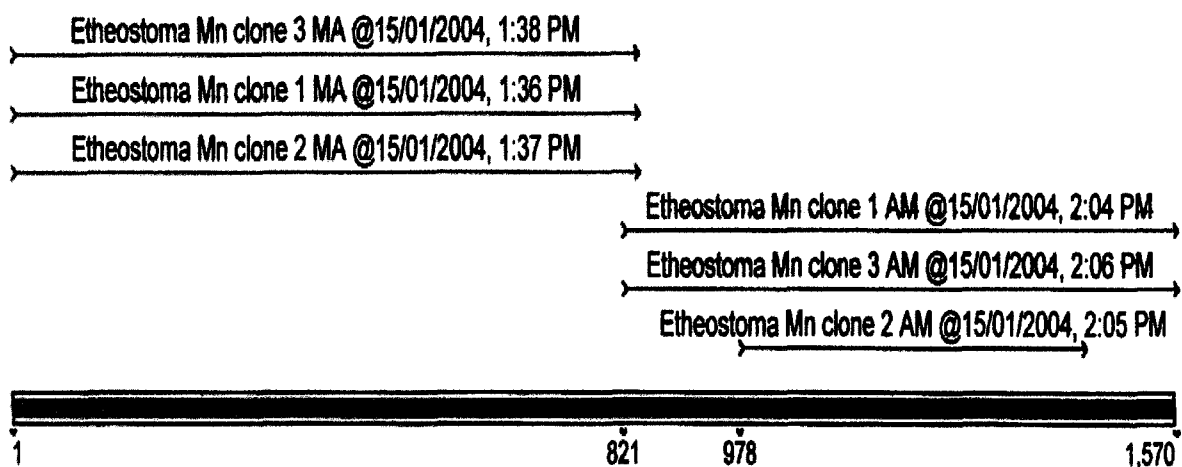


Figure 24. Clones of *Myxobolus* sp. from Johnny darter. Three clones from each of two primer sets with ambiguous regions removed. Differences are noted in text.

CTAACATGCTGGATGTAGCTTGCTACATTTGGCCGCATTTATTAGATAATACCAATCACATGA
 GCAATCATGTGTGATGAATCTAGATAACTTTGCTGATCGCATGACCTTGTGTGCGGCGACATTT
 CGATTGAGTTTCTGCCCTATCACTTGATGCTAGTGTATTGAACTAGCATGGGAGTTACGGGT
 AACGGAGGATCAGGGTTCGATCCCGGAGAGGGAGCCTTAGAAACGGCTACCACATCCAAG
 GAAGGCAGCAGGCGCGCAAATTACCCAATCCAGACAATGGGAGGTGGTGACGAAGAGTAC
 CGGATGACTAAAAAATTTTTAGTCAGTCTGGAATGAAATCAATTTAAGCAATTGAGTTGAGT
 AACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGGTAATTCCAGCTCCAGTAGCGTGTC
 TCAAAGTTGCTGCGCTTGAAACGCTCGTAGTTGGATGAGAACTACACATTGGATAGTTATTA
 TCTAGGTTATAGACTACATATTTTCGTATATCTGTGATGCTTTCAGTGTCCAGATAATATGGGT
 ATTAGTTGAAGTAGTAGTAACATATAACTGGTGTGTAGTAATTGCGTATGAGAATGATCATTG
 ACCTTGAGTGTGTTGAAGATCGTGTCTCATGGGATGTGCCTTGAGTAAATCAGAGTGCTCAA
 AGCAGGCGTTATGCCTGAATGTTAATAGCATGGAACGAACAATAGTGTAGATGTGTATTGAA
 GATGGTAGTATGTTATTATAACTTAATAATATACTACTACAATATGCAACACCGACCGCCAATT
 ACGGATGTTGGTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGGGGGCATTGGTATTT
 GGCAGCGAGAGGTGAAATTCTTGGACCTGCCAAGGACTAACAAATGCGAAGGCATTTGTCC
 AGACCGTATCCATTAATCAAGAACGATAGTGAGAGGTTTGAAGACGATCAGATACCGTCCTA
 GTTCTCACTGTAAACTATGCCGACCCGGGATTAGTCCTATGCCTTATTTTTACGGCTTGGGT
 GGTCCCCCTGGGAAACCTAAAGTTTTTCGGTTGCGGGGGGAGTATGGTCGCAAGGCTGAAA
 CTAAAGGAATTGACGGAAGGGCACCACCAGGGGTGGAGCCTGCGGCTTAATTTGACTCAA
 CACGGGAAAACCTACCGGATCAGGACATCAAAGGATTGTCAGACCGAAGATCTTACATGAT
 CTGGTGACTGGTGGTGATGGCCGTTCTTAGTTCGTGAAGTGATTTGTCAGGTTTATTCCGG
 TAACGGACGAGATCACAATCTCCATTTGACGAACTGGAGTAAATGCAAAATAGTCATAATTAAT
 TGTAAGGATTGATTGTGATATGATTTGCATTGTTGGTAAATGTGAAAGATATAATCATAAT
 GATGAAATATAGGGTAAAACTATATTGAATTAGAATGAGTATGGAAGTTATTTTTACCTCCA
 GTTAAACAGTGTATAGTTAACTTTAACTGTGTGCTGTATCATGGAGAGACAACGGAAATAAT
 AAAAATCGAGGAA

Figure 25. DNA sequence of small subunit 18S rDNA myxobolid sp. from Johnny darter.

Sequences producing significant alignments		Bit Score	E Value
gi 18071158 gb AF378344.1 	<i>Henneguya zschokkei</i> 18S ribosoma...	<u>428</u>	e-117
gi 27369410 gb AY165182.1 	<i>Henneguya weishanensis</i> 18S ribos...	<u>420</u>	e-114
gi 11066099 gb AF195510.1 	<i>Henneguya ictaluri</i> small subunit...	<u>412</u>	e-112
gi 18071167 gb AF378353.1 	<i>Neoactinomyxum</i> sp. KAB-2001 18S ...	<u>412</u>	e-112
gi 13172455 gb AF306793.1 AF306793	<i>Tetraspora discoidea</i> 18S...	<u>412</u>	e-112
gi 13172454 gb AF306792.1 AF306792	<i>Triactinomyxon</i> sp. 18S r...	<u>412</u>	e-112
gi 13172451 gb AF306794.1 AF306794	<i>Henneguya lesteri</i> 18S ri...	<u>412</u>	e-112
gi 4103271 gb AF021878.1 	<i>Aurantiactinomyxon mississippiensis</i> ...	<u>404</u>	e-110
gi 13172452 gb AF306790.1 AF306790	<i>Sphaeractinomyxon ersei</i> ...	<u>404</u>	e-110
gi 4103274 gb AF021881.1 	<i>Henneguya exilis</i> 18S small subuni...	<u>396</u>	e-107
gi 21729516 gb AF378341.2 	<i>Myxobolus spinacurvatura</i> 18S rib...	<u>396</u>	e-107
gi 18071170 gb AF378356.1 	<i>Aurantiactinomyxon</i> sp. KAB-2001 ...	<u>391</u>	e-105
gi 18071166 gb AF378352.1 	<i>Raabeia</i> sp. KAB-2001 18S ribosom...	<u>391</u>	e-105
gi 18071151 gb AF378337.1 	<i>Myxobolus ichkeulensis</i> 18S ribos...	<u>391</u>	e-105
gi 23429476 gb AY129318.1 	<i>Myxobolus bizerti</i> 18S ribosomal ...	<u>375</u>	e-101

Figure 26. BLAST results for small subunit 18S rDNA sequence from yellow perch isolates against those sequences available in GenBank.

Sequences producing significant alignments		Bit Score	E Value
gi 27369410 gb AY165182.1	<i>Henneguya weishanensis</i> 18S ribos...	<u>434</u>	e-118
gi 11066099 gb AF195510.1	<i>Henneguya ictaluri</i> small subunit...	<u>426</u>	e-116
gi 18071167 gb AF378353.1	<i>Neoactinomyxum</i> sp. KAB-2001 18S ...	<u>426</u>	e-116
gi 13172455 gb AF306793.1	AF306793 <i>Tetraspora discoidea</i> 18S...	<u>426</u>	e-116
gi 13172454 gb AF306792.1	AF306792 <i>Triactinomyxon</i> sp. 18S r...	<u>426</u>	e-116
gi 13172451 gb AF306794.1	AF306794 <i>Henneguya lesteri</i> 18S ri...	<u>426</u>	e-116
gi 4103271 gb AF021878.1	<i>Aurantiactinomyxon mississippiensis</i> ...	<u>418</u>	e-113
gi 13172452 gb AF306790.1	AF306790 <i>Sphaeractinomyxon ersei</i> ...	<u>418</u>	e-113
gi 4103274 gb AF021881.1	<i>Henneguya exilis</i> 18S small subuni...	<u>410</u>	e-111
gi 21729516 gb AF378341.2	<i>Myxobolus spinacurvatura</i> 18S rib...	<u>410</u>	e-111
gi 18071170 gb AF378356.1	<i>Aurantiactinomyxon</i> sp. KAB-2001 ...	<u>404</u>	e-109
gi 18071166 gb AF378352.1	<i>Raabeia</i> sp. KAB-2001 18S ribosom...	<u>404</u>	e-109
gi 23429472 gb AY129315.1	<i>Myxobolus ichkeulensis</i> 18S ribos...	<u>404</u>	e-109
gi 23429474 gb AY129317.1	<i>Myxobolus exiguus</i> 18S ribosomal ...	<u>402</u>	e-109
gi 23429470 gb AY129314.1	<i>Myxobolus muelleri</i> 18S ribosomal...	<u>402</u>	e-109

Figure 27. BLAST results for myxobolid small subunit 18S rDNA sequence from Johnny darter against those sequences available in GenBank.

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Perch      GTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGGGGGCATTGGTATT
Etheostoma GTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGGGGGCATTGGTATT

Perch      TGGCAGCGAGAGGTGAAATTCTTGGACCTGCCAAGGACTAACAAATGCGA
Etheostoma TGGCAGCGAGAGGTGAAATTCTTGGACCTGCCAAGGACTAACAAATGCGA

Perch      AGGCATTTGTCCAGACCGTATCCATTAATCAAGAACGATAGTGAGAGGTT
Etheostoma AGGCATTTGTCCAGACCGTATCCATTAATCAAGAACGATAGTGAGAGGTT

Perch      CGAAGACGATCAGATACCGTCCTAGTTCTCACTGTAAACTATGCCGACCC
Etheostoma CGAAGACGATCAGATACCGTCCTAGTTCTCACTGTAAACTATGCCGACCC

Perch      GGGATTAGTCCTATGCCTTATTATTACGGCTTGGGTGGTCCCCCTGGGA
Etheostoma GGGATTAGTCCTATGCCTTATTTTACGGCTTGGGTGGTCCCCCTGGGA

Perch      AACCTAAAGTTTTTCGGTTGCGGGGGGAGTATGGTCGCAAGGCTGAAACT
Etheostoma AACCTAAAGTTTTTCGGTTGCGGGGGGAGTATGGTCGCAAGGCTGAAACT

Perch      TAAAGGAATTGACGGAAGGGCACCACCAGGGGTGGAGCCTGCGGCTTAAT
Etheostoma TAAAGGAATTGACGGAAGGGCACCACCAGGGGTGGAGCCTGCGGCTTAAT

Perch      TTGACTCAACACGGGAAAACTTACCGGATCAGGACATCAAAAGGATTGTC
Etheostoma TTGACTCAACACGGGAAAACTTACCGGATCAGGACATCAAAAGGATTGTC

Perch      AGACCGAAGATCTTACATGATCTGGTGATTGGTGGTGCATGGCCGTTCTT
Etheostoma AGACCGAAGATCTTACATGATCTGGTGATTGGTGGTGCATGGCCGTTCTT

Perch      AGTTCGTGAAGTGATTTGTCAGGTTTATTCCGGTAACGGACGAGATCACA
Etheostoma AGTTCGTGAAGTGATTTGTCAGGTTTATTCCGGTAACGGACGAGATCACA

Perch      ATCTCCATTTGACGAGCTGGAGTAAATGCAAATCTTCTTTATAGACTGTA
Etheostoma ATCTCCATTTGACGAACTGGAGTAAATGCAAATAGTCATAATTAATTGTA

Perch      ACAGGTCTGTAT- TGATATG- ATTTGAATTGTTGGTAAGGTGTGAAAGAG
Etheostoma AAAGATCTGATTGTGATATGTATTTGCATTGTTGGTAAAATGTGAAAGAT

Perch      ATACTC-----TGAAGCATTGGGTGTGTTTATGAC- ATGCTTGGTGTG
Etheostoma ATAATCATAATGATGAAATATAGGGTA-----AAAACATA TTGAATTAG

Perch      GATGGGTGTTGAAGTTGCACTTTCCCTCCAGTCAAACATTGCATGG
Etheostoma AATGAGTATGGAAGTTATATTTTACCTCCAGT TAAACAGTGTATAG

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Figure 28. Clustal X alignment of *Myxobolus* SSU 18S rDNA Isolated from yellow perch and Johnny darter. Data was obtained from sequencing of PCR products using myxozoan specific primers Act3F-MX3. Differences are noted in red.

Intercellular	GATAACCGTGGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTG
Intracellular	GATAACCGTGGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTG
Etheostoma	GATAACCGTGGGAAATCTAGAGCTAATACATGCTAACATGCTGGATG
Intercellular	TGGTTCGCTGCATTTCAGCCGCATTTATTAGAG-AATACCAATCGCATG
Intracellular	TGGTTCGCTGCATTTCAGCCGCATTTATTAGAG-AATACCAATCGCATG
Etheostoma	TAGCTTGCTACATTTGGCCGCATTTATTAGATTAATACCAATCACATG
Intercellular	AGCAATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGCATGGCCTTT
Intracellular	AGCAATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGCATGGCCTTT
Etheostoma	AGCAATCATGTGTGATGAATCTAGATAACTTTGCTGATCGCATGACCTT-
Intercellular	GAGCCGGCGACATTTTCGATTGAGTTTCTGCCCTATCACCTTGATGCTAGT
Intracellular	GAGCCGGCGACATTTTCGATTGAGTTTCTGCCCTATCACCTTGATGCTAGT
Etheostoma	GTGTCGGCGACATTTTCGATTGAGTTTCTGCCCTATCACCTTGATGCTAGT
Intercellular	GTATTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGG-TTCGATC
Intracellular	GTATTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGGGTTTCGATC
Etheostoma	GTATTGAACTAGCATGGGAGTTACGGGTAACGGAGGATCAGGGTTTCGATC
Intercellular	CCGGAGAGGGAGCCTTAGAAACGGCTACCACATCCAAGGAAGGCAGCAG
Intracellular	CCGGAGAGGGAGCCTTAGAAACGGCTACCACATCCAAGGAAGGCAGCAG
Etheostoma	CCGGAGAGGGAGCCTTAGAAACGGCTACCACATCCAAGGAAGGCAGCAG
Intercellular	GCGCGCAAATTACCCAATCCAGACAATGGGAGGTGGTGACGAAGAGTAC
Intracellular	GCGCGCAAATTACCCAATCCAGACAATGGGAGGTGGTGACGAAGAGTAC
Etheostoma	GCGCGCAAATTACCCAATCCAGACAATGGGAGGTGGTGACGAAGAGTAC
Intercellular	CGGATGACTGATATATTATT-AGTCAGTCTGGAATGAAATCAATTTAAGC
Intracellular	CGGATGACTGATATATTATT-AGTCAGTCTGGAATGAAATCAATTTAAGC
Etheostoma	CGGATGACTAAAAATTTTATTAGTCAGTCTGGAATGAAATCAATTTAAGC
Intercellular	AATTGAGTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGG
Intracellular	AATTGAGTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGG
Etheostoma	AATTGAGTTGAGTAACTACTGGAGGGCAAGTCTGGTGCCAGCAGCCGCGG
Intercellular	TAATTCCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAAACGCTC
Intracellular	TAATTCCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAAACGCTC
Etheostoma	TAATTCCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAAACGCTC
Intercellular	GTAGTTGGATGAGAATCTATATATCGGATAGTTATCTACTAGATTTAAGG
Intracellular	GTAGTTGGATGAGAACTACATATCGGATAGTTATTTACTAGATTTAGGA
Etheostoma	GTAGTTGGATGAGAACTACACATTGGATAGTTATTATCTAGGTTATAGA
Intercellular	CTAATTATTA-CTTATTTCCGTGATGCTTATGGTATCCGGACAATA TTGA
Intracellular	CTAATTATTA-CATATTTCCGTGATGCTTATGGTATCCGGATAATA GTGA
Etheostoma	CTACATATTTTCGTATATCTGTGATGCTTTCAGTGTCAGATAATATGGG
Intercellular	TATTGGTTCAGCTAGTGATAACATTAACTGGTATGTAGTAATTGCGTAT
Intracellular	TATTGGTTCAGCTAGTGATAACATTAACTGGTATGTAGTAATTGCGTAT
Etheostoma	TATTAGTTGAACTAGTAGTAACATATAACTGGTGTGTAGTAATTGCGTAT
Intercellular	GAGGATGACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATGGGATG
Intracellular	GAGGATGACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATGGGATG
Etheostoma	GAGAATGATCATTGACCTTGAGTGTGTTGAAGATCGTGTCTCATGGGATG

Intercellular	TGCCTTGAGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAATGTTG
Intracellular	TGCCTTGAGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAATGTTG
Etheostoma	TGCCTTGAGTAAATCAGAGTGCTCAAAGCAGGCGTTATGCCTGAATGTTA
Intercellular	ATAGCATGGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTAGTGTA
Intracellular	ATAGCATGGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTAGTGTA
Etheostoma	ATAGCATGGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTAGTATG
Intercellular	TTATTACAATGTAATAGTATACTACTACAATATGCAACACCGACCGCCAA
Intracellular	TTATTACAATGTAATAGTATACTACTACAATATGCAACACCGACCGCCAA
Etheostoma	TTATTATAACTTAATAATATACTACTACAATATGCAACACCGACCGCCAA
Intercellular	TTACGGATGTTGGTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGG
Intracellular	TTACGGATGTTGGTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGG
Etheostoma	TTACGGATGTTGGTTCCGTATTGGGGTACTGATTAAGAGGAGCGGTTGG
Intercellular	GGGCATTGGTATTTGGCAGCGAGAGGTGAAATTCTTGGACCTGCACAAG
Intracellular	GGGCATTGGTATTTGGCAGCGAGAGGTGAAATTCTTGGACCTGCACAAG
Etheostoma	GGGCATTGGTATTTGGCAGCGAGAGGTGAAATTCTTGGACCTGC-CAAG

Figure 29. Clustal X Alignment of *Myxobolus procerus* SSU 18S rDNA from both cyst types in trout-perch with homologous region isolated from Johnny darter. Differences are noted in red. There are 52 differences out of a total of approximately 989 basepairs or a 5.2% difference.

Intercellular	TCAGTTATAATCTGCTCGATTGTTATGGTTA
Intracellular	TCAGTTATAATCTGCTCGATTGTTATGGTTA
Perch	CCA--TGCAAT--GTTTGACTG----GA---
Intercellular	TTGGATAACCGTGGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTG
Intracellular	TTGGATAACCGTGGGAAATCTAGAGCTAATACATGCTATAATGTTGGGTG
Perch	--GGGAAAGTGC---AACTTCAACACCCATCCACACCA-----AGCA
Intercellular	TGGTTCGCTGCATTTCAGCCGCATTTATTAGAGAATACCAATCGCA--TGA
Intracellular	TGGTTCGCTGCATTTCAGCCGCATTTATTAGAGAATACCAATCGCA--TGA
Perch	TG--TCA-TAAACACACCCA-ATGCTTCAGAGTATCTCTTTCACACCTTA
Intercellular	GCAATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGCATGGCCTTTG
Intracellular	GCAATCATGTGTGGTGAATCTAGATAACTTTGCTGATCGCATGGCCTTTG
Perch	CCAACAATTCAAATCATATCAATACAGACCTGTT-----ACAGTCTATA
Intercellular	AGCCGGCGACATTTTCGATTGAGTTTCTGCCCTATCACCTTGATGCTAGTG
Intracellular	AGCCGGCGACATTTTCGATTGAGTTTCTGCCCTATCACCTTGATGCTAGTG
Perch	AA--GAAGAT-----TTGAATTT-----ACTCCAGCT
Intercellular	TATTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGG-TTCGATCC
Intracellular	TATTGAACTAGCATGGGAGTAACGGGTAACGGAGGATCAGGTTTCGATCC
Perch	CGTCAA-----TGGAGATTGTG-----ATCTCG-TCCGTTAC
Intercellular	CGGAGAGGGAGCCTTAGAAACGGCTACCAC-ATCCAAGGAAGGCAGCAGG
Intracellular	CGGAGAGGGAGCCTTAGAAACGGCTACCAC-ATCCAAGGAAGGCAGCAGG
Perch	CGGAATA--AACCTGACAAATCACT-TCACGAATAAGAACGGC--CATG
Intercellular	CGC-GCAAATTACCCAATCCAGACAATGGGAGGTGGT--GACGAAGAGTA
Intracellular	CGC-GCAAATTACCCAATCCAGACAATGGGAGGTGGT--GACGAAGAGTA
Perch	CACCACCAATCACCAGATC-----ATGTAAGATCTTCGGTCTGACAATC
Intercellular	CCG--GATGAC-TGATATATTATTAGTCAGTCTGGAATGAAATCAATTTA
Intracellular	CCG--GATGAC-TGATATATTATTAGTCAGTCTGGAATGAAATCAATTTA
Perch	CTTTTGATGTCCTGATCCGGTA--AGTTT-TCCCGTGTGAGTCAAATTA
Intercellular	AGCAATTGAGTTGAGTAAGTACTGGAGGGCAAGTCTGGTGCCAGCAGCCG
Intracellular	AGCAATTGAGTTGAGTAAGTACTGGAGGGCAAGTCTGGTGCCAGCAGCCG
Perch	AGCCGCAGGCT----CCACCCCTGG-----TGGTGCC--CTTCCG
Intercellular	CGGTAATTCCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAACG
Intracellular	CGGTAATTCCAGCTCCAGTAGCGTGTCTCAAAGTTGCTGCGCTTAAACG
Perch	T--CAATTCC---TTTAA-----GTTTCAGCCTTGCGAC-CATACTCC
Intercellular	CTCGTAGTTGGATGAGAATCTATATATCGGATAGTTATCTACTAGATTTA
Intracellular	CTCGTAGTTGGATGAGAACTACATATCGGATAGTTATTTACTAGATTTA
Perch	CCCACAACCGAA----AACTTTA----GG-----TTTCCAGG---G
Intercellular	AGGCTAATTATTACTTATTTCCGTGATGCTTATGGTATCCGGACAATATT
Intracellular	GGACTAATTATTACATATTTCCGTGATGCTTATGGTATCCGGATAATAGT
Perch	GGACCAACC-----CAAG---CCGTAATAAT-AAGGCATAGGACTAATCCC
Intercellular	GATATTGGTTTCAGCT---AGTGATAACATTAAACTGGTATGTAGTAATTG
Intracellular	GATATTGGTTCAACT---AGTGATAACATTAAACTGGTATGTAGTAATTG
Perch	GG-GTCGGCATAGTTTACAGTGAGAAC--TAGGACGGTATCTG---ATCG

Intercellular	CGTATGAGGATGACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATG
Intracellular	CGTATGAGGATGACCATTGACCTTAAGTGTGTTGAAGGTCGTGTCTCATG
Perch	TCTTCGA-----ACCTCTCACTAT-----CGTT-----CTTGATTAATG
Intercellular	GGATGTGC-TTGAGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAA
Intracellular	GGATGTGCCTTGAGTAAATCAGAGTGCTCAAAGCAGGCGTGATGCCTGAA
Perch	G-ATACGGTCTGGACAAAT----GCCTTC---GCATTTGTTAGTCCT---
Intercellular	TGTTGATAGCATGGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTA
Intracellular	TGTTGATAGCATGGAACGAACAATAGTGTAGATGTGTATTGAAGATGGTA
Perch	-----TGGCA--GGTCCAAGAAT-----TTCACC-----
Intercellular	GTGTATTATTACAATGTAATAGTATACTACTACAATATGCAACACCGACC
Intracellular	GTGTATTATTACAATGTAATAGTATACTACTACAATATGCAACACCGACC
Perch	---TCTCGCTGC-----CAA-----ATACCA-----ATGC--CCCCAACC
Intercellular	GCCAATTACGGATGTTGGTT-CCGTATTGGGGTACTGATTAAGAGGAGCG
Intracellular	GCCAATTACGGATGTTGGTT-CCGTATTGGGGTACTGATTAAGAGGAGCG
Perch	GCTCCTCTTA---ATCAGTTACCCCAATACGGAACC----AACATCCGTA
Intercellular	GTTGGGGGCATTGGTATTTGGCAGCGAGAGGTGAAATTCTTGACCTGCA
Intracellular	GTTGGGGGCATTG-TATT--GCAGCGAGAG-TGAA-TTCT--GACCTGC-
Perch	ATTGGCGG--TCGGTGTT--GCA-----TAT-----TG--
Intercellular	CAAG
Intracellular	CAAG
Perch	-AAG

Figure 30. Clustal X alignment of *Myxobolus procerus* SSU 18S rDNA from both cyst types in trout-perch with homologous region isolated from yellow perch. As is evident there is a marked difference in the sequences. Differences are noted in red.

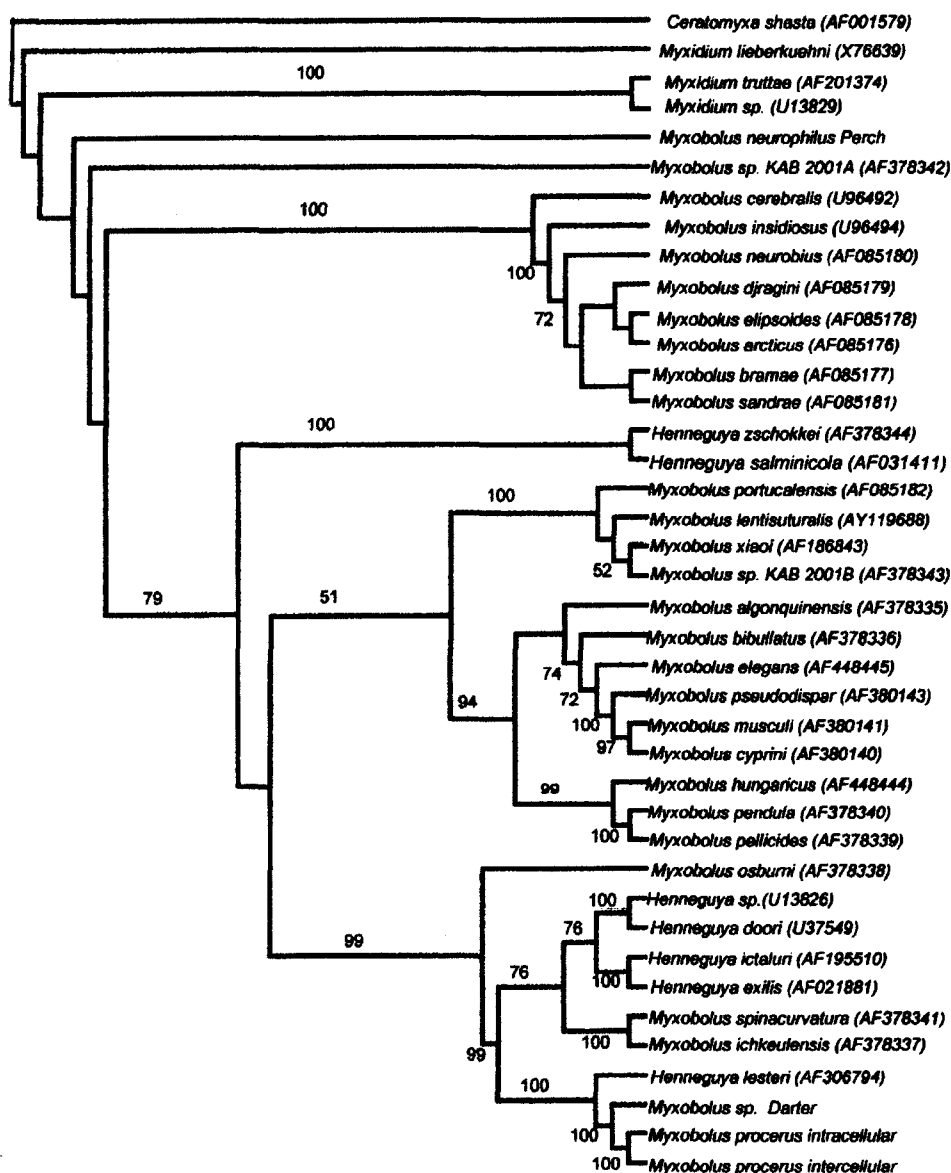


Figure 31. Neighbour-joining tree illustrating SSU 18S rDNA sequences of myxosporeans including data acquired from those species of *Myxobolus* included in the present work. The tree is rooted at *Ceratomyxa shasta*. Bootstrap values and GenBank Accession number (in parentheses) are included.

9. TABLES

Myxosporean	Fish host	Actinosporean	Invertebrate host	References
<i>Myxobolus cerebralis</i>	<i>Oncorhynchus mykiss</i>	triacetinomyxon	<i>Tubifex tubifex</i>	Wolf and Markiw 1984
<i>Myxobolus cotti</i>	<i>Cottus gobio</i>	triacetinomyxon	mixed oligochaetes	El-Matbouli and Hoffmann 1989
<i>Myxobolus pavlovskii</i>	<i>Hypophthalmichthys molitrix</i>	hexactinomyxon	mixed oligochaetes	Ruidisch et al. 1991
<i>Myxobolus culus</i>	<i>Carassius auratus</i>	raabeia	<i>Branchiura sowerhyi</i>	Yokoyama et al. 1995
<i>Myxobolus carassii</i>	<i>Leuciscus idus</i>	triacetinomyxon	<i>Tubifex tubifex</i>	El-Matbouli and Hoffmann 1993
<i>Myxobolus arcticus</i> (Canada)	<i>Oncorhynchus nerka</i>	triacetinomyxon	<i>Stylodrilus heringianus</i>	Kent et al. 1993b
<i>Myxobolus arcticus</i> (Japan)	<i>Oncorhynchus masu</i>	triacetinomyxon	<i>Lumbriculus variegatus</i>	Urawa 1994*
<i>Myxobolus drjagini</i>	<i>Hypophthalmichthys molitrix</i>	triacetinomyxon	<i>T. tubifex</i>	El-Mansy and Molnár 1997a
<i>Myxobolus portucalensis</i>	<i>Anguilla anguilla</i>	triacetinomyxon	<i>T. tubifex</i>	El-Mansy et al. 1998a
<i>Myxobolus hungaricus</i>	<i>Abramis abramis</i>	triacetinomyxon	<i>T. tubifex</i> , <i>L. hoffmeisteri</i>	El-Mansy and Molnár 1997b
<i>Myxobolus dispar</i>	<i>Cyprinus carpio</i>	raabeia	<i>T. tubifex</i>	Molnár et al. 1999a
<i>Myxobolus pseudodispar</i>	<i>Rutilus rutilus</i>	triacetinomyxon	<i>T. tubifex</i> , <i>L. hoffmeisteri</i>	Székely et al. 1999, 2000
<i>Myxobolus hyamue</i>	<i>Abramis brama</i>	triacetinomyxon	<i>T. tubifex</i>	Eszterbauer et al. 2000
<i>Henneguya exilis</i>	<i>Ictalurus punctatus</i>	<i>Aurantiactinomyxon jani-szewskai</i>	<i>Dero digitata</i>	Lin et al. 1999
<i>Henneguya ictaluri</i>	<i>Ictalurus punctatus</i>	<i>aurantiactinomyxon</i>	<i>D. digitata</i>	Burtle et al. 1991; Styer et al. 1991; Pote et al. 2000
<i>Hoferellus carassii</i> (Germany)	<i>Carassius auratus</i>	<i>aurantiactinomyxon</i>	mixed species	El-Matbouli et al. 1992b
<i>Hoferellus carassii</i> (Japan)	<i>Carassius auratus</i>	neoactinomyxon	<i>B. sowerhyi</i>	Yokoyama et al. 1993
<i>Hoferellus cyprini</i>	<i>Cyprinus carpio</i>	<i>aurantiactinomyxon</i>	<i>Nais</i> sp.	Grossheider and Körting 1992
<i>Thelohanellus nikolskii</i>	<i>Cyprinus carpio</i>	<i>aurantiactinomyxon</i>	<i>B. sowerhyi</i>	Székely et al. 1998
<i>Thelohanellus hovorkai</i>	<i>Cyprinus carpio</i>	<i>aurantiactinomyxon</i>	<i>B. sowerhyi</i>	Yokoyama 1997; Székely et al. 1998; Anderson et al. 2000
<i>Sphaerospora renicola</i>	<i>Cyprinus carpio</i>	undetermined neoactinomyxon	unknown <i>B. sowerhyi</i>	Grossheider and Körting 1993; Molnár et al. 1996b
<i>Sphaerospora truttae</i>	<i>Salmo trutta</i>	echinactinomyxon	<i>L. variegatus</i>	Özer and Wootten 1999
<i>Ceratomyxa shasta</i>	<i>Oncorhynchus mykiss</i>	tetractinomyxon	<i>Manayunkia speciosa</i>	Bartholomew et al. 1997
<i>Zschokkella</i> sp.	<i>Carassius auratus</i>	echinactinomyxon	<i>B. sowerhyi</i>	Yokoyama et al. 1993
<i>Zschokkella nova</i>	<i>Carassius carassius</i>	siedleckiella	<i>T. tubifex</i>	Uspenkaya 1995
<i>Myxidium giardi</i>	<i>Anguilla anguilla</i>	<i>aurantiactinomyxon</i>	<i>T. tubifex</i>	Benajiba and Marques 1993
PKX	<i>Oncorhynchus mykiss</i>	<i>Tetracapsula bryosalmonae</i>	<i>Plumutella</i> sp. and <i>Fred-ericella sultana</i>	Longshaw et al. 1999

* Urawa, S. 1994. Life cycle of *Myxobolus arcticus*, a myxosporean parasite of salmonid fishes. In: Program and Abstracts Intl. Symposium, Aquat. Animal Health Seattle, WA. 4-8 Sept. 1994. University of California, Davis. p. W-10.3.

Table 1. A summary of life cycles of myxozoans (Kent et al., 2000).

Primer	Sequence	Annealing Temperature (°C)	Reference
MX5	5' ctgcggacggctcagtaaatacgt 3'	55	Kent et al., 2001
MX3	5' ccaggacatcttagggcatcacaga 3'	55	Kent et al. 2001
18e	5' ctggttgatcctgccagt 3'	55	Hillis and Dixon 1991
18g	5' ggtagtagcgacgggcggtgtg 3'	55	Hillis and Dixon 1991
Myxgen2r	5' caratgcyttcgwyttgtta 3'	55	Kent et al., 2000
18r	5' ctacggaaacctgttacg 3'	55	Whipps et al. 2003
MX3-1n	5' tgtatcatggagagacaacg 3'	55	This paper
MX3-3n	5' cattcaggcatcacgcctgc 3'	55	This paper
18M-3n	5' ggttacacttcccttccg 3'	55	This paper
Myxgp2F	5' tggataaccgtgggaaa 3'	55	Kent et al. 2001
Act1R	5' aatttcacctctcgctgcca 3'	55	Hallet and Diamant 2001
Act3F	5' catggaacgaacaat 3'	55	Hallet and Diamant 2001

Table 2. Primer sequences used to amplify SSU 18S rDNA from isolates of myxobolids.

<i>Myxobolus procerus</i> intercellular	Primer	Number of clones	Differences/base pairs sequenced
	18e	3	0/647
	MX3	3	0/752

<i>Myxobolus procerus</i> intracellular	Primer	Number of clones	Differences/base pairs sequenced
	MX3	3	0/631
	Myxgen2r (18r)	2	0/473

Table 3. Aligning clones of *Myxobolus procerus*. There are no differences within those clones generated for each primer set noted.

In (μM)	<i>Myxobolus procerus</i> Interacellular cyst type	<i>Myxobolus procerus</i> Intracellular cyst type	Kruskal-Wallis
Spore length	14.13 (0.97)	12.50 (0.91)	0.000
Spore width	6.22 (0.55)	6.22 (0.57)	0.748
Polar Capsule 1 Length	5.94 (0.61)	5.81 (0.62)	0.486
Polar Capsule 1 Width	1.71 (0.30)	1.76 (0.39)	0.573
Polar Capsule 2 length	5.89 (0.62)	5.82 (0.65)	0.329
Polar Capsule 2 Width	1.71 (0.30)	1.73 (0.28)	0.939

Table 4. Statistical analysis for *Myxobolus procerus* spore measurements - ANOVA. *Myxobolus procerus* morphotypes of intercellular and intracellular spores average dimensions. Measurements are of distinct spore types as viewed under light microscopy. The only statistically significant difference between the spores was in spore length.

	<i>Myxobolus exiguus</i>	<i>Myxobolus muelleri</i>	<i>Aurantiactinomyxon mississippiensis</i>	<i>Henneguya exilis</i>	<i>Henneguya ictaluri</i>
<i>Myxobolus procerus</i> Intracellular Cyst	276/296 (93%)	276/296 (93%)	353/392 (90%)	356/397 (89%)	356/397 (89%)
<i>Myxobolus procerus</i> Intercellular Cyst	268/287 (93%)	268/287 (93%)	355/397 (89%)	355/397 (89%)	355/397 (89%)

Table 5. BLAST results of SSU 18S rDNA consensus sequences from spores within each cyst type.

Phylum Myxozoa

Class Myxosporea

Order Bivalvulida (marine and freshwater, with two valves to spore)

Suborder Variisporina (marine and freshwater, mostly coelozoic)

Includes *Ceratomyxa*, *Chloromyxum*, *Hoferrellus*, *Myxidium*, *Myxobilatus*, *Ortholinea*, *Parvicapsula*, *Polysporoplasma*, *Sinuolinea*, *Sphaerospora*, *Zschokkella*,

Suborder Platysporina (marine and freshwater, mostly histozoic)

Includes *Myxobolus*, *Henneguya*, and *Thelohanellus*

Suborder Sphaeromyxina (marine, with ribbon-like polar filaments in polar capsules at opposing end of spore)

Sphaeromyxa

Order Multivalvulida (marine, with greater than 2 spores valves)

Includes *Hexacapsula*, *Kudoa*, *Trilospora*, and *Unicapsula*

Class Malacosporea (freshwater, with soft valves, parasites of bryozoans; one order, family and genus)

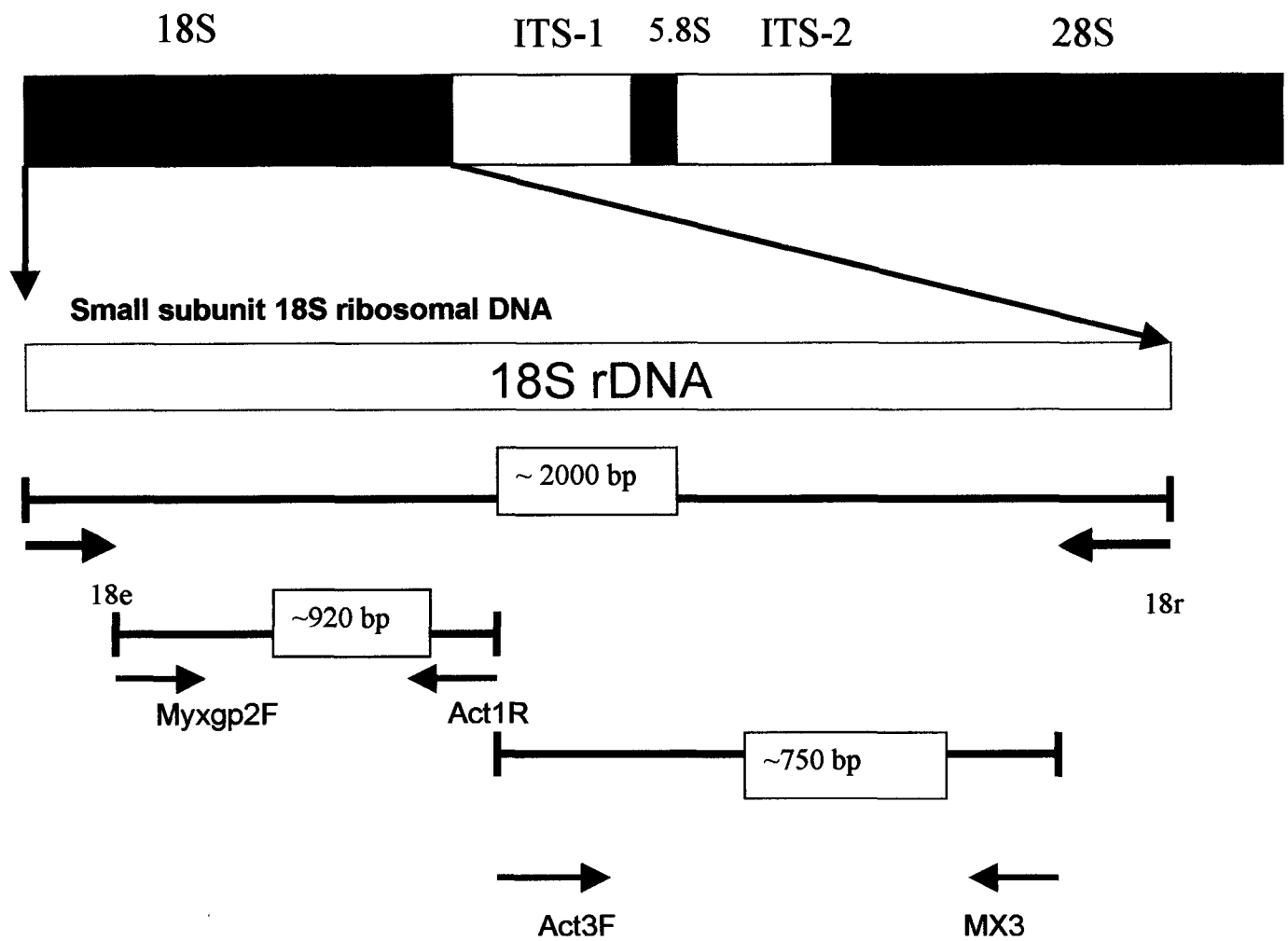
Order Malacovalvulida

Tetracapsula (with 4 polar capsules)

Table 6. Abbreviated classification of the class Myxosporea based primarily on Lom and Dyková (1992).

10. APPENDICES

Appendix A Ribosomal DNA



Appendix B

Glossary

ANOVA (Analysis of variance): Used to uncover the main and interaction effects of categorical independent variables (called "factors") on an interval dependent variable.

Algorithm: A procedure or formula for solving a problem.

Bit-score: An indication of the relatedness of the sequence in question to the sequences it is aligned against. The higher the bit score the greater the similarity.

Bootstrapping: A method in which one takes a sub sample of the sites in an alignment and creates trees based on those sub samples. That process is iterated multiple times (usually 1000) and the results are compiled to allow an estimate of the reliability of a particular grouping.

Capsulogenic: Cells that are to become the polar capsules.

Coelozoic: Cavity dwellers that attach to walls or float freely in cavities such as urinary or gall bladder. Typically invade epithelial cells in early stages.

Ectoparasitic: Parasites that live on the outside of the body of the host.

Endoparasitic: Parasites that live on the inside of the body of the host.

E-Value: The number of matches to the current nonredundant sequence database that are expected by chance alone. The smaller the E value, the more likely that the similarity is "real".

Fusiform: Spindle-shaped; tapering from the middle towards each end

Heterogeneity: Having the characteristic of containing dissimilar constituents.

Heuristic: As an adjective, heuristic pertains to the process of gaining knowledge or some desired result by intelligent guesswork rather than by following some pre-established formula.

Homogeneity: Having the characteristic of containing similar constituents.

Histozoic: Intracellular tissue dwellers, found in solid tissues such as cartilage or Muscle.

In situ Hybridization: A method of detecting the presence of specific nucleic acid sequences within a cytological preparation. A DNA or RNA probe is labeled radioactively or chemically and hybridized to a cytological preparation to detect RNA or to a denatured cytological preparation to detect DNA. The hybridization is detected by autoradiography (for radioactive probes) or by chromomeric reactions or fluorescent (for chemically-labeled probes).

Intercellular: That existing between cells.

Intracellular: That existing inside cells.

Intergenic: taking place or existing between neighboring genes.

Long-Branch Abstractions: An artifact of phylogenetic analyses of taxa with differing evolution rates.

Maximum likelihood: A method of creating phylogenetic trees by maximizing the likelihood of observing the data.

Monophyletic: Derived from a single ancestral line.

Morphology: The comparison of the forms of organisms and their parts in order to identify homologous structures.

Myxosporea: A subphylum of the Kingdom Protista, characterised by the presence of spores of multicellular origin, usually with two or three valves, two or more polar filaments, and an amoeboid sporoplasm; parasitic in lower vertebrates, especially common in fishes. Important genera include *Ceratomyxa*, *Henneguya*, *Leptotheca*, *Myxidium*, and *Myxobolus*.

Neighbour joining: A method of constructing phylogenetic trees by manipulating a distance matrix reducing it in size at each step then reconstructing the tree from that series of matrices.

Ontogeny: The development or course of development, especially that of an individual organism.

p Value: The probability of getting something more extreme than your result, when there is no effect in the population.

Parsimony: In effect, this rule states that if there exists two answers to a problem or a question, and if, for one answer to be true, well-established laws of logic and science must be re-written, ignored, or suspended in order to allow it to be true, and for the other answer to be true no such accommodation need be made, then the simpler-the second-of the two answers is much more likely to be correct.

Phylogeny: The evolution of a genetically related group of organisms as distinguished from the development of the individual organism.

Polar capsule: Apical, thick walled vesicle of a myxozoan spore containing spirally coiled, extrusible polar filament.

Polar filament: Tube-like structure coiled within the polar capsule of myxozoans. Serves as anchor for hatching spore (cf. polar tube of microsporeans).

Polymerase: Any enzyme which catalyses the formation of a polymer, esp. a polynucleotide. For example the enzyme responsible for deoxyribonucleotide incorporation is designated as 'polymerase'.

Polymerase Chain Reaction (PCR): An in vitro DNA synthesis procedure in which two oligonucleotides annealed to a DNA template oriented so that the primers direct DNA synthesis on complementary, opposing strands towards each other. With successive cycles of DNA synthesis, denaturation, and repriming, the region between the primers accumulates exponentially as a double-stranded DNA fragments with the ends consisting of the oligonucleotide primers.

Polyphyletic: Composed of members that originated, independently, from more than one evolutionary line.

Pseudoplasmodium: A plasmodium with vegetative and generative nuclei in the same cell.

Restriction Fragment Length Polymorphism: Restriction fragment length polymorphism is the identification of specific restriction enzymes that reveal a pattern difference between the DNA fragment sizes in individual organisms.

Ribosomal RNA: Ribosomes are the site of protein synthesis in the cell. The sizes of various subunits of the ribosome are given in Svedberg units. In all organisms, the ribosomes are made up of two subunits of unequal size. In eukaryotes, there are 4 segments of rRNAs (18S piece in the smaller subunit and 5.8S and 28S in the larger subunit).

Taxon: A taxonomic group, as a genus or species.

Topology: The way in which constituent parts are interrelated or arranged.

Svedberg Unit (ex.18S): The Svedberg is a unit that measures the rate at which a particle sediments in a sucrose solution during ultra centrifugation..

Taq Polymerase - A DNA polymerase that is very stable at high temperatures, isolated from the thermophilic bacterium *Thermus aquaticus*. Very useful in PCR

reactions which must cycle repetitively through high temperatures during the denaturation step.

Triploblast: An organism having three primary germ layers

Vector: A bacteriophage, phage or plasmid that transfers genetic material from one bacterium to another; also, a used to transfer extraneous DNA into a cell.

X-Gal: Chromogenic substrate for the enzyme beta-galactosidase yielding an intense blue colour. Very widely used in blue-white colour selection of bacteria containing recombinant plasmids, and in histochemistry of tissues or cells expressing beta-galactosidase as a reporter gene.

Appendix C List Of Reagents

DNA Extraction

- i. Proteinase K (200µg/ml)
- ii. Sodium Dodecyl Sulfate (10%)
- iii. Sodium Acetate 3 M
- iv. Phenol:Chloroform:Isoamyl alcohol 25:24:1
- v. TE (Tris 10mM – EDTA 1 mM pH 8.0)
- vi. 100% Ethanol
- vii. 70% Ethanol

Polymerase Chain Reaction – 50 µL reaction

- i. *Taq* Polymerase (1.25U)
- ii. Magnesium buffer (1.5mM)
- iii. 1 X PCR buffer
- iv. dNTP (250 µM each)
- v. Primers 1 µM each
- vi. ddH₂O to 50 µl
- vii. DNA to 300 ng

Gene Clean

- i. Sodium iodide
- ii. Guanidium hydrochloride
- iii. TE (10mM Tris-1mM EDTA pH 8.0)
- iv. Glass milk

Cloning

- i. X-galactosidase (X-gal)
- ii. LB (Luria-Bertani) Broth:
 - a. 1% tryptone
 - b. 0.5% Yeast extract
 - c. 1% NaCl
 - d. pH 7.0
- iii. Carbenicillin (final concentration at 25 µg/ml)

DNA Sequencing

- i. ET dye terminator
- ii. PCR primers (see Materials and Methods)

Appendix D TOPO Cloning Vector

Sequence for TOPO Cloning Vector

AGCGCCCAATACGCAAACCGCCTCTCCCCGCGCGTTGGCCGATTCATTAATGCAGCTGGCA
 CGACAGGTTTCCCGACTGGAAAGCGGGCAGTGAGCGCAACGCAATTAATGTGAGTTAGCTC
 ACTCATTAGGCACCCCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAATTGT
 GAGCGGATAACAATTTACACACAGGAAACAGCTATGACCATGATTACGCCAAGCTATTTAGGT
 GACGCGTTAGAATACTCAAGCTATGCATCAAGCTTGGTACCGAGCTCGGATCCACTAGTAAC
 GGCCGCCAGTGTGCTGGAATTCGCCCTTAAGGGCGAATTCTGCAGATATCCATCACACTGG
 CGGCCGCTCGAGCATGCATCTAGAGGGCCCAATTCGCCCTATAGTGAGTCGTATTACAATTC
 ACTGGCCGCTCGTTTTACAACGTCGTGACTGGGAAAACCCCTGGCGTTACCCAACCTTAATCGCC
 TTGCAGCACATCCCCCTTTCGCCAGCTGGCGTAATAGCGAAGAGGGCCCGCACCGATCGCCC
 TTCCCAACAGTTGCGCAGCCTATACGTACGGCAGTTTAAAGTTTACACCTATAAAAGAGAGA
 GCCGTTATCGTCTGTTTGTGGATGTACAGAGTGATATTATTGACACGCCGGGGCGACGGAT
 GGTGATCCCCCTGGCCAGTGCACGTCTGCTGTCAGATAAAGTCTCCCGTGAACCTTACCCG
 GTGGTGCATATCGGGGATGAAAGCTGGCGCATGATGACCACCGATATGGCCAGTGTGCCG
 GTCTCCGTTATCGGGGAAGAAGTGGCTGATCTCAGCCACCGCGAAAATGACATCAAAAACG
 CCATTAACCTGATGTTCTGGGGAATATAAATGTCAGGCATGAGATTATCAAAAAGGATCTTCA
 CCTAGATCCTTTTACGTAGAAAGCCAGTCCGCAGAAACGGTGCTGACCCCGGATGAATGT
 CAGCTACTGGGCTATCTGGACAAGGGAAAACGCAAGCGCAAAGAGAAAGCAGGTAGCTTGC
 AGTGGGCTTACATGGCGATAGCTAGACTGGGCGGTTTTATGGACAGCAAGCGAACCAGGAAT
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 CTCGCCGCCAAGGATCTGATGGCGCAGGGGATCAAGCTCTGATCAAGAGACAGGATGAGG
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 TGAAGTCAAGACGAGGCAGCGCGGCTATCGTGGCTGGCCACGACGGGCGTTCCTTGCGC
 AGCTGTGCTCGACGTTGTCACTGAAGCGGGAAGGGACTGGCTGCTATTGGGCGAAGTGCC
 GGGGCAGGATCTCCTGTCATCTCACCTTGCTCCTGCCGAGAAAGTATCCATCATGGCTGAT
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 CGAAGAGCATCAGGGGCTCGCGCCAGCCGAACTGTTCCGCCAGGCTCAAGGCGAGCATGCC
 CGACGGCGAGGATCTCGTCTGACCCATGGCGATGCCTGCTTGCCGAATATCATGGTGGAA
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 CGGGACTTCGTGGAGGACGACTTCGCCGGTGTGGTCCGGGACGACGTGACCCTGTTTCATC
 AGCGCGGTCCAGGACCAGGTGGTGCCGGACAACACCCTGGCCTGGGTGTGGGTGCGCGG
 CCTGGACGAGCTGTACGCCGAGTGGTCCGGAGGTGCTGTCCACGAACCTTCCGGGACGCCTC
 CGGGCCCGGCCATGACCGAGATCGGCGAGCAGCCGTGGGGGCGGGAGTTCCGCCCTGCGCG
 ACCCGGCCGGCAACTGCGTGCACTTCGTGGCCGAGGAGCAGGACTGACACGTGCTAAAAC
 TTCATTTTTAATTTAAAAGGATCTAGGTGAAGATCCTTTTTGATAATCTCATGACCAAAATCCC
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 GTGGTTTGTGGCCGATCAAGAGCTACCAACTCTTTTTCCGAAGGTAAGTGGCTTCAGCAG
 AGCGCAGATACCAATACTGTCCTTCTAGTGTAGCCGTAGTTAGGCCACCACTTCAAGAACT
 CTGTAGCACCGCCTACATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGTGG

CGATAAGTCGTGTCTTACCGGGTTGGAAGACGATAGTTACCGGATAAGGCGCAGCGG
 TCGGGCTGAACGGGGGGTTCGTGCACACAGCCCAGCTTGGAGCGAACGACCTACACCGAA
 CTGAGATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTTCCCGAAGGGAGAAAGGCG
 GACAGGTATCCGGTAAGCGGCAGGGTCGGAACAGGAGAGCGCACGAGGGAGCTTCCAGG
 GGGAAACGCCTGGTATCTTTATAGTCCTGTCTGGGTTTCGCCACCTCTGACTTGAGCGTCGAT
 TTTTGTGATGCTCGTCAGGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCGGCCTTTTT
 ACGGTTCTGGGCTTTTGCTGGCCTTTTGCTCACATGTTCTTTCTGCGTTATCCCCTGATTC
 TGTGGATAACCGTATTACCGCCTTTGAGTGAGCTGATACCGCTCGCCGCAGCCGAACGACC
 GAGCGCAGCGAGTCAGTGAGCGAGGAAGCGGAAG

The TOPO TA Cloning System

The TOPO TA Cloning System (TOPO Cloning) is a method for cloning polymerase chain reaction (PCR) products into a plasmid vector

PCR is a method, which essentially isolates a specific segment of DNA by amplification. PCR uses specific primers, which allow *Taq* DNA polymerase to make copies of only a specific segment. *Taq* DNA polymerase is used for its ability to withstand the high heat (95°C) necessary for the many cycles of PCR. A unique aspect of *Taq* DNA polymerase is that it adds a single deoxyadenosine (A) to the 3' ends of PCR products. The resulting products of PCR are many copies of a specific DNA sequence with 3' A overhangs.

TOPO Cloning ligates the PCR product into the pCR 2.1-TOPO plasmid vector (vector) with topoisomerase I. This can be done quickly and efficiently because of the unique aspects of the vector. The vector has been engineered to be a linearized plasmid with 3' deoxythymidine (T) overhangs that is activated by being covalently bonded to topoisomerase I. The 3' A overhangs of the PCR product complement the 3' T overhangs of the vector and allow for fast ligation with the already present topoisomerase I. The plasmid can then be transformed into competent bacterial cells.

Other useful features of the vector are its ampicillin and kanamycin resistance markers, lacZ reporter gene, T7 promoter, EcoR I sites flanking the PCR insertion site, and the f1 origin of replication. The ampicillin and kanamycin resistance inserts in the plasmid allow for quick selection of bacterial colonies that take up the vector plasmid during transformation. The lacZ gene in bacteria causes colonies to have a blue color. If PCR DNA is inserted in the vector it will insert in the middle of the lacZ gene causing the colonies to be white and easily selected. The T7 promoter region in the vector allows for *in vitro* RNA transcription/translation by the T7 phage. If T7 phage infects the bacteria it will make proteins from the DNA sequence of the vector and insert which allows scientists to sequence the subsequent protein and therefore DNA sequence of the insert. The *EcoR* I sites on either side of the insert enable the insert to be easily removed by *EcoR* I restriction enzymes. The f1 origin of replication is a necessary component of the plasmid and makes single-strand rescue possible.

The entire TOPO Cloning process can be done in only five minutes at room temperature with 95% efficiency. This is much faster and less complicated than the PRIME PCR Cloner Cloning System we used in lab and most other cloning

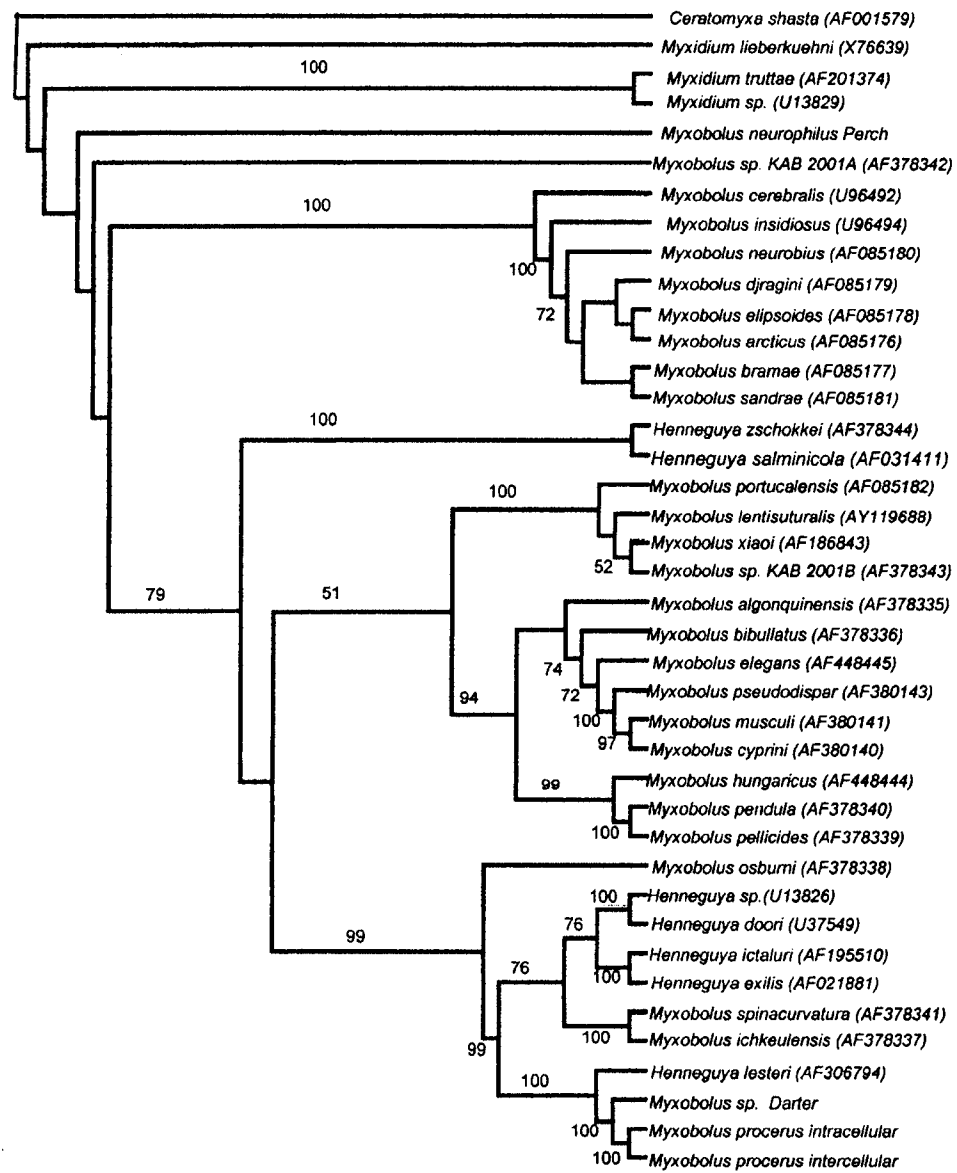
procedures. TOPO Cloning system is ideal for scientists that need to quickly clone PCR products into bacteria.

Reference:

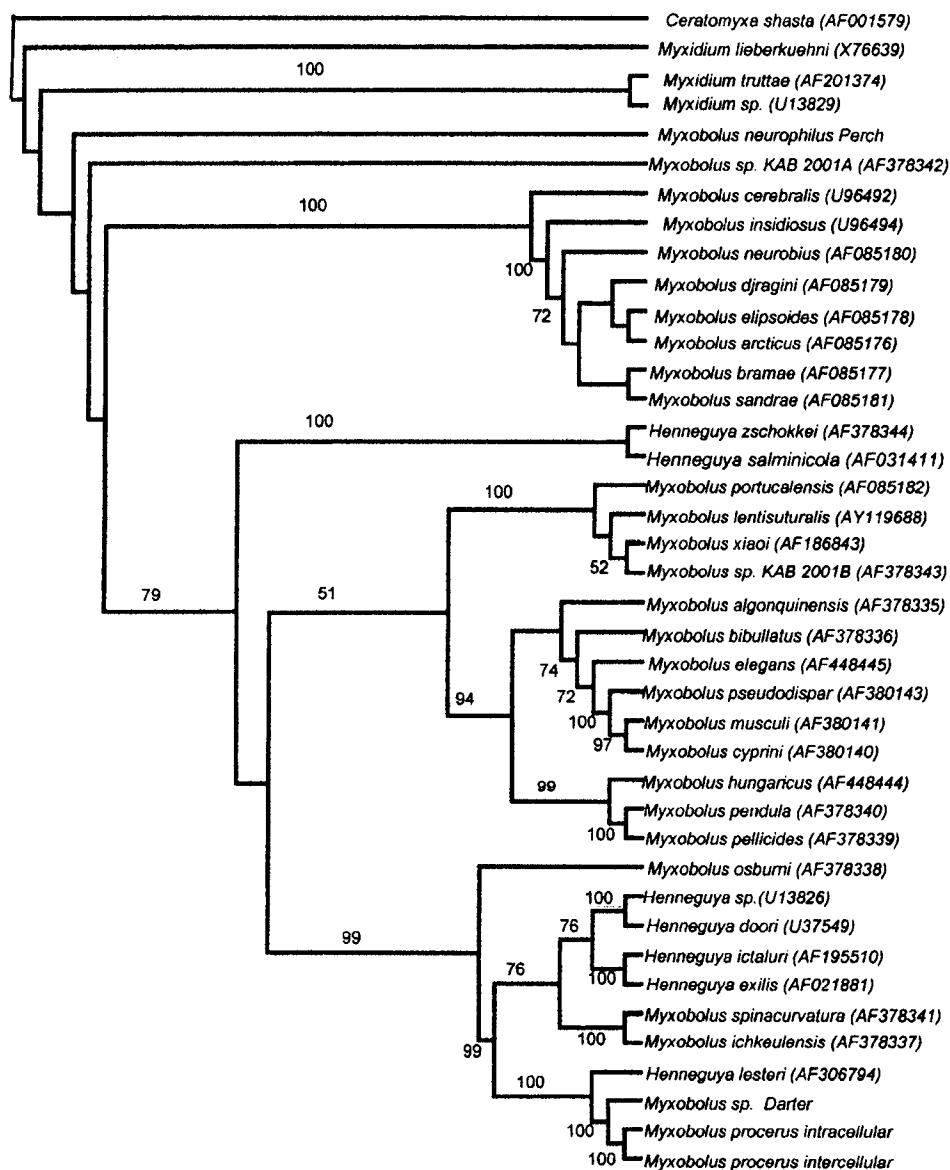
Campbell NA, Biology. 4th ed. Menlo Park, CA: The Benjamin/Cummings Publishing Company, Inc.; 1996. p 379-380.

Hoover C, 1998 July 8. Invitrogen web catalog.

Appendix E Likelihood tree



Appendix F Parsimony Tree



Appendix G Information on *Myxobolus* Sp. used in phylogenetic tree

1. *Myxidium* species

Myxidium truttae GenBank AF201374 *Myxidium* sp. GenBank U13829, *Myxidium lieberkuehni* GenBank X766390,

Host: *Perca flavescens*, *Melanogrammus aeglefinus*, *Oncorhynchus*

Site: kidney, intestine, urinary bladder, gall bladder.

Distribution: AB, BC, ON

2. *Henneguya* species

Henneguya sp. GenBank U13826,

Henneguya lesteri GenBank AF306794,

Henneguya doori GenBank U37549,

Henneguya ictaluri GenBank AF195510,

Host: *Coregonus*, *Luxilis*, *Oncorhynchus*

Site: gills or not specified

Distribution: AB, ON

3. *Henneguya salminicola* GenBank AF031411

Host: *Oncorhynchus* sp.

Site: muscle cysts

Distribution: BC

4. *Henneguya exilis* GenBank AF021881

Host: *Ameiurus*, *Ictalurus*

Site: gills

Distribution: ON

5. *Henneguya zschokkei* GenBank AF378344

Host: *Coregonus*, *Cottus*, *Prosopium*, *Rhinichthys*, *Richarsonius*, *Thymallus*

Site: muscle, gills, kidney

Distribution: BC, ON

6. *Myxobolus osburni* GenBank AF378338

Host: *Lepomis gibbosus*

Site: mesenteries

Distribution: ON

7. *Myxobolus neurobius* GenBank AF085180

Host: *Oncorhynchus*, *Prosopium*, *Salmo*, *Salvelinus*

Site: brain, spinal cord

Distribution: BC, NF

8. *Myxobolus arcticus* GenBank AF085176

Host: *Oncorhynchus* sp.

Site: brain

Distribution: BC

9. *Myxobolus musculi* GenBank AF380141

Host: *Notemigonus crysoleucas*

Site: eye, heart, kidney, muscle, ureters, urinary bladder

Distribution: ON

10. *Myxobolus pendula* GenBank AF378340

Host: *Semotilus atromaculatus*, *Sematolilus margarita*

Site: gills

Distribution: ON

11. *Myxobolus pellicides* GenBank AF378339

Host: *Semotilus atromaculatus*

Site: gills

Distribution: ON

12. *Myxobolus bibullatus* GenBank AF378336

Host: *Catostomus*, *Cottus*, *Mylocheilus*, *Ptychocheilus*

Site: gall bladder, muscle, kidney

Distribution: BC, NS , ON

13. *Myxobolus* sp.

Myxobolus lentisuturalis GenBank AY119688, *Myxobolus* KAB-2001B GenBank AF378343, *Myxobolus xiaoi* GenBank AF186843, *Myxobolus portucalensis* GenBank F085182, *Myxobolus cerebrealis* GenBank U96492, *Myxobolus insidiosus* GenBank U96494, *Myxobolus elipsoides* GenBank AF085178, *Myxobolus dfragini* GenBank AF085179, *Myxobolus bramae* GenBank AF085177, *Myxobolus sandrae* GenBank AF085181, *Myxobolus* sp. KAB2001A GenBank AF378342, *Myxobolus pseudodispar* GenBank AF380143, *Myxobolus cyprini* GenBank AF380140, *Myxobolus hungaricus* GenBank AF448444, *Myxobolus elegans* GenBank AF448445, *Myxobolus algonquinensis* GenBank AF378335

Host: All of the above

Site: gills, fins, gall bladder, intestine, heart, kidney, liver, brain, spinal cord, gonads

Distribution: BC, MB, NB, NS, ON

14. *Ceratomyxa shasta* GenBank AF001579

Host: *Oncorhynchus* sp.

Site: pyloric caeca, intestine, gall bladder

Distribution: BC, YT

15. *Myxobolus procerus*

Host: *Percopsis omiscomaycus*

Site: skin, muscle

Distribution: ON

16. *Myxobolus ichkeulensis* GenBank AF378337

Host: ND

Site: gill arches

Distribution: Mugil, Mediterranean Sea: off coast of Tunisia

17. *Myxobolus spinacurvatura* GenBank F378341

Host: ND

Site: mesenteries and blood

Distribution: Mugil, Mediterranean Sea: off coast of Tunisia

Appendix H Clustal X alignment of species of *Myxobolus*.

```

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interleave datatype=DNA gap= -;

matrix
Melipsoides -----
Mdjragini -----
Mbramae -----
Mneurobius -----
Marcticus -----
Msandrae -----
Minsidiosus -----
Mcerebralis -----AGATATACGCTTTTCTCTAAGACTAAGC
Mlentisuturalis -ACCTGGTTGATCCTGCCAGTGAGCGTACGCTTTTCTCAAGGACTCAGCC
Mxiao -----GAGCGTACGCTTTTCTCAAGGACTCAGCC
Myxo. -----
Mportucalensis -----
Mmusculi -----
Mcyprini -----
Mpseudodispar -----
Mbibullatus -----AGAATCATGCTATTCTTGAAGATTAAGCC
Melegans -----
Mpendula -----CCCTTTGGGAAGAAACATGCTATTCTCAAAGACTAAGCC
Mpellicides -----CCCTTTGGGAAGAAA-ATGCTATTCTCAAAGACTAAGCC
Mhungaricus -----
Myxiditruttae -----
Myxidium -----TGGCATATGCTCGTCTCTAAGATT-AAGC
Ethclone1AM -----
Ethclone3AM -----
Ethclone2AM -----
Mdiaphanus -----
M.lieberkuehni AACCTGGTTGATCCTGCCAGTAATCATATGCTTGTCTCTAAGCTT-AAGC
Myxosp. -----
Hennzschokkei -----AAACATACGCTGTTTTCTAAGACT-AAGC
Hennsalminicola -----AAACATACGCTGTTTTCTAAGACTTAAGC
Ethclone2MA -----
Ethclone3MA -----
Ethclone1MA -----
Myxobprocerus -----
Mprocerus -----
Henneguya -----TTCCGTATTCTGCCAGTGGGCATACGCTCTTTTTTAAAGATTTAGC
Hsp. -----GCGTATACGCTCTTTTTTAAAGATTAAGC
Hdoori -----ATGC-----
Hennictaluri -AACCTGGTTGATCCTGCCAGTGCGTATACGCTCTTTTTTAAAGACTAAGC
Hennexilis -AACCTGGTTGATCCTGCCAGTGCGTATACGCTCTTTTTTAAAGATTAAGC
Mspinacurvatura -----
Michkeulensis -----
Mosburni -----ACGCTGTTCTTAAAGATTTAGC

```

Muvuliferus	-----
Ceratomyxa	-----
Malgonquinensis	-----AAGATTAAGC
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	-----GYCGGCTC
Mcerebralis	CATGCATGTTNAAGTTCATACGTAGTAAAACGTGAGACTGCGGACGGCTC
Mlentisuturalis	ATGCAAGTGTTAAAGTTCATACGATTTAAAAGTGAGACTGCGGACGGCTC
Mxiao	ATGCAAGTGTTAAAGTTCATACGATTTAAAAGTGAGACTGCGGACGGCTC
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-TGTAGGTGCCAA-GTTCATACAATATTATTGTGAGACTGCGGACGGCTC
Melegans	-----
Mpendula	ATGCACGTGCCAA-GTTCATACGATCTTATCGTGAGACTGCGGACGGCTC
Mpellicides	-TGCACGTGCCAA-GTTCATACGATCTTATCGTGAGACTGCGGACGGCTC
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	CATGCATGTCTGAGTTCATACCTAGTTAA-CGTGAGACTGCGAAAGGCTC
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	CATGCATGTCTAAGTTCAAACAT--TTAA-TGTGAGACTGCGAAAGGCTC
Myxosp.	-----
Hennzschokkei	CATGCAGGTGCTACAGTACATACGTGAATTTCGTGAGACTGCGGAAGGCTC
Hennsalminicola	CATGCAGGTGCTACACTACATACGTGAATTTCGTGAGACTGCGGAAGGCTC
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----CTGCGGACGGCTC
Mprocerus	-----CTGCGGACGGCTC
Henneguya	CATGCATGTGCCAG-TTCATACATTAAAAATGTGAGACTGCGGACGGCTC
Hsp.	CATGCAGGTGCTAG-TTCATACATATGTAATGTGAGACTGCGGACGGCTC
Hdoori	----CAGGTGCTAGCTTCATACATATGTAATGTGAGACTGCGGACGGCTC
Hennictaluri	CATGCATGTGCCAG-TTCATACATATTTAATGTGAGACTGCGGACGGCTC
Hennexilis	CATGCAAGTGCTAG-TTCATACATATTTAATGTGAGACTGCGGACGGCTC
Mspinacurvatura	-----
Michkeulensis	-----
Mosburni	CATGCAAGTGAAAG-TTCAAACATTTAAAAATGTGAGACTGTGGACGNCTC
Muvuliferus	-----
Ceratomyxa	-----TTAAATGATGAAACTGCGAAGGCGTC
Malgonquinensis	CATGTATGTGCCAAGTTCATACG--TTAGACGTGAGACTGCGGACGGCTC
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----

Melipsoides	-----TATCATCTATTTGATTGTCTA--CCCA--TTGGATAACCG
Mdjragini	-----TATCATCTATTTGNTTGTCTA--CCCA--TTGGATAACCG
Mbramae	-----TATCATCTATTTGATTGTCTA--CCCA--TTGGATCCCCG
Mneurobius	-----TATCATCTATTTGNTTGTCTA--CCCA--TTGGATAACCG
Marcticus	-----TATCATCTATTTGATTGTCTA--CCCA--TTGGATAACCG
Msandrae	-----TATCATCTATTTGATTGTCTA--CCCA--TTGGATAACCG
Minsidiosus	AGTAAATCAGTYATCATCTATTTGATTGTCTA--CCCA--TTGGATAACCG
Mcerebralis	AGTAAATCAGTTATCATCTATTTGATTGTCTA--CCCA--TTGGATAACCN
Mlentisuturalis	AGTAAATCAGCAATGATCTATCTGATTGTAGAGACCCA--TTGGATAACCG
Mxiao	AGTAAATCAGCGATGATCTATCTGATTGTACAT--CCCA--TTGGATAACCG
Myxo.	-----GATGATCTATCTGATTGTACAT--CCCA--TTGGATAACCG
Mportucalensis	-----CATGATCTATCTGATTGTATAG--CCCA--TTGGATAACCG
Mmusculi	-----GAT-ATCTATTTGATTGTCTAA--CCTA--TTGGATAACCG
Mcyprini	-----GAT-ATCTGTTTGGATTGTCTAA--CCTA--TTGGATAACCG
Mpseudodispar	-----GAT-ATCTGCTGATTGTCTAT--CCTA--TTGGATAACCG
Mbibullatus	AGTAAATCAGTGATTATCTGTTTGGATTGTATAA--CCCCATTGGATAACCG
Melegans	-----GAT-ATCTATTTGATTGTCTA--CCCA--TTGGATAACCG
Mpendula	AGTATATCAGTGATTATCTGTTTGGATTGTCTTA--CCCA--TTGGATAACCG
Mpellicides	AGTATATCAGTGATTATCTGTTTGGATTGTCTTA--CCCA--TTGGATAACCG
Mhungaricus	-----GAT-ATCTGTTTGGATTCTCTTC--GCCA--TTGGATAACCG
Myxiditruttae	-----
Myxidium	AGTACATCAGTTATAGTTTATTCGATTGT--GAATCCAC--ATGGATAACCG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	AGTAAATCAGCTATAATCTATTTGATTGTAACTCCCA--GTGGATAACCG
Myxosp.	-----CTATTTGATTGT--ACAGCCCA--ATGGATAACCG
Hennzschokkei	AGTAAATCAGTTATCATCTATTTGATTGT--CTAGCCCA--TTGGATAACCG
Hennsalminicola	AGTAAATCAGTTATCATCTATTTGATTGT--CTAGCCCA--TTGGATAACCG
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	AGTATATCAGTTATAATCTGCTCGATTGTTATGGTTAT--TGGATAACCG
Mprocerus	AGTAAATCAGTTATAATCTGCTCGATTGTTATGGTTAT--TGGATAACCG
Henneguaya	AGTATATCAGTTATAATCTGCTCGATTGTTAAGGTTAT--TGGATAACCG
Hsp.	AGTATATCAGTTATAATCTGCTCGATTGTTCAAGGTTAT--TGGATAACCG
Hdoori	AGTATATCAGTTATAATCTGCTCGATTGTTGAGGTTAG--TGGATAACCG
Hennictaluri	AGTATATCAGTTATAATCTGCTCGATTGTTAAGGTTAT--TGGATAACCG
Hennexilis	AGTATATCAGTTATAATCTGCTCGATTGTTAAGGTTAT--TGGATAACCG
Mspinacurvatura	-----
Michkeulensis	-----TATAATCTGCTCGATTGTATAGGTTAT--TGGATAACCG
Mosburni	AGTAAATCAGTTATAATCTGCTCGATTGTCAAGGTTAT--TGGATAACCG
Muvuliferus	-----
Ceratomyxa	AGTAAATCAGTTATCGTCTGTTTCGATCGATACATGCCA--TGGATAACTG
Malgonquinensis	AGTAAATCAGTGACAATCTATTTGATTTTCTCT--CCCA--TTGGATAACCG
Perchclone1AM	-----CTAGCCCATGCAATGTTTGACTG-----G
Perchclone2AM	-----CTAGCCCATGCAATGTTTGACTG-----G
Perchclone3AM	-----CTAGCCCATGCAATGTTTGACTG-----G
Melipsoides	TGGGNAA-TCTAGNGCTAATAC--ATGCAG-----TTTGTAGTT
Mdjragini	TGGGNAA-TCTAGNNCTAATAC--ATGCAG-----TTTGNANT
Mbramae	TGGGAAA-TCTAGANNNTTTC--NTNCAG-----TTTGTAGAT
Mneurobius	TGGGNAA-TCTAGNGCTAATAC--ATGCAG-----TTTGTAGNT
Marcticus	TGGGAAA-TCTAGAGCTAATAC--ATGCAG-----TTTGTAGAT
Msandrae	TGGGAAA-TCTAGAGCTAATAC--ATGCAG-----TTTGTAGAT
Minsidiosus	TGGGAAA-TCTAGAGCTAATAC--ATGCAG-----TCTTGGGAT

Mcerebralis	NNGGGAAATCTAGAGCTAATAC--ATGCAG-----TTTTGGG-C
Mlentisuturalis	TGGGAAA-TCTAGAGCTAATAC--ATGCGTTGAAATTACG----TGG---
Mxiao	TGGGAAA-TCTAGAGCTAATAC--ATGCGTTGATATAACGATATTGGAGT
Myxo.	TGGGAAA-TCTAGAGCTAATAC--ATGCGTTGATATAAATGTATTTGGAG-
Mportucalensis	TGGGAAA-TCTAGAGCTAATAC--ATGCGTTTA-ATCGTACAGGTAGCT-
Mmusculi	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----GTTTATGG---
Mcyprini	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----GTAAATGG---
Mpseudodispar	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----GTTTATGG---
Mbibullatus	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----GTTTATGG---
Melegans	TGGGAAA-TCTAGAGCTAATAC--ATGCAA-----GTATTTGGCAG
Mpendula	TGGGAAA-TCTAGAGCTAATAC--ATGCAA-----ATAATTGGCGT
Mpellicides	TGGGAAA-TCTAGAGCTAATAC--ATGCAA-----ATAATTGGCGT
Mhungaricus	TGGGAAA-TCTAGAGCTAATAC--ATGCAG-----TTTATTGGTCT
Myxiditruttae	-----
Myxidium	TGG-AAA-TCTAGAGCTAATAC--ATGCGAAAAAT---CTTTCAAGGTG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	TGGGAAA-TCTAGAGCTAATAC--ATGCA-A-----CTAAGCCGTGC
Myxosp.	TGGGAAA-TCTAGAGCTAATAC--ATGCACA-----CTTATTTGAAA
Hennzschokkei	TGGGAAA-TCTAGAGCTAATAC--ATGTAGTTAA-----TTGGTGGTGT
Hennsalminicola	TGGGAAA-TCTAGAGCTAATAC--ATGTAGTTAA-----TTGGTGGTGT
Ethclone2MA	-----CTA-----ACATGCTGGATG
Ethclone3MA	-----CTA-----ACATGCTGGATG
Ethclone1MA	-----
Myxobprocerus	TGGGAAA-TCTAGAGCTAATAC--ATGCTA-----TAATGTTGGGTG
Mprocerus	TGGGAAA-TCTAGAGCTAATAC--ATGCTA-----TAATGTTGGGTG
Henneguya	TGGGAAA-TCTAGAGCTAATAC--ATGC-A-----GTTTGCGGTGGT
Hsp.	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----CGCCTCGGTT-A
Hdoori	TGGGAAA-TCTAGAGCTAATAC--ATGCAA-----CGCCTCAGTTTG
Hennictaluri	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----CCTCTCGGCGAA
Hennexilis	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----CCTCTCGTCCGG
Mspinacurvatura	-----CTAATAC--ATGTA-----ATTTGGGGA
Michkeulensis	TGGGAAA-TCTAGAGCTAATAC--ATGTA-----ATT-GGGAG
Mosburni	TGGGAAA-TCTAGAGCTAATAC--ATGCA-----GTTTGTGAGGG
Muvuliferus	-----
Ceratomyxa	TGGCAAA-CCTAGAGCTAATAC--ATGCAAA-----ATTCTTTGTTTT
Malgonquinensis	TGGGAAA-TCTAAAGCTAATAC--GTGCGG-----TATCTTGGCG
Perchclone1AM	AGGGAAAGTGCAACTTCAACACCCATCCAC-----
Perchclone2AM	AGGGAAAGTGCAACTTCAACACCCATCCAC-----
Perchclone3AM	AGGGAAAGTGCAACTTCAACACCCATCCAC-----
Melipsoides	AG-----CGT-AA-GTTGTC-----TCACGGCAT-TTA-TTGG-
Mdjrugini	AG-----CGT-AA-GTTGTC-----TCACNGCAT-TTA-TTGGG
Mbramae	AG-----CGT-AA-GTTGTC-----TCACGGCAT-TTA-TTGG-
Mneurobius	AG-----CGT-AA-GTTGTC-----TCACGGCAT-TTA-TTGG-
Marcticus	AG-----CGT-NAAGTTGTC-----TCACGTCAT-TTA-TTGG-
Msandrae	AG-----CGT-AA-GTTGTC-----TCACGGCAT-TTA-TTGG-
Minsidiosus	AG-----CGT-AA-GTTGTC-----TCACGGCAT-TTA-TTGG-
Mcerebralis	AG-----CGTTAAACTGTC-----TCACGGCAT-TTA-TTGG-
Mlentisuturalis	TG---GGTAA--AATCAT-CA-----CGTAAAGCAT-TTA-TTGG-
Mxiao	TG---TGTTAGCAATAATGCAATGAATTGTCGTTAAGCAT-TTA-TTAG-
Myxo.	-G---CATTTGTAAGATGTACTTGAATA-CATTAAAGCAT-TTA-TTAG-
Mportucalensis	TG---CGCTGCC-----TGT-----TGAAGCAT-TTA-TTAG-
Mmusculi	-----TCG---GGCTTGCT-----CGGCCAAGCAT-TTA-TTAG-
Mcyprini	-----TCG---GGCTTGCT-----CGGCCAAGCAT-TTA-TTAG-

Mpseudodispar	-----TCG---GGCTTGCT-----TGGCCAAGCAT-TTA-TTAG-
Mbibullatus	CAAAATCTTTG---GGTTTG-----TGTCAAAGCAT-TTA-TTAG-
Melegans	TA--CCCTCG---GG--TGC-----TGTCAAAGCAT-TTA-TTAG-
Mpendula	TG-----CTGCAAGGCGCG-----CCAAAGCAT-TTA-TTAG-
Mpellicides	TG-----CTGCAAGGCGCG-----CCAAAGCAT-TTA-TTAG-
Mhungaricus	TG-----TCGAAAGGCTTGA-----TCAAAGCAT-TTA-TTAG-
Myxiditruttae	-----ATC---ATTTTC-----AGGGTGATGCAT-TTA-TTAG-
Myxidium	AG-----CATC---ATTTTG-----AAGGTGATGCAT-TTAATTAG-
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	CC-----TGTC---AAAGGG-----GTGCGGTAGCAT-TTA-TTGG-
Myxosp.	GG-----CGTC---AAAACC-----TATCAAGGCAT-TTA-TTAG-
Hennzschokkei	AG-----TATTTATATTGTA-----CCACCAATGCAT-TTA-TTAG-
Hennsalminicola	AG-----TATTTATATTGTA-----CCACCAATGCAT-TTA-TTAG-
Ethclone2MA	TA-----GCTTGCTACATTT-----GGCC---GCAT-TTA-TTAGA
Ethclone3MA	TA-----GCTTGCTACATTT-----GGCC---GCAT-TTA-TTAGA
Ethclone1MA	-----
Myxobprocerus	TG-----GTTCGCTGCATTC-----AGCC---GCAT-TTA-TTAGA
Mprocerus	TG-----GTTCGCTGCATTC-----AGCC---GCAT-TTA-TTAGA
Henneguya	TG-----CTTTCGGGCGACT-----GGCGT-GGCAT-TTA-TTAGA
Hsp.	CG-----GTTCC-GCCGTGA-----ACCGAATGCAT-TTA-TTAGA
Hdoori	CG-----GTTCC-GCCGCGA-----GCTGAATGCAT-TTA-TTAGA
Hennictaluri	AA-----GCTT---GCTTTTC-----GTCGATTGCAT-TTA-TTAGA
Hennexilis	AA-----GCTT---GCTTTTC-----GTCGATTGCAT-TTA-TTAGA
Mspinacurvatura	TG-----ATTTTCGGTTCGTT-----CTCGTGGCAT-TTA-TTAGA
Michkeulensis	CG-----GTTTCGGCCGT-----CTCATGGCAT-TTA-TTAGA
Mosburni	CA-----AC-----CTCATGGCAT-TTA-TTAGA
Muvuliferus	-----
Ceratomyxa	CG-----GCAAA-----GAAGCAT-TTATCCAG-
Malgonquinensis	TA-----TCTTCGGGTACG-----TCAAAGCAT-TTA-TTAG-
Perchclone1AM	-----ACCAAGCATGTCA--TAA-
Perchclone2AM	-----ACCAAGCATGTCA--TAA-
Perchclone3AM	-----ACCAAGCATGTCA--TAA-
Melipsoides	ACAAAACCAACTAC-CGA-CGTANCAGCTT--GCTGTTGCGACGCGTA--
Mdjragini	ACAAAACCAACTAC-CGA-CGTANCAGCTT--GCTGTTGCGTTCGCGTA--
Mbramae	ACAAAACCAACTAC-CGA-CGTANCAGCTT--GCTGTTGCGACGCGTA--
Mneurobius	ACAAAACCAACTAC-CGA-CGTANCAGCTT--GCTGTTGCGACGCGTA--
Marcticus	ACAAAACCAACTAC-CGA-CGTACCAGCTT--GCTGTTGCAACCCGTA--
Msandrae	ACAAAACCAACTAC-CGA-CGTAGCAGCTT--GCTGTTGCGACGCGTA--
Minsidiosus	ACAAAACCYACTAC-CGA-CGTAGCAGTTT--GCTGTTGCGACGCGTN--
Mcerebralis	ACTAAACCAACTAC-CGT-TGCATTGGTTTACGCTGATGTAGCGAGTA--
Mlentisuturalis	TAGAAACCAACTGT-TGG-TGCATGTAACAGTGTG---CCA---ATAT-
Mxiao	AATAAACCAACTGC-TAA-TGTATGTGATA-TGCA---TTA---GTGT-
Myxo.	TGTAAACCAACTAT-TGT-AATATGTGATA-TATT---ACA---GTGT-
Mportucalensis	AGCAA-CCAGCCAC-TGTGTGCATGGGTAACTAAACATACA---GTGT-
Mmusculi	TTTTAACCAATTAC-TGC--TT--GCA-----GTA--
Mcyprini	TTTTAACCAATTAC-TGC--GCAAGCA-----GTA--
Mpseudodispar	TTTTAACCAATTAC-TGC--GCAAGCA-----GTA--
Mbibullatus	ACTTTACCAACTAT-TAC--ATGGGCAAC-----CATGTA---GTA--
Melegans	ACCAAACCAACTGT-TGT--GCGATCATTG---GTCGTACA---GCA--
Mpendula	ACTTAACCATCTAC-TGT--GTG-GCAAC-----ATACA---GTA--
Mpellicides	ACTTAACCATCTAC-TGT--GTG-GCAAC-----ATACA---GTA--
Mhungaricus	ACTTAACCATCTAC-TGC--GTG-GCAAC-----ACGTG---GTA--
Myxiditruttae	ATTAAGCCAGTCAA-CGATGTGGTA---TTTACG-TATTGCATTGTTGT

Myxidium	ATTAAACCAATCAT-TGTTTCAATAG---TTTAGGCTATTGTTGCATTGT
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	A--AATACCAACTT-TGGGCGAGTGGGGTTAAACTTGCTCGTCCAGATA-
Myxosp.	ACTAATACCAACTG-TATGTTAGTGG---TTAACGCTTCTGACGTACAA-
Hennzschokkei	ATGAAGCCAA-CTG-GTTGTTGGGGG---TGCAAATCTCCAACTTGTA-
Hennsalminicola	ATGAAGCCAA-CTG-CTTGTTGGGGG---TGCAAATCTCCAACTTGTA-
Ethclone2MA	TAA-TACCAAT----CACATGAGCAATCAT-----GTGTG
Ethclone3MA	TAA-TACCAAT----CACATGAGCAATCAT-----GTGTG
Ethclone1MA	-----
Myxobprocerus	GAA-TACCAAT----CGCATGAGCAATCAT-----GTGTG
Mprocerus	GAA-TACCAAT----CGCATGAGCAATCAT-----GTGTG
Henneguya	GAA-TACCAATGGCACGTACGAGAGATCGT-----GCGAG
Hsp.	GCGATACCAATGA--TGTACGTGAGTACAT-----GT---
Hdoori	GCGATACCAACCA--TGCACGTGAGTGCA-----
Hennictaluri	GCGATACCAACCG--TGTGTT---ACACAC-----GT---
Hennexilis	GCGATACCAACCG--TGTGC---AACAC-----GT---
Mspinacurvatura	CAATTACCAATCG--TGGGGC--AACCCAT-----GT---
Michkeulensis	CAATTACCAATCG--CGGGGC--AACCCGT-----GT---
Mosburni	TAATTACCAATTGAATGAGAGTAAAAATCTC-----ATTCA
Muvuliferus	-----
Ceratomyxa	-CTAAACCAATCGG--GCCTTAAAACCCA-----GTA--
Malgonquinensis	ACTAAACCACTTAC-TATGCTCCGGCATA-----GTA--
Perchclone1AM	ACACACCCAA-----
Perchclone2AM	ACACACCCAA-----
Perchclone3AM	ACACACCCAA-----
Melipsoides	--AGGTGAATCTAGATAACT-TTGCTGATCGTAT-GGCCTA-ATG--CCG
Mdjragini	--AGGTGAATCTAGNTAACT-TTGCTGATCGTAT-GGCCTA-ATG--CCG
Mbramae	--AGGTGAATCTAGATAACT-TTGCTGATCGTAT-GGCCTA-GTG--CCG
Mneurobius	--AGGTGAATCTAGATAACT-TTGCTGATCGTAT-GGCCTA-NTG--CCG
Marcticus	--AGGTGAATCTAAATAACT-TTGCTNATCGTAT-GGCCTA-TTG--CCG
Msandrae	--AGGTGAATCTAGATAACT-TTGCTGATCGTAT-GGCCTA-NTG--CCG
Minsidiosus	--YGGTGAYTCTAGATAACT-TTGCTGATCGTATTGGCCTA-GTG--CCG
Mcerebralis	--AGGTGAATCTAGATAACT-TTTCTGATCGTAT-GGCCTA-TAG--CCG
Mlentisuturalis	--GGATGAATCTAGATAACT-TTGCTGATCGTAGTGGCGTA-ATA-GCCG
Mxiaoi	--GGGTGAGTCTAGATAACT-TTGCTGATCGTAGTGGCATA-ATA-GCCG
Myxo.	--GGGTGAATCTAGATAACT-TTGCTGATCGTAGTGGCGTA-ATATGCCG
Mportucalensis	--GGGTGAATCTAGATAACT-TGTCTGATCGTAGTGGC-CA-TGAAGCCG
Mmusculi	--AGGTGAATCTAGATAACT-TTGCTGATCGTA-TGGCTTT-G---CCA
Mcyprini	--AGGTGAATCTAGATAACT-TTGCTGATCGCA-TGGCTCT-G---CCA
Mpseudodispar	--AGGTGAATCTAGATAACT-TTGCTGATCGCA-CGGCTTT-G---CCG
Mbibullatus	--AGGCGAATCTAGATAACT-TTGCTGATCGTA-TGGCCTT-GTG--CCG
Melegans	--AGGTGAATCTGGATAACT-TTGCTGATCGTA-TGGCCTA-GAG--CCG
Mpendula	--AGGCGAATCTAGATAACT-TTGCTGATCGTA-TGGCCTA-GTG--CCG
Mpellicides	--AGGCGAATCTAGATAACT-TTGCTGATCGTA-TGGCCTA-GTG--CCG
Mhungaricus	--AGGCGAATCTAGATAACT-TTGCTGATCGTA-TGGCCTT-GTG--CCG
Myxiditruttae	TGTGGTGAGTCTGGATAACT-GTGCCGATCGTA-TGGCCTT-GAG--CCG
Myxidium	TGTGGTGAGTCTGGATAACT-GTGCCGATCGTA-TGGCCTA-GAG--CCG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	---GGTGAATCTGGATAACT-G-GCTGATCGTA-TGGCCTT-GTG--CTG
Myxosp.	---GGTGAATCTAGATAACT-TTGCCGATCGCA-TGGCTTC-GGG--CCG

Hennzschokkei	---GGGGAGTCTAGATAACT-GTGCAGATCGTA-TGGCCTT-GTG--CTG
Hennsalminicola	---GGTGAGTCTAGATAACT-GTGCAGATCGTA-TGGCCTT-GTG--CTG
Ethclone2MA	---ATGAATCTAGATAACT-TTGCTGATCGCA-TGACCTT-GTG--TCG
Ethclone3MA	---ATGAATCTAGATAACT-TTGCTGATCGCA-TGACCTT-GTG--TCG
Ethclone1MA	-----
Myxobprocerus	---GTGAATCTAGATAACT-TTGCTGATCGCA-TGGCCTTTGAG--CCG
Mprocerus	---GTGAATCTAGATAACT-TTGCTGATCGCA-TGGCCTTTGAG--CCG
Henneguya	CGGGGTGAATCTAGATAACT-GTGCAGATCGCA-TGGCCTT-GAG--CCG
Hsp.	---GGTGAATCTAGATAACT-GTGCAGATCGTA-TGGCCTT-GAG--CTG
Hdoori	-----AATCTAGATAACTCGTGAGATCGTA-TGGCCTT-GAG--CTG
Hennictaluri	---GGCGAATCTAGATAACT-GTGCAGATCGTA-TGGCCTT-GAG--CTG
Hennexilis	---GGCGAATCTAGATAACT-GTGCAGATCGTA-TGGCCTT-GAG--CTG
Mspinacurvatura	---GGCGAATCTAGATAACTTG-GCAGATCGTA-TGGCCTT-GAG--CCG
Michkeulensis	---GGTGAATCTAGATAACTTA-GCAGATCGTA-TGGCCTA-GTG--CCG
Mosburni	--GGTTGAATCTAGATAACT-ATGCAGATCGTA-CGGCCTC-GTG--CTG
Muvuliferus	-----
Ceratomyxa	---GGTGAATCTAGATAACT-GTGCCGATCGTT---GCTTT-ATG---CG
Malgonquinensis	---AGGGAATCTGGATAACT-CTGCTGATCGTA-TGGCCTA-GTG--CCG
Perchclone1AM	-----TGCTTCAGAGTATCTCTTTTCACACCTTACCAACAAT-----TCA
Perchclone2AM	-----TGCTTCAGAGTATCTCTTTTCACACCTTACCAACAAT-----TCA
Perchclone3AM	-----TGCTTCAGAGTATCTCTTTTCACACCTTACCAACAAT-----TCA
Melipsoides	GCGTC-TTTC AATTGAA-TTTC TGCCC-TATT-AACTT--GTTGGTANTA
Mdjrugini	GCGNAGTTTCAATTGAA-TTTC TGCCC-TATT-AACTT--GTTGGTAGTA
Mbramae	GCGACGTTTCAATTGAA-TTTC TGCCC-TATT-AACTT--GTTGGTAGTA
Mneurobius	GCGNCNTTTC AATTGAA-TTTC TGCCC-TATT-AACTT--GTTGGTAGTA
Marcticus	GCAACTTTNCAATTGAA-TTNC TGCCC-TATT-AACTT--GTNGGTATTA
Msandrae	GCGACNTTTC AATTGAA-TTTC TGCCC-TATT-AACTT--GTTGGTAGTA
Minsidiosus	GCGACGTTTCAATTGAA-TTTC TGCCCCTATT-AACTT--GTTGGTAGRA
Mcerebralis	GCCACGTTTCAATTGAA-TTTC TGCCC-TATT-AACTA--GTTGGTAGTA
Mlentisuturalis	GCGACATTTCAAGTTGAA-TTTC TGCCC-TATC-AAATTG--GTTGGTAAGG
Mxiaoi	GCGACATTTCAAGTTGAA-TTTC TGCCC-TATC-AAATTG--GTTGGTAAGG
Myxo.	GCGACGTTTCAAGTTGAA-TTTC TGCCC-TATC-AAATTG--GTTGGTAAGG
Mportucalensis	GCGACATTTCAAGTTGAAATCTCTGCCC-TATC-AACTT--GTTGGTAAGG
Mmusculi	GCGACATTTCAATTGAG-TTTC TGCCC-TATC-AAATT--GTTGGTAAGG
Mcyprini	GCGACGTTTCAATTGAG-TTTC TGCCC-TATC-AAATT--GTTGGTAAGG
Mpseudodispar	GCGACGTTTCAATTGAG-TTTC TGCCC-TATC-AAATT--GTTGGTAAGG
Mbibullatus	ACGACGTTTCAATTGAG-TTTC TGCCC-TATC-AACTT--GTTGGTAAGG
Melegans	GCGACGTTTCAATTGAG-TTTC TGCCC-TATC-AACTA--GTTGGTAATG
Mpendula	ACGACGTTTCAATTGAG-TTTC TGCCC-TATC-AAATT--GTTGGTAAGG
Mpellicides	ACGACGTTTCAATTGAG-TTTC TGCCC-TATC-AAATT--GTTGGTAAGG
Mhungaricus	ACGACGTTTCAATTGAG-TTTC TGCCC-TATC-AAATT--GTTGGTAAGG
Myxiditrutae	GCGACATTTCAAGTTAAA-TTTC TGCCC-TATC-AACTT--GTTGGTAAGG
Myxidium	GCGACATTTCAAGTTAAA-TTTC TGCCC-TATC-AACTT--GTTGGTAAGG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	ACGACGTTTCAATTGAA-TTTC TGCCC-TATC-AACTT--GTTGGTAAGG
Myxosp.	GCGACGTTTCAATTGAA-TTTC TGCCC-TATC-AACTT--GTTGGTAAGG
Hennzschokkei	ACGACGTTTCAATTGAA-TTTC TGCCC-TATC-AACTA--GTTGGTAAGG
Hennsalminicola	ACGACGTTTCAATTGAA-TTTC TGCCC-TATC-AACTA--GTTGGTAAGG
Ethclone2MA	GCGACATTTCAAGTTGAG-TTTC TGCCC-TATC-AC-TT--GATGCTAGTG
Ethclone3MA	GCGACATTTCAAGTTGAG-TTTC TGCCC-TATC-AC-TT--GATGCTAGTG
Ethclone1MA	-----
Myxobprocerus	GCGACATTTCAAGTTGAG-TTTC TGCCC-TATC-ACCTT--GATGCTAGTG
Mprocerus	GCGACATTTCAAGTTGAG-TTTC TGCCC-TATC-ACCTT--GATGCTAGTG

Henneguya	GCGACATTTTCGATTGAG-TTTCTGCCC-TATC-ACCTA--GATGCAAGTG
Hsp.	ACGACGTTTCGATTGAA-TTTCTGCCC-TATC-AACTT-TGTCGACAGTG
Hdoori	ACGACGTTTCGATTGAA-TTTCTGCCC-TATC-AACTCATGTCGACAGTG
Hennictaluri	ACGACATTTTCGATTGAA-TTTCTGCCC-TATC-AACTC-TGTTGGCAGTG
Hennexilis	ACGACATTTTCGATTGAA-TTTCTGCCC-TATC-AACTC-TGTTGGCAGTG
Mspinacurvatura	GCGACGTTTCGATTGAA-TTTCTGCCC-TATC-AACTATTGTCGGTAGTG
Michkeulensis	GCGACGTTTCGATTGAA-TTTCTGCCC-TATC-AACTATTGTCGGTAGTG
Mosburni	ACGACGATTTCGATTGAG-TTTCTGCCT-TATC-AGCTT-TGTCGTGGCGG
Muvuliferus	-----
Ceratomyxa	GCGATATTTTCGATTGAG-TTTCTGCCC-TATC-AACTT--GTTGGTATGG
Malgonquinensis	GCGACGTTTCAATTGAG-TTTCTGCCC-TATCCAACCTT--GTTGGTAAGG
Perchclone1AM	A-ATCATATCAATACAG-----ACCTGTTAC-AGTCT--ATAAAGAAGA
Perchclone2AM	A-ATCATATCAATACAG-----ACCTGTTAC-AGTCT--ATAAAGAAGA
Perchclone3AM	A-ATCATATCAATACAG-----ACCTGTTAC-AGTCT--ATAAAGAAGA
Melipsoides	TANTTGCCTAC-CAAGGTTGCGACGGGTGACGGGGAATCAGG----GTTT
Mdjrugini	TANTTGCCTAC-CAAGGTTGCGACGGGTGACNGGGAATCAGG----GTTT
Mbramae	TAGTTGCCTAC-CAAGGTTGCGACGGGTGACCGGGAATCAGG----GTTT
Mneurobius	TAGTTGCCTAC-CAAGGTTGCGACGGGTGACCGGGAATCAGG----GTTT
Marcticus	TANTTGCCTAC-CACGGTTGCGACGGGTGACCGGGAATCAGG----GTTT
Msandrae	TAGTTGCCTAC-CAACGTTGCGACGGGTGACCGGGAATCAGG----GTTT
Minsidiosus	TAGTTGCCTAC-CAARGTTCCGACGGGTGACCGGGAATCAGG----GTTT
Mcerebralis	TAGAAGCCTAC-CAAGGTTGCGATGGGTAACGGGGAATCAGG----GTTT
Mlentisuturalis	TAGTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGGATCAGG----GTTT
Mxiao	TAGTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGGATCAGG----GTTT
Myxo.	TAGTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGGATCAGG----GTTT
Mportucalensis	TAGTGGCTTAC-CAAGGTTACAACGGGTAACGGGGAATCAGG----GTTT
Mmusculi	TATTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGAATCAGG----GTTT
Mcyprini	TATTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGAATCAGG----GTTT
Mpseudodispar	TATTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGAATCAGG----GTTT
Mbibullatus	TATTGGCTTAC-CAAGGTTGCAACGGGTAACGGGGAATCAGG----GTTT
Melegans	TATTGGATTAC-CAAGGTAGCAACGGGTAACGGGGAATCAGG----GTTT
Mpendula	TATTGGCTTAC-CAAGGTTGCAACGGGTAACGGGGAATCAGG----GTTT
Mpellicides	TATTGGCTTAC-CAAGGTTGCAACGGGTAACGGGGAATCAGG----GTTT
Mhungaricus	TATTGGCTTAC-CAAGGTTGCAACGGGTAACGGGGAATCAGG----GTTT
Myxiditrutae	TCGTTGCTTAC-CAAGGTGATTACGGGTAACGGGGAATCAGG----GTTT
Myxidium	TAGTGGCTTAC-CAAGGTGAT-ACGGGTAACGGGGAATCAGG----GTTT
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	TAGTGGCTTAC-CAAGGTTGTAACGGGTAACGGGGAATCAGG----GTTT
Myxosp.	TAGATGCTTAC-CAAGGTGGCGACGGGTAACGGGGGATCAGG----GTTT
Hennzschokkei	TAGATGCTAAC-CAAGGTTGTGACGGGTAACGGGGAATCAGG----GTTT
Hennsalminicola	TAGTAGTCAAC-CAAGGTTGTGACGGGTAACGGGGAATCAGG----GTTT
Ethclone2MA	TATTGAACTAG-CATGGGAGTTACGGGTAACGGAGGATCAGG----GTTT
Ethclone3MA	TATTGAACTAG-CATGGGAGTTACGGGTAACGGAGGATCAGG----GTTT
Ethclone1MA	-----
Myxobprocerus	TATTGAACTAG-CATGGGAGTAACGGGTAACGGAGGATCAGG-----TTC
Mprocerus	TATTGAACTAG-CATGGGAGTAACGGGTAACGGAGGATCAGG----GTTT
Henneguya	TATTGTACTTG-CATGGGGGTCACGGGTGACGGAGGATCAGG----GTTT
Hsp.	TAGTGGACTGC-CGAGGTTTTTTACGGGTAACGGGGAATCAGG----GTTT
Hdoori	TAGTGGACTGC-CGAGGTTTTTTACGGGTAACGGGGAATCAGG----GTTT
Hennictaluri	TAGTGGACTGC-CAAGGTTTTTTACGGGTAACGGGGAATCAGG----GTTT
Hennexilis	TAGTGGACTGC-CAAGGTTTTTTACGGGTAACGGGGAATCAGG----GTTT
Mspinacurvatura	TAGTGGACTAC-CGAGGTTTTTTACGGGTGACGGGGGATCAGG----GTTT
Michkeulensis	TAGTGGACTAC-CGAGGTTTTTTACGGGTGACGGGGGATCAGG----GTTT

Mosburni	TGAGGGCCCCA-CGTGGCAGTGACGAGTACGGGTGAATAAGG----GTTT
Muvuliferus	-----
Ceratomyxa	TATTGGCCTAC-CAAGGTTTGTACGGGTAACGGGGAATCAGG----GTTT
Malgonquinensis	TATTGGCCTAC-CAAGGTTGTCAACGGGTAACGG-GAATCAGG----GTTT
Perchclone1AM	TTTGAATTTACTCCAGCTCGTCA-----AATGGAGATTGTGATCTCGTCC
Perchclone2AM	TTTGAATTTACTCCAGCTCGTCA-----AATGGAGATTGTGATCTCGTCC
Perchclone3AM	TTTGAATTTACTCCAGCTCGTCA-----AATGGAGATTGTGATCTCGTCC
Melipsoides	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGGGAAGGC
Mdjragini	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGG-AAGGC
Mbramae	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGGNAAGGC
Mneurobius	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGGGAAGGC
Marcticus	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGG-AAGGC
Msandrae	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGGGAAGGC
Minsidiosus	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGG-AAGGC
Mcerebralis	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCATGG-AAGGC
Mlentisuturalis	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Mxiaoi	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Myxo.	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Mportucalensis	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Mmusculi	GATTCCGGAGAGGGAGCCTGAGAAATCGGCTACCAC-ATCCAAGG-AAGGC
Mcyprini	GATTCCGGAGAGGGAGCCTGAGAAATCGGCTACCAC-ATCCAAGG-AAGGC
Mpseudodispar	GATTCCGGAGAGGGAGCCTGAGAAATCGGCTACCAC-ATCCAAGG-AAGGC
Mbibullatus	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Melegans	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Mpendula	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-GAGGC
Mpellicides	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-GAGGC
Mhungaricus	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-AAGGC
Myxiditruttae	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-ACGGC
Myxidium	GATTCCGGAGAGGGAGCCTGAGAAACGGCTACCAC-ATCCAAGG-ACGGC
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	GATTCCGGAGAGGGAGCCTGAGAAATGGCTACCAC-ATCCAAGG-GAGGC
Myxosp.	GATTCCGGAGAGGGAGCCTGAGAAATGGCTACCAC-ATCCAAGG-ACGGC
Hennzschokkei	GATTCCGGAGAGGGAGCCTGAGAAATGGCTACCAC-ATCCAAGG-AAGGC
Hennsalminicola	GATTCCGGAGAGGGAGCCTGAGAAATGGCTACCAC-ATCCAAGG-AAGGC
Ethclone2MA	GATCCCGGAGAGGGAGCCTTAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Ethclone3MA	GATCCCGGAGAGGGAGCCTTAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Ethclone1MA	-----
Myxobprocerus	GATCCCGGAGAGGGAGCCTTAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Mprocerus	GATCCCGGAGAGGGAGCCTTAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Henneguya	GATCCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Hsp.	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Hdoori	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Hennictaluri	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Hennexilis	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Mspinacurvatura	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Michkeulensis	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Mosburni	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATNCAAGG-AAGGC
Muvuliferus	-----
Ceratomyxa	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-TTCTAAGG-AAGGC
Malgonquinensis	GATTCCGGAGAGGGAGCCTGAGAAAACGGCTACCAC-ATCCAAGG-AAGGC
Perchclone1AM	GTTACCGGAAT--AAACCTGACAAATCACT-TCACGAACTAAGA-ACGGC
Perchclone2AM	GTTACCGGAAT--AAACCTGACAAATCACT-TCACGAACTAAGA-ACGGC
Perchclone3AM	GTTACCGGAAT--AAACCTGACAAATCACT-TCACGAACTAAGA-ACGGC

Melipsoides	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGGGAGGTGGTGACGA
Mdjragini	AGCAGGCGC-GCAAATTACCCAATCC-ANACACTGGGGAGGTGGTGACGA
Mbramae	AGCAGGCGC-GCAAATTACCCAATCC-AG-CACTGGGG-GGTGGTGACGA
Mneurobius	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGGGAGGTGGTGACGA
Marcticus	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGGGAGGTGGTAACGA
Msandrae	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGGGAGGTGGTGACGA
Minsidiosus	AGCAGGCGC-CCAAATTACCCYATCCYAGANACCGGG-AGGTGGTGACGA
Mcerebralis	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGG-AGGTGGTGACGA
Mlentisuturalis	AGCAGGCGC-GCAAATTACCCAATCC-AGACAGTGGG-AGGTGGCGACGA
Mxiao	AGCAGGCGC-GCAAATTACCCAATCC-AGACAGTGGG-AGGTGGCGACGA
Myxo.	AGCAGGCGC-GCAAATTACCCAATCC-AGACAGTGGG-AGGTGGCAACGA
Mportucalensis	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGG-AGGTAGTAACGA
Mmusculi	AGCAGGCGC-GCAAATTACCAAATCT-AGACAGTAGG-AGGTGGTGAAGA
Mcyprini	AGCAGGCGC-GCAAATTACCAAATCT-AGACAGTAGG-AGGTGGTGAAGA
Mpseudodispar	AGCAGGCGC-GCAAATTACCAAATCT-AGACAGTAGG-AGGTGGTGAAGA
Mbibullatus	AACAGGCGC-GCAAATTACCCAATCT-AGACAGTAGG-AGGTGGTGAAGA
Melegans	AACAGGCGC-GCAAATTACCCAATCT-AGACAGTAGG-AGGTGGTGAAGA
Mpendula	AGCAGGCGC-GCAAATTACCCAATCT-AGACAGTAGG-AGGTGGTGAAGA
Mpellicides	AGCAGGCGC-GCAAATTACCCAATCT-AGACAGTAGG-AGGTGGTGAAGA
Mhungaricus	AGCAGGCGC-GCAAATTACCCAATCT-AGACAGTAGG-AGGTGGTGAAGA
Myxiditruttae	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGG-AGGTGGTGACGA
Myxidium	AGCAGGCGC-GCAAATTACCCAATCC-AGACACTGGG-AGGTGGTGACGA
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	AGCAGGCGC-GCAAATTACCCAATCC-ACAATGTGGG-AGGTGGTGACGA
Myxosp.	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Hennzschokkei	AGCAGGCGC-GCAAATTACCCAATTC-AGACAGTGAG-AGGTGGTGACGA
Hennsalminicola	AGCAGGCGC-GCAAATTACCCAATTC-AGACAGTGAG-AGGTGGTGACGA
Ethclone2MA	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Ethclone3MA	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Ethclone1MA	-----
Myxobprocerus	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Mprocerus	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Henneguya	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Hsp.	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGG--AGGTGGTGACGA
Hdoori	AGCAGGCGC-GCAAATTACCCAATCC-AGACGATGGG-AGGTGGTAACGA
Hennictaluri	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Hennexilis	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Mspinacurvatura	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Michkeulensis	AGCAGGCGC-GCAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Mosburni	AGCAGGCGC-GAAAATTACCCAATCC-AGACAATGGG-AGGTGGTGACGA
Muvuliferus	-----
Ceratomyxa	AGCAGGCGC-GCAAATTACCCAATCC-AGACATTGGG-AGGTAGTGACGA
Malgonquinensis	AACAGGCGC-GCAAATTACCCAATCT-AGACAGTAGG-AGGTGGTGAAGA
Perchclone1AM	--CATGCACCACCAATCACCAGATC-----ATGT-----A
Perchclone2AM	--CATGCACCACCAATCACCAGATC-----ATGT-----A
Perchclone3AM	--CATGCACCACCAATCACCAGATC-----ATGT-----A
Melipsoides	NAA-GTACTAAGTGGTGGCCCT-TA-GGGTC--GCTAGC-TTGGGAATGG-
Mdjragini	GAA-GTACTAAGTGGTGGCCCC-TA-GGGTC--GCTAGC-TTGGGAATGG-
Mbramae	NAA-GTACTAAGTGGTGGCCCT-TA-GGGTC--GCTAGC-TTGGGAATGG-
Mneurobius	NAA-GTACTAAGTGGTGGCCCT-TA-GGGTC--GCTAGC-TTGGGAATGG-
Marcticus	NAA-GTACTAAGTGGTGGCCCT-TA-GGGTC--GCTAGC-TTGGGAATGG-
Msandrae	GAA-GTACTAAGTGGTGGCCCT-TA-GGGT--GCTAGC-TTGGGAATGG-

Minsidiosus	GNAAGTACTAAGTGGTGGCCCT-TA-GGGTC--GCTAGC-TTGG RATGG-
Mcerebralis	GAA-GTACTAAGTGGTGGCCCT-TA-GGGTC--GCCAGC-TTGG AATGG-
Mlentisuturalis	GAA-TTACTGAGTGGTAGCCAT-AATGG-TT--ATCGAC-TTGA AATGA-
Mxiao	GAA-TTACTAAGTGGTAGCCTT-TATGG-TT--ATCGAC-TTGA AATGA-
Myxo.	GAA-TTACTGGGTGGTAGCCTT-TATGG-TT--ATCGAC-TTGA AATGA-
Mportucalensis	GAA-TTACTGAGTAGTGGTCCG-AATGGGCT--ACTAGT-TTGG AATGA-
Mmusculi	GAA-GTACTAAGTGGTGGTT----ATTTACT--ACCAGC-TTGG AATGG-
Mcyprini	GAA-GTACTAAGTGGTGGTT----ATTTACT--ACCAGC-TTGG AATGG-
Mpseudodispar	GAA-GTACTAAGTGGTGGTT----ATTTACT--ACCAGC-TTGG AATGG-
Mbibullatus	GAA-GTACTAAGTAGTGGTCC--AATGGATT--ACTAGC-TTGG AATGA-
Melegans	GAA-TTACTAAGTGGTGGCCCT--TATG-GCT--ACCAGC-TTGG AATGA-
Mpendula	GAA-GTACTTAGTGGTGACC----AAATGGT--CCCAAC-TTGG AATGA-
Mpellicides	GAA-GTACTTAGTGGTGACC----AAATGGT--CCCAAC-TTGG AATGA-
Mhungaricus	GAA-GTACCGGTGGTGGTTC----AATGACT--ACCAAC-TTGG AATGA-
Myxiditruttae	GAA-ATACCAGGTCTGTTCCC--AAAAGGAT--TCGACA-CTGG AATGA-
Myxidium	GAA-ATACCAGGTCTGTTCCC--AAAAGGAT--TCGACA-CTGG AATGA-
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	GAA-ATATTAGGTCTGTTTTCCCTCAGGGAAT--GCGATA-CTG AAAATGA-
Myxosp.	GAA-ATACTAGTCAGTGTCTC--AAAAGGAC--GCTGAT-CTGG AATGG-
Hennzschokkei	GAA-TTACCAAGTGGTGACC--ATAAGGTT--GCCAAT-TTGG AATGA-
Hennsalminicola	GAA-TTACCAAGTGGTGACC--ATAAGGTT--GCCAAT-TTGG AATGA-
Ethclone2MA	AGA-GTACCGGATGACTAAA--AAATTTTTTAGTCAGT-CTGG AATGA-
Ethclone3MA	AGA-GTACCGGATGACTAAA--AAATTTTTTAGTCAGT-CTGG AATGA-
Ethclone1MA	-----
Myxobprocerus	AGA-GTACCGGATGACTGAT---ATATTATT-AGTCAGT-CTGG AATGA-
Mprocerus	AGA-GTACCGGATGACTGAT---ATATTATT-AGTCAGT-CTGG AATGA-
Henneguya	AAA-GTACCGAATAAC-GCC---AAATGGTT--GTTAGT-TCAG AATGGG
Hsp.	AAA-GTACCAAAATGACGCCC--TAACGGGT--GTCAGT-TTGG AATGA-
Hdoori	AAA-GTACCAAAATGACGCCC--TAACGGGT--GTCAGT-TTGG AATGA-
Hennictaluri	AAA-GTACCGAATGGCAGCT---TA-TTGT--GCCAGT-TTGG AATGA-
Hennexilis	AAA-GTACCGAATGGCAGCT---TA-TTGT--GCCAGT-TTGG AATGA-
Mspinacurvatura	AAA-GTACCAGATGACGATC---TAA-GATT--GTCAAT-TTGG AATGG-
Michkeulensis	AAA-GTACCAGATGACGCTC---AAA-GACT--GTCAAT-CTGG AATGG-
Mosburni	AAA-GTACCGAATCTGGCCC---TAGTGGTT--AG-AGT-TTGG AATGG-
Muvuliferus	-----
Ceratomyxa	GAA-ATACCGGACTGGATCT----TACGAT---CCAGTACTGGA AATGA-
Malgonquinensis	GAA-TTACTAAATGGTGGTCT--TATTGATC--ATTTGT-TTGG AATGA-
Perchclone1AM	AGATCTTCGGTCTGACAATCC-----TT-----TTG--ATGT-
Perchclone2AM	AGATCTTCGGTCTGACAATCC-----TT-----TTG--ATGT-
Perchclone3AM	AGATCTTCGGTCTGACAATCC-----TT-----TTG--ATGT-
Melipsoides	ACGTAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Mdjragini	ACGTAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Mbramae	ACGTAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Mneurobius	ACGTAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Marcticus	ACGTAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Msandrae	ACGTAYT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGACAA-GTC-TG
Minsidiosus	ACGTAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Mcerebralis	ACGTAAAT-TTAAGTAATTCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Mlentisuturalis	ACGTGATATTAAGAAATCCG-TTGAGTAACAACCTGGAGGGCAA-GTC-TG
Mxiao	ACGTGAT-TTAAGAAATCCG-TTGAGTAACAACCTGGAGGGCAA-GTC-TG
Myxo.	ACGTGAT-TTAAGAAATCCG-TTGAGTAACAACCTGGAGGGCAA-GTC-TG
Mportucalensis	ACGTGAT-TTAGTACATCCG-ATGAGTAACAACCTGGAGGGCAA-GTC-TG
Mmusculi	ACGTAAT-TTAAGCAATTCG-ATGAGTAACCTACTGGAGGGCAA-GTCCTG

Mcyprini	ACGTAAT-TTAAGCAATTTCG-ATGAGTAACTACTGGAGGGCAA-GTCCTG
Mpseudodispar	ACGTAAT-TTAAGCAATTTCG-ATGAGTAACTACTGGAGGGCAA-GTCCTG
Mbibullatus	ACGTAAT-TTAAGCAATTTCG-ATGAGTAACTACTGGAGGGCAAAGTCCTG
Melegans	ACGTAAT-TTAAGCAATTTCG-ATGAGTAACTACTGGAGGGCAA-GTCCTG
Mpendula	GGCCAAT-TTAAGAAATTCG-TCGAGTAACTACTGGAGGGCAA-GTCCTG
Mpellicides	GGCCAAT-TTAAGAAATTCG-TCGAGTAACTACTGGAGGGCAA-GTCCTG
Mhungaricus	ACGTAAT-TTAAGCAATTTCG-ATGAGTAACTACTGGAGGGCAA-GTCCTG
Myxiditruttae	GTGCAAT-TTAAAAATTTCA-ACGAGTAACTACTGGAGGGCAA-GTC-TG
Myxidium	GTGCAAT-TTAAAAATTTCA-ACGAGTAACTACTGGAGGGCAA-GTC-TG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	GCACAAT-TTAGAACATTTG-TCGAGTAACTACTGAAGGGCAA-GTC-TG
Myxosp.	ACGCAAT-TTAAGCAATTTCG-ATGAGTAACTACTGGAGGGCAA-GTC-TA
Hennzschokkei	ACGTAAT-TTAAGAAATTCG-ATGAGTAGCACTGGAGGGCAA-GTC-CG
Hennsalminicola	ACGTAAT-TTAAGAAATTCG-ATGAGTAACTACTGGAGGGCAA-GTC-CG
Ethclone2MA	AATCAAT-TTAAGCAATTGAGTTGAGTAACTACTGGAGGGCAA-GTC-TG
Ethclone3MA	AATCAAT-TTAAGCAATTGAGTTGAGTAACTACTGGAGGGCAA-GTC-TG
Ethclone1MA	-----
Myxobprocerus	AATCAAT-TTAAGCAATTGAGTTGAGTAACTACTGGAGGGCAA-GTC-TG
Mprocerus	AATCAAT-TTAAGCAATTGAGTTGAGTAACTACTGGAGGGCAA-GTC-TG
Henneguya	AATCAAT-TTAAGCAATTGA-TTGAGTAACGACTGGAGGGCAA-GTC-TG
Hsp.	AAACAAT-TTAGGAATTTGAATTGAGTAACGACTGGAGGGCAA-GTC-TG
Hdoori	AAACAAT-TTAGGAATTTGAATTGAGTAACGACTGGAGGGCAA-GTC-TG
Hennictaluri	AAACAAT-TTAAGTAATTGAATTGAGTAACGACTGGAGGGCAA-GTC-TG
Hennexilis	AAACAAT-TTAAGTAATTGAATTGAGTAACGACTGGAGGGCAA-GTC-TG
Mspinacurvatura	AATCAAT-TTAAGCAATTGA-TTGAGGAACGACTAGAGGGCAA-GTC-TG
Michkeulensis	AATCAAT-TTAAGCAATTGA-TTGAGGAACGACTAGAGGGCAA-GTC-TG
Mosburni	A--CGAT-ANATAAACATGATCGAGAATTCAACTGGAGGGCAA-GTC-TG
Muvuliferus	-----
Ceratomyxa	ACGATAT-GTAATCATTTTCG-ATGAGGATCTACTGGAGGGCAA-GTC-TG
Malgonquinensis	ACGTAAC-TTAAGAAATTCG-TTGAGAAACAACTGGAGGGCAA-GTCCTG
Perchclone1AM	-CCTGAT-CCGGTAAGTTTCCCGTGT-----TGA--GTCAA-----
Perchclone2AM	-CCTGAT-CCGGTAAGTTTCCCGTGT-----TGA--GTCAA-----
Perchclone3AM	-CCTGAT-CCGGTAAGTTTCCCGTGT-----TGA--GTCAA-----
Melipsoides	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Mdjragini	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Mbramae	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Mneurobius	GTGCCAGCAGTC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Marcticus	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Msandrae	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Minsidiosus	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Mcerebralis	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATTTTAAA-G
Mlentisuturalis	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTACCTCAAAA-G
Mxiao	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTACCTCAAAA-G
Myxo.	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTACCTCAAAA-G
Mportucalensis	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTGGC-GTACTTTAAA-G
Mmusculi	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTGGC-GTGATTTAAA-G
Mcyprini	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTGGC-GTGATTTAAA-G
Mpseudodispar	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTGGC-GTGATTTAAA-G
Mbibullatus	GTGCCAACAGCCCGCGGTAATTCCAGCTCCAAGTTGT-GTGATTTAAAAG
Melegans	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTGAT-CTCGTTTAAA-G
Mpendula	GTGCCAGCAGCCCGCGGTAATTCCAGCTTC-AGTGGCCGTGATTTAAA-G
Mpellicides	GTGCCAGCAGCCCGCGGTAATTCCAGCTTCAGTGGCCGTGATTTAAA-G
Mhungaricus	GTGCCAGCAGCC-GCGGTAATTCCAGCTCC-AGTGGC-GTGATTTAAA-G

Myxiditrutae	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGT-GTATTCAAAC-G
Myxidium	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGT-GTATCCAAAC-G
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	GTGCCAGCAGCC-GCGGTAATTCCAGCTTCA-GTAGT-GTATATTAAT-G
Myxosp.	GTGCCAGCAGCC-GCGGTAATTCTAGCTCCA-GTGGT-GTATTTTAAT-G
Hennzschokkei	GTGCCAGCAGCC-GCGGTAATTCCGGCTCCA-GTAGC-GTATTTTAAA-A
Hennsalminicola	GTGCCAGCAGCC-GCG-TAATTCCGGCTCCA-GTAGC-GTATTTTAAA-A
Ethclone2MA	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTGTCTCAAA-G
Ethclone3MA	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTGTCTCAAA-G
Ethclone1MA	-----
Myxobprocerus	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTGTCTCAAA-G
Mprocerus	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTGTCTCAAA-G
Henneguya	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATCTCAAA-G
Hsp.	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATCTCAAA-G
Hdoori	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATCTCAAA-G
Hennictaluri	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATCTCAAA-G
Hennexilis	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATCTCAAA-G
Mspinacurvatura	GTGCCAGCAGCC-GCGGTAATTCCAGCTCTA-GTAGC-GTATCCCAAA-G
Michkeulensis	GTGCCAGCAGCC-GCGGTAATTCCAGCTCTA-GTAGC-GTATCCCAAA-G
Mosburni	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGC-GTATCGCAAA-G
Muvuliferus	-----
Ceratomyxa	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGT-GTATATCAAC-A
Malgonquinensis	GTGCCAGCAGCC-GCGGTAATTCCAGCTCCA-GTAGT-TTGCTTTAAA-G
Perchclone1AM	--ATTA--AGCC-GCAG--GCTCCACCCCTG-GTGGT-GCCCTTCC---G
Perchclone2AM	--ATTA--AGCC-GCAG--GCTCCACCCCTG-GTGGT-GCCCTTCC---G
Perchclone3AM	--ATTA--AGCC-GCAG--GCTCCACCCCTG-GTGGT-GCCCTTCC---G
Melipsoides	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Mdjragini	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Mbramae	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Mneurobius	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Marcticus	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Msandrae	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Minsidiosus	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Mcerebralis	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Mlentisuturalis	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACATG--GTTG-GGTC
Mxiao	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACATG--GTTAAGGAT
Myxo.	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACATG--GTTAGGTAT
Mportucalensis	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTGTA---GT
Mmusculi	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCAAGCGGTGACA---GT
Mcyprini	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCATGCGGTGACA---GC
Mpseudodispar	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCAAGCGGTGTA---GT
Mbibullatus	TTGTTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACCCAGTGACA---GT
Melegans	TTGTTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGCATCA---TC
Mpendula	TTGCTGCGTTTAAAAACGCTCGTAG--TTGGGATCATGCAGTGATA---TT
Mpellicides	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCATGCAGTGATA---TT
Mhungaricus	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ATCACGCAGTAGTA---TT
Myxiditrutae	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ACAAGAGAGTAACCTTGATG
Myxidium	TTGCTGCGTTTAAAAACGCTCGTAG--TTGG-ACAAGAGAGTAGCTTGGAG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	CTGCGGCGGTTAAAAACGCTCGTAG--TTGA-ATGAC---GAAGCTCGGA

Myxosp.	TTGCTGCGCTTTAAAACGCTCGTAG--TTGG-ATTAC---GCAGTGTGAGT
Hennzschokei	TTGTTGCGCTTAAAACGCTGGTAG--TTGG-ATGAC---ACAGTCATGAT
Hennsalminicola	TTGTTGCGCTTAAAACNCTCGTAG--TTGG-ATGAC---ACAGTCATGAT
Ethclone2MA	TTGCTGCGCTTGAAAACGCTCGTAG--TTGG-ATGAGAACTACAC---AT
Ethclone3MA	TTGCTGCGCTTGAAAACGCTCGTAG--TTGG-ATGAGAACTACAC---AT
Ethclone1MA	-----
Myxobprocerus	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATGAGAATCTATAT---AT
Mprocerus	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATGAGAACTACAT---AT
Henneguya	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATCATTAACTGTGT---GT
Hsp.	TTGCTGC-CTTAAAACGCCCCGTAG--T-GG-A-CACGCACTGTGT---GT
Hdoori	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATCACGCACTGTGT---GT
Hennictaluri	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATCACGCACTATGC---AT
Hennexilis	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATCACGCACTATGC---AT
Mspinacurvatura	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATCACAACTATAC---GC
Michkeulensis	TTGCTGCGCTTAAAACGCTCGTAG--TTGG-ATCACAACTATAT---TC
Mosburni	NTGCTGCGCTTAAAACGNTCGTAAGTTGGG-ATCACAGTCTGTGT---GT
Muvuliferus	-----
Ceratomyxa	TTGTTGCGGTTAAAACGCTCGTAG--TTGG-ATAACGAG-----
Malgonquinensis	TTGTTGCGGTTAAAACGCTCGTAG--TTGG-ATCACGCAGTGATG---TT
Perchclone1AM	TCAATTCCCTTTAAG-----TTTCAGCCTTGCGACCA-----
Perchclone2AM	TCAATTCCCTTTAAG-----TTTCAGCCTTGCGACCA-----
Perchclone3AM	TCAATTCCCTTTAAG-----TTTCAGCCTTGCGACCA-----
Melipsoides	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTG---GTG
Mdjragini	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTG---GTG
Mbramae	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTG---GTG
Mneurobius	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTG---GTG
Marcticus	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTG---GTG
Msandrae	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTG---GTG
Minsidiosus	TGGTAAGCTGATTG---AATGGTGCT-----CCAACTGTTTTYG---GTG
Mcerebralis	TGGTAGGCTGATCG---AATGGTGCTA-----CTAACTGCTCCA---GCG
Mlentisuturalis	TAGTGAaaaaaacgt---GTTAGTACGCT-AAATCCTCAAGTGTAGATGTG
Mxiao	CAGTGAATTGGCAT---GTAGGTACAAT-AAATCCTCAAGTGTAGATGTA
Myxo.	CAGTGAATTACCAT---GTTGGCACAAA-AAATCCTCAAGTGTAGGTGTG
Mportucalensis	CAGTGGAATGGCGC---AAT-GTGCAAT-AAACCCCTCACCGGTTGATGTG
Mmusculi	TTGTAGTCTGCGGT---TG-----GC-----ATTTGGCTGCTTC---AG
Mcyprini	CTGTAGTCTGCGGC---TT-----GC-----ATTTGGCTGCTTC---AG
Mpseudodispar	TTGTAGTCTGCGAT---TA-----GT-----ATCTGACTGCTTC---AG
Mbibullatus	CGGTTATTTTCGGT---TAT-ACGCT-----AACTGTCGATTTT---TG
Melegans	TGGTAATCT-TGAT---AGT-GAGCATA-AAAGCTG--CACTTTGAGGAG
Mpendula	CAGTGACCATAAAT---GGT-ACAT-----GTATACGATTTT---CT
Mpellicides	CAGTGACAATAAAT---GGT-ACAT-----GTATACCATTTT---CT
Mhungaricus	CAGTAATCTAGGCC---AAT-GTAT-----GCTGACGATTG-----CC
Myxiditrutae	CGGGCGGGGAGTGT---AATTACGCTAT-GTGTTCACTCGTTGGG--TGA
Myxidium	CGG--CGGGAGTGT---AAT-GCTCTGT-GTGATCACTGATTAAAG--GAA
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	CGTACTTTAACGT---A--TGCTACAT-CAAACCG-----TG
Myxosp.	CGGTCAATTAGCAG---AGCTTCACATT-AAAACCG-----AG
Hennzschokei	CAGTTGATTGTCAT---GGTTATACATT-AAACTTT-----
Hennsalminicola	CAGTTGATTGTCAT---GGTTATACATT-AAACTTT-----
Ethclone2MA	TGGATAGTTATTATCTAGGTTATAGACT-ACATATTTTCGTATATCTGTG
Ethclone3MA	TGGATAGTTATTATCTAGGTTATAGACT-ACATATTTTCGTATATCTGTG
Ethclone1MA	-----GGTTATAGACT-ACATATTTTCGTATATCTGTG
Myxobprocerus	CGGATAGTTATCTACTAGATTTAAGGCT-A-ATTATTACTTATTTCCGTG

Mprocerus	CGGATAGTTATTTACTAGATTTAGGACT-A-ATTATTACATATTTCCGTG
Henneguya	GGGGTAAGTTGGAACGATGTT-TGCCCT-ACACCG----ACATGGCCGCG
Hsp.	TGGTGGATGTGCAA---T-C-GTGCTAT-GACCCCTCTTCTTCCAGGCACG
Hdoori	TGGTGGATGCGCAT---TGC-GTGCTATTGACCCCTCTTCTTCTGGGCGCG
Hennictaluri	TGGTGTACTCAAGTGTTCG-ATGGTA--AACCTAG-GCT--CGGTTTTTC
Hennexilis	TGGTGTATCCAAGTGTTCG-ATGGTA--AGCTTTGTGCT--CAGACT-C
Mspinacurvatura	GAGTTGGTGTGG-----TGCTGTATCAA-ATTTATGAA---TCATGCAAG
Michkeulensis	GAGTTGGTGTGG-----TGCTGTGTCAA-AATTTTAAAGATTTTCCCGGG
Mosburni	GTGTGAATCGGAA---TGCGGTGCGTTCAACTTTTATAATTTACATGCT
Muvuliferus	-----
Ceratomyxa	-GGTAAAT-----AATTATATGA-----
Malgonquinensis	CAGTAATCTTTAG---AGGGGTGTGCTCAAATGTATCCGTC-----
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Mdjragini	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Mbramae	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Mneurobius	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Marcticus	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Msandrae	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Minsidiosus	ATA-A-----ATTTCTA--TTTATTACTAAAAACAGTGT-----
Mcerebralis	TTG-A-----ATTTCAA--TTCAGTGTGGAGTAGTGT-----
Mlentisuturalis	ACG-GTGTGTTATCACCGTTGCAA--TTTATATGT--TGTGGAGT-----
Mxiao	GTG-GGGTCTTATCCCTACTACAA--TTTATATGT--TGTGGAGT-----
Myxo.	ACG-GTGTGCTATCACCGTTGCAA--TTTATATAT--TGTGGAGT-----
Mportucalensis	TCT-ACGTTCACCGTGGACACAT--TCAATTGACG-TGAGGAGT-----
Mmusculi	TC-----GATG-----CTGATC-----GTGG-----
Mcyprini	TC-----GATG-----C-AGCC-----GTGG-----
Mpseudodispar	TC-----GATG-----CTTTTC-----GTAG-----
Mbibullatus	TCC-AGATATGCAAATAGATGGCA--CTAATCTGATGGGTAG-----
Melegans	TT-----TTCACGTAAGTGTAAGGCCGATT-AAAGTGTAGTGTCT----
Mpendula	CCA-C-----CGAGTTTAT--TGAATC-----TGTNCC-----
Mpellicides	CCA-A-----CGAGTTTAT--TGAATC-----TGTACC-----
Mhungaricus	TCA-----GGGCGTAT--CGGATC-----TTCATT-----
Myxiditruttae	-TG--CGTTTGCTGTTTTGCGAAAAGTGAAGCAGTTGCGAGGAGCC-TTTC
Myxidium	GTG-TCGCTCGCTAT----CGAAAAGTTA-GCAAGTACTATTGACTATTCC
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	CTG-TAGTATTGCGAAGCAGACCACGTCTGTCTA-GTGGTGCTGCAGTAC
Myxosp.	GCG-TTGTCTTCATGAAAACACGGTTACCGTGAACGTGACTCAGCGTCTT
Hennzschokkei	-GA-CTCTTTACTGTAGATGTCTAACGACGTATATGGTGTGAGT-----
Hennsalminicola	-GA-CTCTCTACTGTAGATGTCTAACGACGTATATG-TGTTGAGT-----
Ethclone2MA	----ATGCTTTCA--GTGTCCAGATAATATGGGTATTAGTTGAAC-----
Ethclone3MA	----ATGCTTTCA--GTGTCCAGATAATATGGGTATTAGTTGAAC-----
Ethclone1MA	----ATGCTTTCA--GTGTCCAGATAATATGGGTATTAGTTGAAC-----
Myxobprocerus	----ATGCTTATG--GTATCCGGACAATATTGATATTGGTTCAGC-----
Mprocerus	----ATGCTTATG--GTATCCGGATAATAGTGATATTGGTTCAAC-----
Henneguya	TTGCATGTTCTCAT-GTTTCGTGACA-TGTTGG--TTGGCGGAAC-----
Hsp.	----GTATCATA--CGTGCACCTGGAGCTGAGGGAGTGCGTAATG-----
Hdoori	----GTATCATTACCGTGCACCTGGGGCTGGGGGAGTGCGTAGTG-----
Hennictaluri	----GTTCTACG--ATTCTCCGCAGTAAAGG--TGTGTAAGGC-----
Hennexilis	----GTTCCACG--ATGTTCTGCAGCTAAGG--TGTGCAGCAC-----
Mspinacurvatura	----GTGTCCAC-----ATTTTGT--ATGATTCT-TATGTGGTGCA-----

Michkeulensis	-----GTGTCATC-----ACTTCGGTAATGATT---TATGTGGTGCA-----
Mosburni	-----ATGTAAAA-----TGTGAAAGAGTGCGTCTATTATTC-----
Muvuliferus	-----GCAAA-----
Ceratomyxa	-----ACGTACGGCCATTTGACGTCACCCGCT-----
Malgonquinensis	-----
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----GC-TTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Mdjragini	-----GC-TTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Mbramae	-----GC-TTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Mneurobius	-----GC-TTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Marcticus	-----GCCTTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Msandrae	-----GC-TTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Minsidiosus	-----GCCTTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Mcerebralis	-----GCCGTCTTTTCAGTTATT-CGCC-AATT-----TACACTACTTA
Mlentisuturalis	-----ACTGCACGTTTTTTGTTA-GCT-AGGC-----CCAACGATTG
Mxiaoi	-----ACTGCATGTCGATTGTTA-GCT-GAAC-----TTAACTGATTG
Myxo.	-----GCTGCATGTTGATTGTTA-GCT-GAGC-----CTAACTGATTG
Mportucalensis	-----ACATTGTGCTATTGTAA-GCT-GATT-----GACATTACTTG
Mmusculi	-----AT-----T-AA---AACGCA-AGCT-----GTCACTATTTA
Mcyprini	-----AT-----TTAA---AGCGCA-GGCT-----GTCACTATTTA
Mpseudodispar	-----AT-----T-AA---AACGCA-AACT-----GTCACTATTTA
Mbibullatus	-----ACCGTAAATAAA---AACGCC-GAAT-----GTTACTATTTG
Melegans	-----CACCG---ATGGATATTAACGCC-AGGT-----GTTGCTAGTTG
Mpendula	-----ATTT---TGCGTGTTTATGCTTGAAT-----GTTACTATTTG
Mpellicides	-----ATTT---TGCGTGTTTATGCT-GAAT-----GTTACTATTTG
Mhungaricus	-----GGCT---CAGATGTTTATGCT-GAAT-----GCTACTATTTG
Myxiditruttae	AGGATGATGCATAGTTGTATCACTTGTGCGACCATGCTGCCAGGTTACTTG
Myxidium	TTGTTGTAGCATAGTAGTAGCACTTGTGCGACCATGCT-CCAGGCTACTTG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	GGCGTGGCGTATGTTAATA--AACAGTCGTTTC-----CGGGCTACTAG
Myxosp.	GGTGTGCTGCATGTTGATT--ATCAGCCGGCC-----CACACTACTTG
Hennzschokkei	---TGAAAGTATAAATATGGCAATTGTGTGCTGAATG---TGACTAGTTG
Hennsalminicola	---TGAAAGTATAAATATGGCAATTGTGTGCTGAATG---TGACTAGTTG
Ethclone2MA	-----TAGTAGTAACATATAACTG---GTGTGTAGTAATTG
Ethclone3MA	-----TAGTAGTAACATATAACTG---GTGTGTAGTAATTG
Ethclone1MA	-----TAGTAGTAACATATAACTG---GTGTGTAGTAATTG
Myxobprocerus	-----TAGTGATAACATTAAACTG---GTATGTAGTAATTG
Mprocerus	-----TAGTGATAACATTAAACTG---GTATGTAGTAATTG
Henneguya	-----GCCGAACAACACATAACCG---ACATGCAGTAATTG
Hsp.	-----TGC-ATCGTAAGCTG---ATACACAGTAATTT
Hdoori	-----TGC-GTCGTAAGCCG---ATACACAGTAATTT
Hennictaluri	-----TTGA-GTAATTAGCCA---GTGCATAGTAATTG
Hennexilis	-----TTGG-GTAAATAGCCA---GTGTGTAGTAATTG
Mspinacurvatura	-----GTGCTGCTACCGTAAGCTA---GTGTATAGTAATTG
Michkeulensis	-----GTGTTGCTAC-GTAAGCTA---GTGTATAGTAATTG
Mosburni	-----TGATTGGAAGCGC---GCCACAGTAATTG
Muvuliferus	-----
Ceratomyxa	-----TGCTCATTGATTA-----TTTTGCTCTTTT
Malgonquinensis	-----TCAGATATCATGCTGGACG-----TCGCTAGTTG
Perchclone1AM	-----TACTCCCCC
Perchclone2AM	-----TACTCCCCC

Perchclone3AM -----TACTCCCC

Melipsoides CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Mdjragini CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Mbramae CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Mneurobius CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Marcticus CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Msandrae CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Minsidiosus CG-----CGTAAGGATGGCAGTTGACCTTTAGTGCGTCGATTGCCG-TG
Mcerebralis CG-----CGTAAGGATGGCAGTTG-CCTTTAGTGCGTCGATTGCCG-TG
Mlentisuturalis CG-----TATGGGGATGGCAATTGACCTTAAGTGAGTCGATTGTTG-TG
Mxiao CA-----TATGGGGATGGTAATTGACCTTAAGTGAGTCGATTATTG-TG
Myxo. CA-----TATGGGGATGGTGATTGACCTTTAGTGAGTCGATCATCG-TG
Mportucalensis CA-----CGTGGGAATGGTCATTGACCTTAGTTGAGTTGACGATTG-TG
Mmusculi AT-----CGCAAAATATGTTGGTTGGCCTTTAGTGAGTCGAGCATCA-TA
Mcyprini AT-----CGCAAAATATGTTGGTTGGCCTTTAGTGAGTCGAGCATCA-TA
Mpseudodispar AT-----CGCAAAATATGTTAGTTGACCTTTAATGAGTTGACTATCA-TA
Mbibullatus CA-----CGCAAGTATGATAGTTGACCTTAAATGCGTTGAGTGTCATCA-TG
Melegans CA-----CGCAGGATGTCAGTTGGTCTTTAGTGACTGAGTGTCATCA-TG
Mpendula CA-----CACAAATATGGCATTGGCCTTTAGTGAGTTGAGTGTCATCA-TG
Mpellicides CA-----CACAAATATGGCATTGGCCTTTAGTGAGTTGAGTGTCATCA-TG
Mhungaricus CA-----CACAAATATGGTATTTGGCCTTAAAGTGAGTTGAGTATCA-TG
Myxiditruttae TAATGTTTCGTGAGAATGGTACATGGTCTTTATTGTACTGTGTGCTT-TG
Myxidium TAATGTTTCGTGAGAATGGTATATGGTCTTCATTGTACTGTGTATTA-TG
Ethclone1AM -----
Ethclone3AM -----
Ethclone2AM -----
Mdiaphanus -----
M. lieberkuehni AT-----TGTGAGAATGGTGAATGGTCTTTACCGACT--GCTCATCGTG
Myxosp. TA-----CGTGAGGATGGCGGTTGACCTTTAGTGCGTTGACTGTTCTGTG
Hennzschokkei CA-----CGTGAGGATGATGGTTGACCTCTGGTGCGTTGACTGTTA-TG
Hennsalminicola CA-----CGTGAGGATGATGGTTGACCTTTAGTGCGTTGACTGTTA-TG
Ethclone2MA CG-----TATGAGAATGATCATTTGACCTTGAGTGTTGA-AGATCGTG
Ethclone3MA CG-----TATGAGAATGATCATTTGACCTTGAGTGTTGA-AGATCGTG
Ethclone1MA CG-----TATGAGAATGATCATTTGACCTTGAGTGTTGA-AGATCGTG
Myxobprocerus CG-----TATGAGGATGACCATTGACCTTAAAGTGTTGA-AGGTCTGTG
Mprocerus CG-----TATGAGGATGACCATTGACCTTAAAGTGTTGA-AGGTCTGTG
Henneguya CG-----CGTGGGGATGGCCTTTGACCTTAAAGTGCGTTGA-GGGCCGTG
Hsp. CA-----CGTGAGGATGATCGCTGACCTTGAGTGTTGCGG-TGATCGTG
Hdoori CA-----CGTGAGGATGATCGCTGACCTTGAGTGTTGCGGTGATCGTG
Hennictaluri CA-----CGTGGGGATGGCCGTTAGCCTTTAGTGCGCTGA-TGGTCTGTG
Hennexilis CA-----CGTGGGGATGGTCTGTTAGCCTTTAGTGCGCTGA-TGATCGTG
Mspinacurvatura CA-----CGTGAGGATAGACATTAGCCTTGAATGAGTAGA-TGTTTGTG
Michkeulensis CA-----CGTGAGGATAGACATTGCGCTTGAATGAGTAGA-TGTTTGTG
Mosburni CA-----CGCGAGGATGACCATTGACCTTTGGTGTTGG-TGGTGGTG
Muvuliferus -----GA--ACTA-----G
Ceratomyxa TA-----TTACAAGGGTCAATACTTTGCTTAAATTGAATTG--TATTG-AA
Malgonquinensis CA-----CGTGAGATAGATTGTTGGCCTTGAATGTGCCGG-TATTCTTG
Perchclone1AM CG-----CAACCGAAAACTTTAG-----G
Perchclone2AM CG-----CAACCGAAAACTTTAG-----G
Perchclone3AM CG-----CAACCGAAAACTTTAG-----G

Melipsoides T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Mdjragini T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Mbramae T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Mneurobius T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Marcticus TACTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T

Msandrae	T--TTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Minsidiosus	T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Mcerebralis	T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Mlentisuturalis	T-CCCATGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Mxiao	T-CCCATGGAGT-GTGCG-TTGAATAAAACAG-----
Myxo.	T-CCCATGGAGT-GTGCC-TTGAATAAAACAGAGTGCTTAAAGCAGGC-T
Mportucalensis	T-CTCACGGGGT-GTGCC-TTGAATAAATCAGAGTGCTTAAAGCAGGC-T
Mmusculi	T-TTTGCGGAGTTGTGCC-TTGAATAAAACAGAGTGCTCAAAGCAGGC-G
Mcyprini	T-TTTGCGGAGTTGTGCC-TTGAATAAAACAGAGTGCTCAAAGCAGGC-G
Mpseudodispar	T-TTTGCAGAGTTGTGCC-TTGAATAAAACAGAGTGCTCAAAGCAGGC-G
Mbibullatus	T-CTTGCGGAGT-GTGCC-TTGAATAAANCAGAGTGCTTAAAGCAGGC-T
Melegans	T-CTTGCGGAGT-GTGCC-TTGAATAAAACAGAGTGCTCAACGCAGGC-A
Mpendula	T-CTTGTGGAGT-GTGCC-TTGAATAAAACAGAGTGCTCAAAGCAGGC-G
Mpellicides	T-CTTGTGGAGT-GTGCC-TTGAATAAANCAGAGTGCTCAAAGCAGGC-G
Mhungaricus	C-CTTGTGGAGT-GTGCC-TTGAATAAAACAGAGTGCTCAAAGCAGGC-G
Myxiditruttae	T-CTTACGTAGT-GTGCC-TTGAATAAATCAGAGTGCTCAATGCAAGC--
Myxidium	T-CTTACGTAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAAGC--
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----TTAACGGCGCGA-A
M.lieberkuehni	T-CTCGCAGAGT-GTGCC-TTGAATAAAACAGAGTGCTCAAAGCAGGC-A
Myxosp.	T-CTTACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC--
Hennzschokkei	T-CTCACGGAGT-GTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Hennsalminicola	T-CTCACGGAGG-TTGCC-TTGAATAAATCAGAGTGCTCAAAGCAGGC-T
Ethclone2MA	T-CTCATGG-GATGTGCC-TTGAGTAAATCAGAGTGCTCAAAGCAGGC-G
Ethclone3MA	T-CTCATGG-GATGTGCC-TTGAGTAAATCAGAGTGCTCAAAGCAGGC-G
Ethclone1MA	T-CTCATGG-GATGTGCC-TTGAGTAAATCAGAGTGCTCAAAGCAGGC-G
Myxobprocerus	T-CTCATGG-GATGTGC--TTGAGTAAATCAGAGTGCTCAAAGCAGGC-G
Mprocerus	T-CTCATGG-GATGTGCC-TTGAGTAAATCAGAGTGCTCAAAGCAGGC-G
Henneguya	T-CTCACGG-GATGTGCC-TTGAGTAAATCAGAGTGCTCAAAGCAGGC-A
Hsp.	T-CTCACGG-G-TGTGCC-TTGAGTAAATCAGAGTGCTTAAAGCAGGC--
Hdoori	T-CTCACGG-G-TGTGCCGTTGAGTAAATCAGAGTG-----
Hennictaluri	T-CTCACGG-GGTGTGCC-TTGAGTAAATCAGAGTGCTTAAAGCAGGC--
Hennexilis	T-CCCACGG-GGTGTGCC-TTGAGTAAATCAGAGTGCTTAAAGCAGGC--
Mspinacurvatura	T-CTCACGG-AGTGTGCC-TTGAATAAATCAAAGTGCTTAAAGCAGGC-T
Michkeulensis	T-CTCACGGGAGTGTGCC-TTGAATAAATCAAAGTGCTTAAAGCAGGCCT
Mosburni	T-CTCGCGG-GGTGTGCC-TTGAGTAAATCAGAGTGCTTAAATGCAAGC--
Muvuliferus	T-CCTGCAG-GTT-----AACGAATC----TCCTT-----
Ceratomyxa	TACTTGTATAGC-GTGCC-TTGAATAAAGCACAGTGCTCAAAGCAAGC-G
Malgonquinensis	T-CTTGCGGGGT-GTGCC-TTGAATAAAACAGAGTGCTTAAAGCAGGT--
Perchclone1AM	T-----TTCC-----CAGGGGGACCAACCCAAGC--
Perchclone2AM	T-----TTCC-----CAGGGGGACCAACCCAAGC--
Perchclone3AM	T-----TTCC-----CAGGGGGACCAACCCAAGC--
Melipsoides	-TTTGCTTGAATGTTAATAGCATGGAACGAACAATTGTGTA-GTAGTATG
Mdjrugini	-TTTGCTTGAATGTTAATAGCATGGAACGAACAATTGTGTA-GTAGTATG
Mbramae	-TTTGCTTGAATGTTAATAGCATGGAACGAACAATTGTGTA-GTAGTATG
Mneurobius	-TTTGCTTGAATATTAATAGCATGGAACGAGCAATTGTGTA-GTAGTATG
Marcticus	-TTTGCTTGAATGTTAATAGCATGGAACGAACAATTGTGTA-GTAGTATG
Msandrae	-TTTGCTTGAATGTTAATAGCATGGAACGAACAATTGTGTA-GTAGTATG
Minsidiosus	-TTTGCTTGAATGTTAATAGCATGGAACGAACAATTGTGTA-GTAGTATG
Mcerebralis	-TTTGCTTGAATGTTAATAGCATGGAACGAANAATTGTGTA-GTAGTGTG
Mlentisuturalis	TGATGCCTGAATGTTAACAGCATGGAACGAACAATTGTGGA-AACATGAT
Mxiao	-----
Myxo.	TGACGCCTGAATGTTAACAGCATGGAACGAACAATTGTGGC-AACATGAT
Mportucalensis	CT-TGCCTGAATGTTGATAGCATGGAACGAACAGTTGTGTT-AACGTGCA

Mmusculi	-AACGCTTGAATGTT-GTAGCATGGAACGAACAAACGTGTA-TTCGCACA
Mcyprini	-AACGCTTGAATGTT-GTAGCATGGAACGAACAAACGTGTA-TTGGCACA
Mpseudodispar	-AACGCTTGAATGTT-GTAGCATGGAACGAACAAACGTGTA-TTCGCATA
Mbibullatus	-AAAGCTTGAATGTT-ATAGCATGGAACGAACAAACGTGTG-ACAGTACG
Melegans	-AAAGCTTGAATGTT-ATAGCATGGAACGAACAAACGTGTT-AACGTGCA
Mpendula	-TATGCTTGAATGTT-ATAGCATGGAACGAACAAACGTGTA-TTTGTGTA
Mpellicides	-TATGCTTGAATGTT-ATAGCATGGAACGAACAAACGTGTA-TTTGNGTA
Mhungaricus	-AAAGCTTGAATGTT-ATAGCATGGAACGAACAAACGTGTA-TTTGCGTA
Myxiditruttae	AATTGCTTGAATGTTAATAGCATGGAACGAATG--TATAGTGAAAGTGTG
Myxidium	TATCGCTTGAATGTTAATAGCATGGAACGAATG--TATAGTGTATGTGCG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	TCGCCCTT-----CATGGASTGAACAATAGTGTA-GATGTGTA
M.lieberkuehni	TGTTGCTTGAATGTTAATAGCATGGAACGAACAAATGTGAT-TTCGTGTA
Myxosp.	TTACGCTTGAATGTTAATAGCATGGAACGAACAAATGTGTTATACGTGTC
Hennzschokkei	TTAAGCTTGAATGTTAATAGCATGGAACGAACAAACGTGTT-TATGTATA
Hennsalminicola	TTAAGCTTGAATGTTAATAGCATGGAACGAACAAACGTGTT-TATGTATA
Ethclone2MA	TTATGCCTGAATGTTAATAGCATGGAACGAACAAATAGTGTAGA-TGTGTA
Ethclone3MA	TTATGCCTGAATGTTAATAGCATGGAACGAACAAATAGTGTAGA-TGTGTA
Ethclone1MA	TTATGCCTGAATGTTAATAGCATGGAACGAACAAATAGTGTAGA-TGTGTA
Myxobprocerus	TGATGCCTGAATGTTGATAGCATGGAACGAACAAATAGTGTAGA-TGTGTA
Mprocerus	TGATGCCTGAATGTTGATAGCATGGAACGAACAAATAGTGTAGA-TGTGTA
Henneguya	TTGCGC-TGAATGTTGATAGCATGGAACGAACAAATCGTGTACAGTGCGCG
Hsp.	GTAAGCTTGAATGTTGATAGCATGGAACGAACAAATCGTGTATGGTGTGTA
Hdoori	-TAAGCTTGAATGTTGATAGCATGGAACGAACAAATCGTGTATGGTGTGTA
Hennictaluri	GTTAGCTTGAATGTTGATAGCATGGAACGAACAAATCGTGTATGGTGTGTG
Hennexilis	GTTAGCTTGAATGTTGATAGCATGGAACGAACAAATCGTGTATGGTGTGTG
Mspinacurvatura	GTACGCTTGAATGTTAA-AGCATGGAACGAACAAATCGTGTAGGGTGCATG
Michkeulensis	TTNCGCTTGAATGTTAA-AGCATGGAACGAACAAATCGTGTAGGGTGCATG
Mosburni	GTGTGCTTGAATGTTGATAGCATGGAACGAACAAATAGTGTATGGTGTGTG
Muvuliferus	-----CATGGAACGAACAAATAGTGTATGGTGTGTG
Ceratomyxa	TAACGCTAGAATGTT-ATAGCATGGAACGAATAGAT-TGAC-----
Malgonquinensis	TATCGCCTGAATGTT-ATAGCATGGAACGAACAAATCGTGTATGGTGTG
Perchclone1AM	---CGT---AATAATAA-GGCATAGGACTAA-----
Perchclone2AM	---CGT---AATAATAA-GGCATAGGACTAA-----
Perchclone3AM	---CGT---AATAATAA-GGCATAGGACTAA-----
Melipsoides	-TTGTGACAC-ATAGCGATCGGTCTTTGA-CTGAATGTTATT-----G
Mdjragini	-TTGTGACAC-ATAGCGATCGGTCTTTGA-CTGAATGCTATT-----G
Mbramae	-TTGTGACAC-ATAGCGATCGGTCTTTGA-CTGAATGCTATT-----G
Mneurobius	-TTGTGACAC-ATAGCGATCGGTCTTTGA-CTGAATGCTATT-----G
Marcticus	-TTGTGACAC-ATAGCGATCGGNCTTTGA-CTGAATGCTATT-----G
Msandrae	-TTGTGACAC-ATAGCGATCGGTCTTTGA-CTGAATGCTATT-----G
Minsidiosus	-TTGTGACAC-ATAGCGATCGGTCTTTGA-CTGAATGCTATT-----G
Mcerebralis	-TTGTGACAA-ATAGCGATCGGTCTTTGA-CTGAATGTTATT-----
Mlentisuturalis	ATTATCTCCACTGG-CAATGTGCTTATTTGTATGTTGTTCATG-----T
Mxiao	-----
Myxo.	GTCGTGCTGATTGGGCGATATACTTTTATGTGTGTCGTTTCA-----A
Mportucalensis	ATGGTTGTATTGGTATCGCGAAGCTTTGAGCCTAGCGT-----G
Mmusculi	--TGTCGTGCGAAGTGAGTGTGCTTCTGGTGCGTCTGCT-TT-----G
Mcyprini	--CGTCGTGCGAAGTGCGGTGCGCTTCTGGTGCGTCTGCT-TT-----G
Mpseudodispar	--CGTCGTGCGAAGTGCGGTGCGCTTCTGGTGCGTCTGCT-TT-----G
Mbibullatus	--TGTTTGCGAACGGCGATGAGCTCATGGTTTAT-TGCTGTT-----G
Melegans	--CGTCGAGT-ACAGCGGTATTCTTATGGATTGC-CATTGTA-----T
Mpendula	--TGNTTGGTGGTAATCAGAGGCAACT---CTATTTACT-----G
Mpellicides	--TGTTTGGTGGTAATCAGAGGCAACT---CTATTTACT-----G

Mhungaricus	--TGTTTAGTGATGATTGGGGGCAACT---CTGATTGTT-----G
Myxiditruttae	-ATGGTGATGG-----CAATTGCTTGT-----
Myxidium	-CTAGTAAGCA-----TCATTTCGTGGT-----
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-TTGGTAA-TGGTAGACG----TTACAA-CGTAATGTTCT-----
M.lieberkuehni	-CTGACAAGTAATGCACAATGATTACAT-TGTTGTGCACT-----
Myxosp.	-TATACTTACAGTTGA-ATGGATTATTT-TGTCTAGCTGT-----
Hennzschokkei	-AT-ATAATTGATTTTGTATTTGTCTATGA-TAGATTATTAT-----G
Hennsalminicola	-AT-ATAATTGATTTTGTATTTGTCTATGA-TAGATTATTGTT-----G
Ethclone2MA	TTGAA---GATGGTAGTATGTTATTATAACTTAATAATA-----TA
Ethclone3MA	TTGAA---GATGGTAGTATGTTATTATAACTTAATAATA-----TA
Ethclone1MA	TTGAA---GATGGTAGTATGTTATTATAACTTAATAATA-----TA
Myxobprocerus	TTGAA---GATGGTAGTGTATTATTACAATGTAATAGTA-----TA
Mprocerus	TTGAA---GATGGTAGTGTATTATTACAATGTAATAGTA-----TA
Henneguya	CTGATATCGGTGCTGGTGTGT-GCTGGCGTGCAAAAGTCGCGTCGCCCCA
Hsp.	TTGAGTACGGTGT--GTGTGGCGTTACTGCGCA-----
Hdoori	TTGAGTACGGTGT--GTGTGGCGTTACTGCGCA-----
Hennictaluri	TTGATAACGGCGT--GT-TGGATCT-TCCTTCA-----
Hennexilis	TTGGTAACGGGGT--GT-TGGATTCGTCCTTCA-----
Mspinacurvatura	TTGGTCAATGGGT--GTGAGGCCTAGTCTTGCA-----
Michkeulensis	TTA-TTTATGGGT--GCAGGGCCTAGCCAGGCA-----
Mosburni	TTGGCCAAGATGT--GA-----TGCAAAATG-----
Muvuliferus	TTGGCAAAGATGT--AA-----TTT--ATTG-----
Ceratomyxa	-----
Malgonquinensis	--CGTCTCTGGGCAGTTGTGAACCTTTGGTTTACA-----G
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Mdjrugini	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Mbramae	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Mneurobius	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Marcticus	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Msandrae	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Minsidiosus	CTGTTGCA---GCATACAGCACCAACCACCAATAACGGATGTTGGYT-CC
Mcerebralis	CAGTTACA---GCATACAGCACCAACCACCAATAACGGATGTTGGTT-CC
Mlentisuturalis	GGGGGCAG---TATCATAGCACCAACCACCAATATCGGATGTTGGTT-CC
Mxiao	-----
Myxo.	TTAGTCGA---GATCATAGCACCAACCACCAATATCGGATGTTGGTT-CC
Mportucalensis	CTGGTCCC---ATGCACAGCACCAACCACCAATATCGGATGTTGGTT-CC
Mmusculi	CTGAC-----GTGTGTATCACCCGCCAA-AATA-CGATTGTTGGTTATC
Mcyprini	CTGAT-----GTGTGTATCACCCGCCAA-AGTA-CGATTGTTGGTTATC
Mpseudodispar	CTGAT-----GTATGCATCACCCGCCAA-AGTA-CGATTGTTGGTTATC
Mbibullatus	TCGAT-----GCGTACAGCACCTACCAA-AATA-TGGATGTTGGTT-CC
Melegans	TAGAT-----GTGCACAGCACCTACCAA-AATA-TGGATGTTGGTT-CC
Mpendula	TCGGC-----ATATGCAGCACCCGCCAA-AATA-CGGATGTTGGTTTTTC
Mpellicides	TCGGC-----ATATGCAGCACCCGCCAA-AATA-CGGATGTTGGTTTTTC
Mhungaricus	CTGGC-----ATACGCAGCACCCACCAA-AATA-CGGATGTTGGTTTTTC
Myxiditruttae	CATTGCTG---TTACACAGCACCAACCAGCGAAGCG-GAGCATTTGGTTGCC
Myxidium	GTTTGCTA---GTACATAGCACCAACCAGCGAAGCG-GAACATTTGGTTACC
Ethclone1AM	-----GTT-CC
Ethclone3AM	-----GTT-CC
Ethclone2AM	-----
Mdiaphanus	ACT--ACA---ATATGCAACACCGACCGCCAATTACGGATGTTGGTT-CC

M.lieberkuehni	AACTGGCA---GTACATAGCACCAACCACCAAG-GCGGATGTTGGTTTCC
Myxosp.	GGGT-GTA---GTTACATACCAACCGCCAATAACGGATGTTGGTTTCC
Hennschokkei	TTTTTGTG---TTGTACTGCACCAACCACCAATAACGGACGTTGGTT-CC
Hennsalminicola	TTTTTGTG---TTGTACTGCACCAACCACCAATAACGGACGTTGGTT-CC
Ethclone2MA	CTACTACA---ATATGCAACACCGACCGCCAATTACGGATGTTGGTT-CC
Ethclone3MA	CTACTACA---ATATGCAACACCGACCGCCAATTACGGATGTTGGTT-CC
Ethclone1MA	CTACTACA---ATATGCAACACCGACCGCCAATTACGGATGTTGGTT-CC
Myxobprocerus	CTACTACA---ATATGCAACACCGACCGCCAATTACGGATGTTGGTT-CC
Mprocerus	CTACTACA---ATATGCAACACCGACCGCCAATTACGGATGTTGGTT-CC
Henneguya	GTGCTGCA---GTGCGCGACACCGACCACCATGTACGGATGTTGGTT-CC
Hsp.	-CATTTCA---ATGCATGACACCGACCACCAATTACGGATGTTGGTTTCC
Hdoori	-CATTTCA---ATGCATGACACCGACCACCAATTACGGATGTTGGTTTCC
Hennictaluri	-CACTGCA---ACACACGACACCGACCGCCAATTACGGATGTTGGTT-CC
Hennexilis	-CACTGCA---ACACACGACACCGACCGCCAATTACGGATGTTGGTT-CC
Mspinacurvatura	-TCCATCA---ACATGTGACACCGACCGCCAATAACGGATGTTGGTTCC-
Michkeulensis	-TTCATCA---ACATGTGACACCGACCACCAATAACGGATGTTGGTTCCC
Mosburni	-CGTCTCA---ACACATGACACCGACCACCAATTACGGATGTT-GGTTCC
Muvuliferus	-CGTCTCA---ACACATGACACCGACCACCAATTACGGATGTTTGGTTTCC
Ceratomyxa	CTGAATCA---GTTTGT-----TGGTAAAAAGTTACGCAAGTAACCTAACC
Malgonquinensis	TTGTTTCGAGATGCGCACGGCACCCACCA--AAATATGGCTGTTGGTT-CC
Perchclone1AM	-----TCCC
Perchclone2AM	-----TCCC
Perchclone3AM	-----TCCC
Melipsoides	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mdjragini	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mbramae	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mneurobius	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Marcticus	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Msandrae	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Minsidiosus	GTATTGGGGTGATGATYAATTAAAAARGAGCGGNTGGGGGCATTGGTATTTGGC
Mcerebralis	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mlentisuturalis	GTATTGGGGTGAT-GATTAAAGAGGAGCGGTTGGGGGCATTGGTATTTTGC
Mxiao	-----
Myxo.	GTATTGGGGTGAT-GATTAAAGAGGAGCGGTTGGGGGCATTGGTATTTTGC
Mportucalensis	GTATTGGGGTGAT-GATTAAAGAGGAGCGGTTGGGGGCATTGGTATTTTGC
Mmusculi	GTATTAGG-TGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTTGA
Mcyprini	GTATTAGG-TGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTTGA
Mpseudodispar	GTATTAGG-TGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTTGA
Mbibullatus	ATATACGG-TGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTTGC
Melegans	ATATACGG-TGAT-GATTAAAAAGGAGCGGTTGGGGGCATTAGTATTTTGC
Mpendula	GTATCAGG-TGAT-GATTAAACAGGAGCGGTTGGGGGCATTGGTATTTTGC
Mpellicides	GTATCAGG-TGAT-GATTAAACAGGAGCGGTTGGGGGCATTGGTATTTTGC
Mhungaricus	GTATCAGG-TGAT-GATTAAACAGGAGCGGTTGGGGGCATTGGTATTTTGC
Myxiditruttae	GACTTCGGGTGAG-GATTAAAGAGGGGCATTTGGGGGCATTAGTATTTTGGT
Myxidium	GACTTCGGGTGAG-GATTAAAGAGGNGCNATTTGGGGGCATTAGTATTTTGGC
Ethclone1AM	GTATTGGGGTACT-GATTAAAGAGGAGCGGTTGGGGGCATTGGTATTTTGC
Ethclone3AM	GTATTGGGGTACT-GATTAAAGAGGAGCGGTTGGGGGCATTGGTATTTTGC
Ethclone2AM	-----
Mdiaphanus	GTATTGGGGTACT-GATTAAAGAGGAGCGGTTGGGGGCATTGGTATTTTGC
M.lieberkuehni	GTTTTGGGGTGAT-GATTAAAAAGGGGCGGTTGGGGGCATTGGTATTTTGC
Myxosp.	GTATTGGGGTGAT-GATTAAAAAGGAGCGATTGGGGGCATTGGTATTTTGC
Hennschokkei	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTTGC
Hennsalminicola	GTATTGGGGTGAT-GATTAAAAAGGAGCGGTTGGGGGCATTGGTATTTTGC
Ethclone2MA	GTATTGGGGTACT-GATTAA-----
Ethclone3MA	GTATTGGGGTACT-GATTAA-----
Ethclone1MA	GTATTGGGGTACT-GATTAA-----

Myxobprocerus	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mprocerus	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGGCATTG-TATT--GC
Henneguya	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Hsp.	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGACATTGGTATTTGGC
Hdoori	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGACATTGGTATTTGGC
Hennictaluri	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Hennexilis	GTATTGGGGTATT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mspinacurvatura	GTATTGGGGTATT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Michkeulensis	GTATTGGGGTATT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Mosburni	GTATTGGGGTACT-GATTAAGAGGAGCGGTTGGGGGCATTGGTATTTGGC
Muvuliferus	GTATTGGGGTACT-GATTAAGAGGAG-GTTTGGGGGCATTGGTATTTGGC
Ceratomyxa	TGAAACGGGTCCCT-GATTAAGAGGGGCATTGAGGATGTTAGTACTTGGT
Malgonquinensis	ATATACGG-TGAT-GATTAACAGGAGCGGTTGGGGGCATCGGTATTTGGC
Perchclone1AM	GGGTCGGCATAGT---TTACAGTGAG-AACTAGGA---CGGTATCTGAT
Perchclone2AM	GGGTCGGCATAGT---TTACAGTGAG-AACTAGGA---CGGTATCTGAT
Perchclone3AM	GGGTCGGCATAGT---TTACAGTGAG-AACTAGGA---CGGTATCTGAT
Melipsoides	-CGCGAG-AGGTGAAA-TTTTGTAGACCGG--CCAAGGACTAACGGAATGC
Mdjragini	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACGGAATGC
Mbramae	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACGGAATGC
Mneurobius	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACG-AATGC
Marcticus	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACG-AATGC
Msandrae	-CGCGAG-AGGTGAAA-TTTTGTAGACCGG--CCAAGGACTAACGGAATGC
Minsidiosus	-CNCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACG-AATGC
Mcerebralis	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACG-AATGC
Mlentisuturalis	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CAAAGGACTAACA-AATGC
Mxiao	-----
Myxo.	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CAAAGGACTAACA-AATGC
Mportucalensis	ACGCGAGGAGGTGAAA-TTCTTGGACCGTTCCCAAGGACTAACA-AATGC
Mmusculi	-CGCGAG-AGGTGAAA-TTCAAAGACCGT--CCAAGGACTAACA-AATGC
Mcyprini	-CGCGAG-AGGTGAAA-TTCAAAGACCGT--CCAAGGACTAACA-AATGC
Mpseudodispar	-CGCGAG-AGGTGAAA-TTCAAAGACCGT--CCAAGGACTAACA-AATGC
Mbibullatus	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACA-AATGC
Melegans	-CGCGAG-AGGTGAAA-TTCTAGAACCGG--TCAAGGACTAACA-AATGC
Mpendula	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CCAAGGACTAACA-GATGC
Mpellicides	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CCAAGGACTAACA-GATGC
Mhungaricus	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CCAAGGACTAACA-GATGC
Myxiditruttae	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CCAAGGACTAACA-AGTGC
Myxidium	-CGCGAG-AGGTGAAA-NTCTTGGACCGG--CCAAGGACTAACA-AGTGC
Ethclone1AM	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CCAAGGACTAACA-AATGC
Ethclone3AM	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CCAAGGACTAACA-AATGC
Ethclone2AM	-----
Mdiaphanus	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CCAAGGACTAACA-AATGC
M.lieberkuehni	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--CCAAGGACTAACA-AATGC
Myxosp.	-CGCGAG-AGGTGAAA-T-CTTGGACCGG--CCAAGGACTAACA-AATGC
Hennzschokkei	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--ACAAGGACTAACA-AATGC
Hennsalminicola	-CGCGAG-AGGTGAAA-TTCTTGGACCGG--ACAAGGACTAACA-AATGC
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CACAAG-----
Mprocerus	-AGCGAG-AG-TGAA--TTCT--GACCTG--C-CAAG-----
Henneguya	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CCAAGGACTAACA-AATGC
Hsp.	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--TCAAGGACTAACA-AATGC
Hdoori	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--TCAAGGACTAACA-AATGC
Hennictaluri	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CCAAGGACTAACA-AATGC
Hennexilis	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--CCAAGGACTAACA-AATGC

Mspinacurvatura	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--TCAAGGACTAACA-AATGC
Michkeulensis	-AGCGAG-AGGTGAAA-TTCTTGGACCTG--TCAAGGACTAACA-AATGC
Mosburni	-AGCGAG-AGGTGAAA-TTCTTAGACCTG--CCAAGGACTAACA-AATGC
Muvuliferus	-AGCGAG-AGGTGAAAA-TTCTTAGACCTG--CCAAGGACTAACA-AATGC
Ceratomyxa	-GGCGAG-AGGTGAAA-TTCTTAGACCCA--CCAAAGACTCACT-ATTGC
Malgonquinensis	-CGCGAG-AGGTGAAA-TTCTTAGACCGG--CCAAGGACTAACA-AATGC
Perchclone1AM	---CG-----TCTTCGAACCT-----CTCACT-----
Perchclone2AM	---CG-----TCTTCGAACCT-----CTCACT-----
Perchclone3AM	---CG-----TCTTCGAACCT-----CTCACT-----
Melipsoides	GGAAGGCATTTGTCTA-GACCGCNTCG-CTTAATC-AAGAACGA-TAGTG
Mdjragini	GGAAGGCATTTGTCTA-GACCGCNTCG-CTTAATC-AAGAACGA-TAGTG
Mbramae	GGAAGGCATTTGTCTA-GACCGCNTCG-CTTAATC-AAGAACGA-TAGTG
Mneurobius	G-AAGGCATTTGTCTA-GACCGCCTCG-CTTAATC-AAGAACGA-TAGTG
Marcticus	G-AAGGCATTTGTCTA-GACCGCNTCG-CTTAATC-AAGAACGA-TAGTG
Msandrae	GGAAGGCATTTGTCTA-GACCGCNTCG-CTTAATC-AAGAACGA-TAGTG
Minsidiosus	G-AAGGCATTTGTCTA-GACCGCCTCG-CTTAATC-AAGAACGA-TAGTG
Mcerebralis	G-AAGGCATTTGCCCA-GACCGCCTCG-CTTAATC-AAGAACGA-TAGTG
Mlentisuturalis	G-AAGGCATTTGCCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
Mxiaoi	-----
Myxo.	G-AAGGCATTTGCCCA-GACCGCATCC-ATTAATC-AAGAACGA-TAGTG
Mportucalensis	GGAAGGCATTTGCCCAAGACNGTATCC-ATTAATC-AAGAACGA-AAGTG
Mmusculi	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-AAGTG
Mcyprini	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-AAGTG
Mpseudodispar	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-AAGTG
Mbibullatus	-GAAGGCATTTGTCTA-GACCGTTTCC-ATTAATC-AAGAACGA-AAGTG
Melegans	-GAAGGCATTTGTCTA-GACCGTTTCC-ATTAATC-AAGAACGA-AAGTG
Mpendula	-GAAGGCGTTTGTCTA-GACCGTTTCC-ATTAATC-AAGAACGA-AAGTG
Mpellicides	CGAAGGCGTTTGTCTA-GACCGTTTCC-ATTAATC-AAGAACGA-AAGTG
Mhungaricus	-GAAGGCGTTTGTCTA-GACCGTTTCC-ATTAATC-AAGAACGA-AAGTG
Myxiditruttae	-GAAAGCATTTGCCAA-GTATGTTCCC-TTAAATC-AAGAACGA-AAGTG
Myxidium	-GAAAGCATTTGCCAA-GTATGTTTCC-CTTAATC-AAGAACGA-AAGTG
Ethclone1AM	-GAAGGCATTTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
Ethclone3AM	-GAAGGCATTTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
Ethclone2AM	-----
Mdiaphanus	-GAAGGCATTTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
M.lieberkuehni	-GAAGGCATCTGCCAA-GACCGTTCCC-GTTGATC-AAGAGCGA-TAGTG
Myxosp.	-GAAGGCAATTGCCAA-GATCGTTTCC-ATTAATC-AAGAACGA-CAGTC
Hennzschokkei	-GAAGGCATTTGCCCA-GACCGTCTCT-ATTAATC-AAGAACGA-TAGAG
Hennsalminicola	-GAAGGCATTTGCCCA-GACCGTCTCT-ATTAATC-AAGAACGA-TAGAG
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
Hsp.	-GAAGGCATCTGTCCA-GACCGTACC--ATTAATTCAGAACGA-TAGTG
Hdoori	-GAAGGCATCTGTCCA-GACCGTATCCGATTAATCGAAGAACGAATAGTG
Hennictaluri	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGAA-AGAG
Hennexilis	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGAA-AGAG
Mspinacurvatura	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
Michkeulensis	-GAAGGCATCTGTCCA-GACCGTATCC-ATTAATC-AAGAACGA-TAGTG
Mosburni	-GAAGGCATCTGTCCA-GACCGTATCC-ATTGATC-AAGAGCGAA-AGTG
Muvuliferus	-GAAGGCATCTGTCCA-GACCGTATCC-ATTGATC-AAGRGCGAA-AGTG
Ceratomyxa	-GAAGGCATTCATCAA-GAATGTTTTTC-ATTAATC-AAGAGCGA-AAGTT
Malgonquinensis	-AAAGGCACTTGTCTA-GACCGTTTCC-ATTAATC-AAGAACGA-AAGTG
Perchclone1AM	-----ATCGTTCTTGATTAAT--GGATACGG-----

Perchclone2AM	-----ATCGTTCTTGATTAAT--GGATACGG-----
Perchclone3AM	-----ATCGTTCTTGATTAAT--GGATACGG-----
Melipsoides	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACTGT-AA
Mdjragini	GGAGG-TTCGAAGACG---ATCAGATACCGNCCTAGTNCC-CACTGT-AA
Mbramae	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACTGT-AA
Mneurobius	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACTGT-AA
Marcticus	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACTGT-AA
Msandrae	GGAGG-TTCGAAGACG---ATCAGATACCGACCTAGTNCC-CACTGT-AA
Minsidiosus	GGAGG-TTCGAAGACG---AACAGATACCGTCCTAGTTCC-CACTGT-AA
Mcerebralis	G-AGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACTGT-AA
Mlentisuturalis	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACCGT-AA
Mxiao	-----
Myxo.	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACCGT-AA
Mportucalensis	GAAGG-TTCCAAGTCG---ATTAGATACCGCCGTAGTTTC-CACTGT-AA
Mmusculi	GGAGG-TTCGAAGACG---ATTAGATACCGTCGTAGTTCC-CACTGT-AA
Mcyprini	GGAGG-TTCGAAGACG---ATTAGATACCGTCGTAGTTCC-CACCGT-AA
Mpseudodispar	GGAGG-TTCGAAGACG---ATTAGATACCGTCGTAGTTCC-CACTGT-AA
Mbibullatus	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCC-CACTGT-AA
Melegans	AGAGG-TTCGAAGACG---ATCAGATACCGTCGTAGTTCT-CACCGT-AA
Mpendula	GGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTAT-AA
Mpellicides	GGAGG-TTCGAAGACG---ATCAGATNCCGTCTAGTTCT-CACTAT-AA
Mhungaricus	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACCGT-AA
Myxiditruttae	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACCGT-AA
Myxidium	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACCGT-NA
Ethclone1AM	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
Ethclone3AM	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
Ethclone2AM	-----G---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
Mdiaphanus	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
M.lieberkuehni	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
Myxosp.	CGAGG-TTCGAAGTCG---ATCAGATACCGACTTAGTTTCG-GACCGT-AA
Hennzschokkei	GAAGGATC-GAAGAGG---ATCAGATACCCTCGTAGTTTC-CTCCGT-AA
Hennsalminicola	GAAGGATNCGAAGAGG---ATCAGATACCCTCGTAGTTTC-CTCCGTAA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
Hsp.	AGAGG-TTCGAAGACG-GATTCAGATACCGTCCTAGTTCT-CACCGT-AA
Hdoori	AGAGG-TTCGAAGAGACGAGTCAGATACCGTCCTAGTTCT-CACCGT-AA
Hennictaluri	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CTCTGT-AA
Hennexilis	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CTCTGT-AA
Mspinacurvatura	AGAGG-TTCGAAGACG---ATCAGATACCG-CCTAGTTCT-CACTGT-AA
Michkeulensis	AGAGG-TTCGAAGACG---ATCAGATACCGTCCTAGTTCT-CACTGT-AA
Mosburni	CGGGG-ATCGAAGATG---ATCAGATACCATCCTAGTCCG-CACCCT-AA
Muvuliferus	CGGGG-ATCGAAGATG---ATCAGATACCATCCTAGTCCG-CACCCT-AA
Ceratomyxa	GGAGA-ATCGAAGACG---ATCAGATACCGTCCTAGTTCCATACAGT-AA
Malgonquinensis	GGAGG-TTCGAAGACG---ATCAGATACCGTCATAGTTCC-CACCGT-AA
Perchclone1AM	-----TCTGGA-----CAAATGCCTTCGCATTT-----
Perchclone2AM	-----TCTGGA-----CAAATGCCTTCGCATTT-----
Perchclone3AM	-----TCTGGA-----CAAATGCCTTCGCATTT-----
Melipsoides	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGCNCTATAT--A-TGCTT
Mdjragini	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGCNCTATAT--A-TGCTT
Mbramae	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGNTCTATAT--A-TGCTT
Mneurobius	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGCTCTATAT--A-TGCTT

Marcticus	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGNTCTATAT--A-TGCTT
Msandrae	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGNNCTATAT--A-TGCTT
Minsidiosus	ACTA--TGC-CGACCCAGGATCAGCATCGA-AGCTCTATAT--A-TGCTT
Mcerebralis	ACTA--TGC-CGACCCGGGATCAGCAT-GA-AGCTCTTTAT--A-CGCTT
Mlentisuturalis	ACTA--TGC-CGACTCGGGATCAGTTTGGT-GCTATTACCA--A-CGCTC
Mxiao	-----
Myxo.	ACTA--TGC-CCACTCGGGATCAGTTTGGT-GCTAATACTA--AACGCTC
Mportucalensis	ACTA--TGC-CAACTCGGGATCAGTTTGGT-GTTA-TACCA--A-CGCTC
Mmusculi	ACTA--TGC-CGACTAGGGATCAGCTTGGT-GATATTACAA--G---CAT
Mcyprini	ACTA--TGC-CGACTAGGGATCAGCTTGGT-GATATTACAA--G---CAT
Mpseudodispar	ACTA--TGC-CGACTAGGGATCAGCTTGGT-GATATTACAA--G---CAC
Mhibullatus	ACTA--TGC-CGACCTGGGATCAGTTTGGG-GATATTACAA--G---CTT
Melegans	ACTA--TGC-CGACCTGGGACCAGTTTGG---ATACTACAA--A---CTC
Mpendula	ACTA--TGC-CGACCTGGGATCAGCTTAGT-GAT-TTACAA--G---CTC
Mpellicides	ACTA--TGC-CGACCTGGGATCAGCTTAGT-GAT-TTACAA--G---CTC
Mhungaricus	ACTA--TGC-CGACCTGGGATCAGCTTAGT-GAT-TTGCAA--G---CAC
Myxiditruttae	ACAA--TGC-CAACCCGGGATCAGTTCGGATGATTTTGTT-----GGATC
Myxidium	ACGA--TGC-CAACCCGGGATCAGTTCGGATGATTTTATTT-----GGATC
Ethclone1AM	ACTA--TGC-CGACCCGGGATTAGTCCTAT--GCCTTATTTTACGGCTT
Ethclone3AM	ACTA--TGC-CGACCCGGGATTAGTCCTAT--GCCTTATTTTACGGCTT
Ethclone2AM	ACTA--TGC-CGACCCGGGATTAGTCCTAT--GCCTTATTTTACGGCTT
Mdiaphanus	ACTA--TGC-CGACCCGGGATTAGTCCTAT--GCCTTATTTTACGGCTT
M.lieberkuehni	ACTA--TGC-CAACCCGGGATCAGTTTTGA--GCTAAAT---AACGCTC
Myxosp.	ACTA--TGC-CGACCCGAGATCAGTTTTGA--GC-TAATTA--AACGCTC
Hennzschokkei	ACTA--TGC-CAACCCGGGATGAGTTTAGA--GC-TAATTAT-AACGCTC
Hennsalminicola	ACTCACTGCGCAACCCGGGATGAGTTTAGA--GC-TAATTAT-AACGCTC
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	ACTA--TGC-CGACCCGGGATCAGCTTGGG--GTTATATACTCA-TGCTC
Hsp.	ACTA--TGC-CGACCCGGGATCAGTTTGGG--GCAGTGTATCCA-CGCTC
Hdoori	ACTA--TGC-CGACCCGGGATCAGTTTGGG--GCAGTGTATCCA-CGCTC
Hennictaluri	ACTA--TGC-CGACCCGGGATCAGTTTGGG--GT-ATATTTCCA-CGCTC
Hennexilis	ACTA--TGC-CGACCCGGGATCAGTTTGGG--GTTATATATCCA-CGCTC
Mspinacurvatura	ACTA--TGC-CGACCCGGGATCAGTTTGGG--GCTCTAT-TCCA-TGCTC
Michkeulensis	ACTA--TGC-CGACCCGGGATCAGTTTGGG--GCTCTAT-TCCA-TGCTC
Mosburni	ACTA--TGC-CAACTCGGGATCAGTTTGGG--GCACAGTATCCA-GGCTC
Muvuliferus	ACTA--TGC-CAACTCGGGATCAGTTTGGG--GTACAATTTCCA-AGCTC
Ceratomyxa	ACTA--TGC-CAGCTTGAGATTAGCTCGGT-----AAACGAGC
Malgonquinensis	ACTA--TGC-CGACCTGGGATCAGTTTAGA-GATGTTACAA-----GCTC
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	GATGTTGGNCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Mdjragini	GATGTTGGACCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Mbramae	GATGTTGGNCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Mneurobius	GATGTTGGTCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Marcticus	GATGTTGGNCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Msandrae	GATGTTGGACCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Minsidiosus	GATGTTGGTCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Mcerebralis	TATGTTGGTCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGAGA
Mlentisuturalis	GAAGTTGATCCCCCTGGGAAA--CCTGAAGTTTTTCGG-TTACGGGGAGA
Mxiao	-----
Myxo.	GAAGTTGCTCCCCCTGGGAAA--CCTTAAGTTTTTCGG-TTACGGGGAGA

Mportucalensis	TAAGTTGGTCCCCCTGGGAAA--CCTTAAGTTTTTGGG-TTACGGGGGGA
Mmusculi	CAAGTTGGTCTCCAAGGGAAA--CCATAAGTTTTTCGG-TTACGGGGGAGA
Mcyprini	CAAGTTGGTCTCCAAGGGAAA--CCATAAGTTTTTCGG-TTACGGGGGAGA
Mpseudodispar	CAAGTTGGTCTCCAAGGGAAA--CCATAAGTTTTTCGG-TTACGGGGGAGA
Mbibullatus	CAGATTGGTCCCCCTGGGAAA--CTTCAAGTTTTTCGG-TTACGGGGGAGA
Melegans	TAGGTTGGTCCCCCTGGGAAA--CCTCAAGTCTTTAGG-TTACGGGGGAGA
Mpendula	TAGGTTGGTCTCTCTGGGAAA--CCTCAAGTTTTTCGGTTACGGGGGAGA
Mpellicides	TAGGTTGGTCTCTCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGGAGA
Mhungaricus	TAAGTTGGTCCCCCTGGGAAA--CCTCAAGTTTTTCGG-TTACGGGGGAGA
Myxiditruttae	CAGGTTGGTCCCCCTGG-AAA--CCTTGAGTTTTTAGG-TTCCGGGGGGA
Myxidium	CAGGTTGTTCCCCCTGG-AAA--CCTTGAGTTTT-AGG-TTCCGGGGGNA
Ethclone1AM	G-GGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
Ethclone3AM	G-GGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
Ethclone2AM	G-GGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
Mdiaphanus	G-GGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
M.lieberkuehni	GTGGTTGGTCCCCCTGGGAAA--CCTTAAGTTTTTAGG-TTCCGGGGGGA
Myxosp.	GAGGTTGCTCTCCTGGGAAA--CCTCAAGTTTTTAGG-TTACGGGGGGA
Hennzschokkei	TAGGTTGGTCCCCCTGGGAAA--CCTGAAGTTTTTCGG-TTGCGGGGGGA
Hennsalminicola	TAGGTTGGTCCCCCTGNGAGATACCTGAAGTTTTTCGG-TTGCGGGGGGA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	GAGGTTGGTCCCCCTGGGAAA--CCTGAAGTTTTTCGG-TTGCGGGGGGA
Hsp.	GAGGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
Hdoori	GAGGTTGGTCCCCCTGGGAAA--CCTAA-GTTTTTCGG-TTGCGGGGGGA
Hennictaluri	GAGGTTGGTCCCCCTGGGAAA--CCTGAAGTTTTTCGG-TTGCGGGGGGA
Hennexilis	GAGGTTGGTCCCCCTGGGAAA--CCTGAAGTTTTTCGG-TTGCGGGGGGA
Mspinacurvatura	GGAGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
Michkeulensis	GGAGTTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTGCGGGGGGA
Mosburni	GAGATTGGTCCCCCTGGGAAA--CCTGAAGTTTTTCGG-TTGCGGGGGGA
Muvuliferus	GAGATTGGTCCCCCTGGGAAA--CCTAAAGTCTTTTCGG-TTGCGGGGGGA
Ceratomyxa	CAAGTTGGTCTCTCCGTGAAA----ACAAGCTTTTCGGG-TTCCGGGGGGA
Malgonquinensis	TAGATTGGTCCCCCTGGGAAA--CCTAAAGTTTTTCGG-TTACGGGGGGA
Perchclone1AM	---GTTAGTC---CTTGGCAGG---TCCAAGAATTTTCACCTCTCGCTGCCA
Perchclone2AM	---GTTAGTC---CTTGGCAGG---TCCAAGAATTTTCACCTCTCGCTGCCA
Perchclone3AM	---GTTAGTC---CTTGGCAGG---TCCAAGAATTTTCACCTCTCGCTGCCA
Melipsoides	-GTATGG--NCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mdjrugini	-GTATGG--NCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mbramae	-GTATGG--NCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mneurobius	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Marcticus	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Msandrae	-GTATGG--NCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Minsidiosus	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mcerebralis	-GTATGG--TCACAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mlentisuturalis	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mxiao	-----
Myxo.	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mportucalensis	-GTATGG--TCGCAAGTCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mmusculi	-GTATGG--TCGCAAGGCT-GAAATTT--AAAGGAATTGACGGAAGGGCA
Mcyprini	-GTATGG--TCGCAAGGCT-GAAATTT--AAAGGAATTGACGGAAGGGCA
Mpseudodispar	-GTATGG--TCGCAAGGCT-GAAATTT--AAAGGAATTGACGGAAGGGCA
Mbibullatus	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Melegans	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mpendula	AGTATGG--TCGCAAGTCT-GAAACTTTAAAGGGAATTGACGGAAGGGCA

Mpellicides	-GTATGG--TCGCAAGTCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mhungaricus	-GTATGG--TCGCAAGTCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Myxiditrutae	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Myxidium	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Ethclone1AM	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Ethclone3AM	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Ethclone2AM	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mdiaphanus	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
M.lieberkuehni	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Myxosp.	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Hennzschokkei	-GTATGG--TTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Hennsalminicola	-GTATGGGGTTGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-GTATGG--TCGCAAGGCTGAAACTTT--AAAGGAATTGACGGAAGGGCA
Hsp.	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Hdooi	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Hennictaluri	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Hennexilis	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Mspinacurvatura	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Michkeulensis	-GTATGG--TCGCAAGGCTGAAACTT--AAAGGAATTGACGGAAGGGCA
Mosburni	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Muvuliferus	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Ceratomyxa	-GTACGG--TCGCAAGTCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Malgonquinensis	-GTATGG--TCGCAAGGCT-GAAACTT--AAAGGAATTGACGGAAGGGCA
Perchclone1AM	AATAC-----CAATGCC-----CCC-----AA
Perchclone2AM	AATAC-----CAATGCC-----CCC-----AA
Perchclone3AM	AATAC-----CAATGCC-----CCC-----AA
Melipsoides	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACNCAACACGGGAAAA
Mdjrugini	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACNCAACACGGGAAAA
Mbramae	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Mneurobius	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Marcticus	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Msandrae	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Minsidiosus	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Mcerebralis	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Mlentisuturalis	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Mxiao	-----
Myxo.	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Mportucalensis	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Mmusculi	CCACC-AGGGGTGGAACTGCGGCTTAATT--TGACTCAACACGGGGAAA
Mcyprini	CCACC-AGGGGTGGAACTGCGGCTTAATT--TGACTCAACACGGGGAAA
Mpseudodispar	CCACC-AGGGGTGGAACTGCGGCTTAATT--TGACTCAACACGGGGAAA
Mbibullatus	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Melegans	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Mpendula	CCACC-AGGGTGGAACTGCGGCTTAATT--TGACTTAAACACGGGGAAA
Mpellicides	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCANACACGGGGAAA
Mhungaricus	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Myxiditrutae	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Myxidium	CCACC-AGGGGTNNNACCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Ethclone1AM	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Ethclone3AM	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Ethclone2AM	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA

Mdiaphanus	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
M.lieberkuehni	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Myxosp.	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Hennzschokkei	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Hennsalminicola	CCACC-AGGAGTGGAG--TGCGGCTTAATT--TGACTCAACACGGGAAAA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	CCACCCAAGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Hsp.	CCACC-AGGGGTGGAGC--TGCGGCTTAATT--TGACTCAACACGGGAAAA
Hdoori	C-ACC-AGGGGTGGCAT--GGCGGCTTAATT--TGACTCAACACGGGAAAA
Hennictaluri	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Hennexilis	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Mspinacurvatura	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Michkeulensis	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Mosburni	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Muvuliferus	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGAAAA
Ceratomyxa	CCACC-AGGAGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGCAA
Malgonquinensis	CCACC-AGGGGTGGAGCCTGCGGCTTAATT--TGACTCAACACGGGGAAA
Perchclone1AM	CCGCT-----CCT----CTTAATCAGTACCCCAATACGG---AA
Perchclone2AM	CCGCT-----CCT----CTTAATCAGTACCCCAATACGG---AA
Perchclone3AM	CCGCT-----CCT----CTTAATCAGTACCCCAATACGG---AA
Melipsoides	CTTACCAGGNCCGGACANCAATAGGATAGACA-GACTGATAGANCT--TT
Mdjrugini	CTTACCAGGNCCGGACANCAATAGGATAGACA-GACTGATAGANCT--TN
Mbramae	CTTACCAGGNCCGGACATCAATAGGATAGACA-GACTGATAGGCTT--NN
Mneurobius	CTTACCAGGTCCGGACATCAATAGGATAGACA-GACTGATAGATCT--TT
Marcticus	CTTACCAGGTCCGGACATCAATAGGATAGACA-GACTGATAGATCT--TT
Msandrae	CTTACCAGGNCCGGACATCAATAGGATAGACA-GACTGATAGNC----TT
Minsidiosus	CTTACCAGGTCCGGACATCAATAGGATAGACA-GACTGATAGATCT--TT
Mcerebralis	CTTACCAGGTCCGGACATCAATAGGATAGACAAGACTGATAGATCT--TT
Mlentisuturalis	CTCACCCGGTCCGGACATCGAAAGGATAGACA-GATTGATAGATCT--TT
Mxiao	-----
Myxo.	CTCACCCGGTCCGGACATCGAAAGGATAAACA-GATTAA-AGGTCT--TT
Mportucalensis	CTCACTTGGGCAAGATGTCAAAAGGATGGCCA-GACTGA-AGTTCT--TG
Mmusculi	CTTACATGGTCCAGACATCGATAGGATAAACA-GACTGATAGATCT--TT
Mcyprini	CTTACATGGTCCAGACATCGATAGGATAAACA-GACTGATAGATCT--TT
Mpseudodispar	CTTACATGGTCCAGACATCGATAGGATAAACA-GACTGATAGATCT--TT
Mbibullatus	CTTACCTGGTCCGGACATCGATAGGATAGACA-GACTGATAGATCT--TT
Melegans	CTTACCTGGTCCGGACATCGATAGGATAGACA-GACTGATAGATCT--TT
Mpendula	CTTACTCGGTCAGGACATCGAAAGGATAGACA-GACTGATAGATCT--TT
Mpellicides	CTTACTCGGTCAGGACATCGAAAGGATAGACA-GACTGATAGATCT--TT
Mhungaricus	CTTACCTGGTCCGGACATCGAAAGGATAGACA-GACTGATAGATCT--TT
Myxiditruttae	CTCACCTGGTCCGGACATCGATAGGATTGACA-GACTAATAGATCT--TT
Myxidium	CTCACCTGGTCCGGACATCGATAGGATTGACA-GACTAATAGATCT--TT
Ethclone1AM	CTTACCGGATCAGGACATCAAAAGGATTGTCA-GACCGA-AGATCT--TA
Ethclone3AM	CTTACCGGATCAGGACATCAAAAGGATTGTCA-GACCGA-AGATCT--TA
Ethclone2AM	CTTACCGGATCAGGACATCAAAAGGATTGTCA-GACCGA-AGATCT--TA
Mdiaphanus	CTTACCGGATCAGGACATCAAAAGGATTGTCA-GACCGA-AGATCT--TA
M.lieberkuehni	CTCACCCGGTCCGGACATCGATAGGATCGACA-GACTGATAGATCT--TT
Myxosp.	CTCACCTGGTCCGGACATCGAAAGGATAGACA-GACTGATAGATCT--TT
Hennzschokkei	CTTACTCGGTTCCGGACATTGACAGGATAAACA-GTTTGATAGAACT--TT
Hennsalminicola	CTTACTCGGTTCCGGACATTGACAGGATAAACA-GTTTGATAGNACT--TT
Ethclone2MA	-----
Ethclone3MA	-----

Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	CTTACCTGGTCCGGACATCCGAAGGATACTCA-GACCTAAGATCT---T
Hsp.	CTTACCTGGTCCGGACATCGAAAAGGATAAAACC-GACCAAAAAGATC---GT
Hdoori	CTTACCTGGTCCGGACATCGAAAAGGATAAAACC-GACCAAAAAGATC---GT
Hennictaluri	CTTACCTGGTCCGGACATCAAAAAGGATAGACA-GACTGCCAGATC---TT
Hennexilis	CTTACCTGGTCCGGACATCAAAAAGGATAGACA-GACTGCCAGATC---TT
Mspinacurvatura	CTTACCTGGTCCGAACATCCGAAGGATACACA-GACTAATAGATC---TT
Michkeulensis	CTTACCTGGTCCGAACATCCGAAGGATACACA-GACTAATAGATC---TT
Mosburni	CTTACCTGGTCCGGACATAAAAAGGAGTTACC-GAGTGCC-GCTCGAATT
Muvuliferus	CTTACCTGGTCCGGACATAAAAAGGAGTTACC-GAGTGCC-GCTCGAATT
Ceratomyxa	CTCACCAGGTCCGGACATTGAAAGGATTGACA-GACTGATAGATCT---T
Malgonquinensis	CTTACCTGGTCCGGACATCGATAGGATAAAACA-GATCAATAGCTCT---T
Perchclone1AM	CC-----AACATCG-----
Perchclone2AM	CC-----AACATCG-----
Perchclone3AM	CC-----AACATCG-----
Melipsoides	-CTTGATATGATGGATAGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mdjragini	-CTTGATATGATGGATAGTGGTGCATGGCC-GTTCTTAGTTTGNNGGAGTG
Mbramae	-CTTGATATGATGGATAGTGGGGCATG-CC-CTTTTTAATTCGGGGAG-G
Mneurobius	TCTTGATATGATGGATAGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Marcticus	-CTTGATATGATGGATAGTGGTGCATG-CC-GTT-TTAGTTTCGTGG-GTG
Msandrae	ACTTGATATGATGGATAGTG-CGCANN-CC-TTTCNNATTTTCGGGGAGAN
Minsidiosus	-CTTGATATGATGGATAGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mcerebralis	-CTTGATATGATGGATAGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mlentisuturalis	-CTTGATACGGTGATTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mxiaoi	-----
Myxo.	-TTTGATACGGTGATTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mportucalensis	-CCAGATATGACAAAAGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Mmusculi	-TTTGATGCGGTGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mcyprini	-TTTGATGCGGTGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mpseudodispar	-TTTGATGCGGTGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mbibullatus	-CTTGATGCGATGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Melegans	-CTTGATGCGATGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mpendula	-CTTGATGCGGTGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Mpellicides	-CTTGATGCGGTGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Mhungaricus	-CTTGATGCGGTGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGCG
Myxiditruttae	-CATGATACGGTGGGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGCG
Myxidium	-CATGATACGGTGGGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Ethclone1AM	-CATGATCTGGTGAAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Ethclone3AM	-CATGATCTGGTGAAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Ethclone2AM	-CATGATCTGGTGAATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Mdiaphanus	-CATGATCTGGTGAATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
M.lieberkuehni	-CGTGATACGGTGATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Myxosp.	-CTTGATACGGTTGGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Hennzschokkei	-TTTGATACGGTGAAATGGTGGTGCATGGCCCGTTTTTAGTTTCGTGGAGTA
Hennsalminicola	-TTTGATACGGTGAAATGGTGGTGCATGGCC-GTTTTTAGTTTCGTGGAGTA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	ATTTGATCTGGTGAATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGAAGTG
Hsp.	TTTTGATACGATGAATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Hdoori	TTTTGATACGATGAATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Hennictaluri	TCTTGATATGGTGAATGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG

Hennexilis	TCTTGATATGGTGATTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mspinacurvatura	TGTTGATGGGGTGAAAAAGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Michkeulensis	TGTTGATGGGGTTGAAAAAGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Mosburni	TAACGATATTATGAGTAGTGGTGCATGGCC-GTTCTTAGTTGCGTGGAGTG
Muvuliferus	TAACGATATTATGAGTAGTGGTGCATGGCC-GTTCTTAGTTGCGTGGAGTG
Ceratomyxa	TCATGATACAGTGATTGGTGGTGCATGGCC-GTTCTTAGTTGGTGGAGTG
Malgonquinensis	TTTTGATCCGATGAGTGGTGGTGCATGGCC-GTTCTTAGTTTCGTGGAGTG
Perchclone1AM	---TAAT-TGGCGGTTCGGTGTTCATA-----TTGAAG---
Perchclone2AM	---TAAT-TGGCGGTTCGGTGTTCATA-----TTGAAG---
Perchclone3AM	---TAAT-TGGCGGTTCGGTGTTCATA-----TTGAAG---
Melipsoides	ATCTGTCAGGCTAATCCCGGT--AACG-AACGAGATCTTATTCTCCATTT
Mdjragini	ATTTGTCAGGCTAATCCCGGT--AACG-NACGAGANTTTATTTTCCATTT
Mbramae	ATN-GTCAGG-TAATCCCGGT--AACG-AACGAGATNTTATTNTCCATTT
Mneurobius	ATCTGTCAGGCTAATCCCGGT--AACG-AACGAGATCTTATTCTCCATTT
Marcticus	ATCTGTCAGG-TAATCCCGGT--AACGGAACGAGAA-TTATTCTCCATTT
Msandrae	ATCTGTCAGGNANATCCCGGA--ANCG-AACGANATCTTATTCTCCATNA
Minsidiosus	ATCTGTCAGGCTAATCCCGGT--AACG-AACGAGATCTTATTCTCCATTT
Mcerebralis	ATCTGTCAGGCTAATCCCGGT--AACG-AACGAGATCTTATTCTCCATTT
Mlentisuturalis	ATCTGTCAGGCCAATTCGGGT--AACG-AACGAGACCACAATCTTCATTT
Mxiao	-----
Myxo.	ATCTGTCAGGCCATATCCCGGT--AACG-AACGAGACCACAATCTTCATTT
Mportucalensis	ATTTGTCGGGTCGATTCCGAT--AACG-GACGAGACTGCATTCTCCATTT
Mmusculi	ATCTGTCAGGTTTATTCCGGT--AACG-AACGAGACTACAACCTTCATTT
Mcyprini	ATCTGTCAGGTTTATTCCGGT--AACG-AACGAGACTACAACCTTCATTT
Mpseudodispar	ATCTGTCAGGTTTATTCCGGT--AACG-AACGAGACTACAACCTTCATTT
Mbibullatus	ATCTGTCAGGTTTATTCCGGT--AACG-AACGAGACCACATTCTCCATTT
Melegans	ATCTGTCAGGTTAATTCGGGT--AACG-AACGAGACCACAGTCCCCATTT
Mpendula	ATTTGTCAGGTCGATTCCGGT--AACG-GACGAGACCACTGTCTCCATTT
Mpellicides	ATTTGTCAGGTCGATTCCGGT--AACG-GACGAGACCACTGTCTCCATTT
Mhungaricus	ATCTGTCAGGTTTATTCCGGT--AACG-AACGAGACCACCTTCTCCATTT
Myxiditruttae	ATCTGTCAGGTTTATTCCGGT--AACG-AACGAGACCACGTTCTTCATTT
Myxidium	ATCTGTCAGGTTGATTCCGGT--AACG-AACGAGACCATTTGTCTTCATTT
Ethclone1AM	ATTTGTCAGGTTTATTCCGGT--AACG-GACGAGATCACAATCTCCATTT
Ethclone3AM	ATTTGTCAGGTTTATTCCGGT--AACG-GACGAGATCACAATCTCCATTT
Ethclone2AM	ATTTGTCAGGTTTATTCCGGT--AACG-GACGAGATCACAATCTCCATTT
Mdiaphanus	ATTTGTCAGGTTAATTCGGGT--AACG-GACGAGATCACAATCTCCATTT
M.lieberkuehni	ATTTGTCAGGTTTATTCCGGT--AACG-AACGAGACCATATTCTCCATTT
Myxosp.	ATCTGTCAGGTTGATTCCGGT--AACG-GACGAGACCATATTCTCCATTT
Hennzschokkei	ATCTGTCAGGCTAACCCCGGT--AACG-AACGAGATCAGCGTCTCCATTT
Hennsalminicola	ATCTGTCAGGCTAACCCCGGT--AACG-AACGAGATCAGCGTCTCCATTT
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	ATTTGTCAGGTTTATTCCGGT--AACG-GACGAGACTGCGTTCTCCATTT
Hsp.	ATCTGTCAGGCTAATCCCGGT--AACG-AACGAGACCACATTCTCCATTT
Hdoori	ATCTGTCAGGCTAATCCCGGT--AACG-AACGAGACCACATTCTCCATTT
Hennictaluri	ATCTGTCAGGCCAATCCCGGT--AACG-AACGAGACCACATTCTCCATTT
Hennexilis	ATCTGTCAGGCCAATTCGGGT--AACG-AACGAGACCATTCTCCATTT
Mspinacurvatura	ATCTGTCAGGCCAATTCGGGT--AACG-AACGAGACTACGTTCTCCATTT
Michkeulensis	ATCTGTCAGGCCAATCCCGGT--AACG-AACGAGACTACAGTCTCCATTT
Mosburni	ATCTGTTTCCCTTAT--CGGTTAAACG-CGCGAGATGCCGGTCTTCATTT
Muvuliferus	ATCTGTCAGGTTTATTCCGGT--AACG-AGCGAGACCACGATCTCTATTT
Ceratomyxa	ATCTGTCAGGTTTATTCCGGT--AACG-AGCGAGACCACGATCTCTATTT
Malgonquinensis	ATCTGTCAGTTTAAATTACGGT--AACG-AACGAGACCACAGTCCCCATTT

Perchclone1AM	-----	
Perchclone2AM	-----	
Perchclone3AM	-----	
Melipsoides	-----	GATGAGC
Mdjragini	-----	GATGAGC
Mbramae	-----	GATGAGC
Mneurobius	-----	GATGAGC
Marcticus	-----	GATGAGC
Msandrae	-----	GATGAGC
Minsidiosus	-----	GATGAGC
Mcerebralis	-----	GATGAGC
Mlentisuturalis	-----	GAGAAGC
Mxiaoi	-----	
Myxo.	-----	GAGTAGC
Mportucalensis	-----	GGTGAGC
Mmusculi	-----	AAGAGGC
Mcyprini	-----	AAGAGGC
Mpseudodispar	-----	AAGAGGC
Mbibullatus	-----	AAGGCAT
Melegans	-----	AAGAAAC
Mpendula	-----	AAGAATC
Mpellicides	-----	AAGAATC
Mhungaricus	-----	AGGATAC
Myxiditruttiae	-----	GGGGAAC
Myxidium	-----	GGGGAAC
Ethclone1AM	-----	GACGAAC
Ethclone3AM	-----	GACGAAC
Ethclone2AM	-----	GACGAAC
Mdiaphanus	-----	GACGAAC
M.lieberkuehni	ACGGAGTCGATGCAGCATCAGCCTGCCAGGGGCAACTCTGTCAGACGAAG	
Myxosp.	-----	GAGGAA-
Hennzschokkei	-----	GGGGTG
Hennsalminicola	-----	GGGGTG
Ethclone2MA	-----	
Ethclone3MA	-----	
Ethclone1MA	-----	
Myxobprocerus	-----	
Mprocerus	-----	
Henneguya	-----	GACGAGC
Hsp.	-----	GACGAGC
Hdoori	-----	GACGAGC
Hennictaluri	-----	GACGAGC
Hennexilis	-----	GACGAGC
Mspinacurvatura	-----	GACGCGC
Michkeulensis	-----	GACGCAC
Mosburni	-----	GATGAGT
Muvuliferus	-----	GATGAGT
Ceratomyxa	-----	-TC
Malgonquinensis	-----	GAGAGAT
Perchclone1AM	-----	
Perchclone2AM	-----	
Perchclone3AM	-----	
Melipsoides	GGAA-GCAGATGGTG-----	
Mdjragini	GGAA-GCAGATGGTG-----	
Mbramae	GGAA-GCAGATGGTG-----	

Mneurobius	GGAA-GCAGATGGTG-----
Marcticus	GGAA-GCAGATGGTG-----
Msandrae	GGAA-GCAGATGGTG-----
Minsidiosus	GGAA-GCAGATGGTG-----
Mcerebralis	GGAA-GAAGATAGTGT-----
Mlentisuturalis	AGCAGAATTCTGA-----
Mxiao	-----
Myxo.	AACAGAATTCAA-----
Mportucalensis	AAAAGATATTGC-----
Mmusculi	AGAA-GCAGGT-----
Mcyprini	AAAA-GCAGGT-----
Mpseudodispar	AAAA-GCAGGT-----
Mbibullatus	GGAA-GTAGGCGGCTGGATTAAATACTTTAGGGTATTGATTTGGTTGCTGG
Melegans	GAAA-GTAGAG-----
Mpendula	AGTAAGCAAAAGG-----
Mpellicides	AGTA-GCAAAAGG-----
Mhungaricus	GGTA-GCAGGAGG-----
Myxiditruttae	CAG---AGGACAGCTA-----TTGTG-----
Myxidium	TAT---AAGAC-----TTGTG-----
Ethclone1AM	TG---GAGTAAAT-----GCA-----
Ethclone3AM	TG---GAGTAAAT-----GCA-----
Ethclone2AM	TG---GAGTAAAT-----GCA-----
Mdiaphanus	TG---GAGTAAAT-----GTA-----
M.lieberkuehni	TGCGACGATTAAATTGGTTGTTTAGCA-----
Myxosp.	TGCAAAACTCAGT-----CA-----
Hennzschokkei	TACAAGAAATCTATCGGTAGTGCTGCATT-----
Hennsalminicola	TACAAGAAATGTATCGGTAGTGCTGCATT-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	TGAAGTTTGTGCGTTC-----
Hsp.	CGTAGAAGGCGTTGGC-----
Hdoori	CGTAGAAGGCGTTGGC-----
Hennictaluri	CGGAGAAGGCCGTGGT-----
Hennexilis	CGGAGAAGGCGTTGGT-----
Mspinacurvatura	CAAAGAACGCACT-----
Michkeulensis	TGAAGAATGCAGG-----
Mosburni	CAGAGAGGGCGTGGAT-----
Muvuliferus	CAGAGATGGCGTCGAT-----
Ceratomyxa	TTTCAGTAGCAT-----
Malgonquinensis	GAAAGTAGGCGTCTGA-----
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----GCTTGAAAAATTGTCT-CG-----ATGA
Mdfragini	-----GCTTGAAAAATTGTCTTCG-----ATGA
Mbramae	-----GCTTGAAAAATTGTCT-CG-----ATGA
Mneurobius	-----GCTTGAAAAATTGTCT-CG-----ATGA
Marcticus	-----GCTTGAAAAATTGTCT-CG-----ATGA
Msandrae	-----GTTTGAAAAATTGTCT-CG-----NNGA
Minsidiosus	-----GCTTGAAA-TTGTCT-CG-----ATGA
Mcerebralis	-----AGCTCGATGATTGTTT-CG-----GCCA
Mlentisuturalis	-----CAAACGATTGCCG-----
Mxiao	-----

Myxo.	-----CAAGCACTCTCTGAT-----	
Mportucalensis	-----CACGTG-TCACCGGTA-----	ACGG
Mmusculi	-----TTCGCGATTGACTGTG-----	CTTCG-
Mcyprini	-----TATGCGAGAG--GTG-----	CTTCGC
Mpseudodispar	-----TTCGCGATTGACTATA-----	CTT---
Mbibullatus	TGGTAAAATTAGTGATCCATTGTAAATTTAGCAAAGGGAGCAATCTTGAT	
Melegans	-----TGTGCAAGAACGGGTA-----	TTTCG-
Mpendula	-----CTGGTGTGGTACCGC-----	TTTT-
Mpellicides	-----CTGGTGTGGTACCGC-----	TTTT-
Mhungaricus	-----CCAG---GAGGCGGT-----	TTTCG-
Myxiditrutiae	---GG--TTGTAGTGTTTTGAATA-----	AGTGG
Myxidium	---GT--TTATATTGTGTGTTTATC-----	ATTGA
Ethclone1AM	---A-----ATAGTCATAATTA-----	ATTG-
Ethclone3AM	---A-----ATAGTCATAATTA-----	ATTG-
Ethclone2AM	---A-----ATAGTCATAATTA-----	ATTG-
Mdiaphanus	---A-----AGCATC-TATGTG-----	TTTGG
M.lieberkuehni	---GACTTGTATTGCTGAATTTTCCTTTTTTC-----	CATTGA
Myxosp.	---G-----GGTGTGTTAGTTC-----	GC
Hennzschokkei	---GTTTCGACAGTGTACATTTA-----	TTGA
Hennsalminicola	---GTTTCGACAGTGTACATTTA-----	TTGA
Ethclone2MA	-----	
Ethclone3MA	-----	
Ethclone1MA	-----	
Myxobprocerus	-----	
Mprocerus	-----	
Henneguya	-----GCACACAGTCATTGGCAACGGTGACTGCGTTGTGACCGTATGG	
Hsp.	-----TATGAAAGTTTTTCTTCGGGAGAGAGTTTGT-----	GGTCA
Hdoori	-----TATGAGTGCTCTC-----GGAGAGAGTTTGT-----	GGTCA
Hennictaluri	-----AATGGAAATTTTTGTGTTACAAAAGTTTCTGC-----	TATCA
Hennexilis	-----ATTGAGGATTTTTGCTTGCAAGGGTCCCCGG-----	TGCTA
Mspinacurvatura	-----	
Michkeulensis	-----	
Mosburni	-----CAGTTCAATTTTTGGCAACAAGAGTTGTTCT-----	TTTCT
Muvuliferus	-----CAGTTCAATTTTTGGCAACAGAGATTGTACT-----	TTTTG
Ceratomyxa	-----TTGTCGTTTC-----	
Malgonquinensis	-----TTCACTCTGCCTCACAGTGGAGCGA-----	ATCGG
Perchclone1AM	-----	
Perchclone2AM	-----	
Perchclone3AM	-----	
Melipsoides	AA-----	TTCAA
Mdjragini	AA-----	TTCAA
Mbramae	AA-----	TTCAA
Mneurobius	AA-----	TTCAA
Marcticus	AA-----	TTCAA
Msandrae	AA-----	TTCAA
Minsidiosus	AA-----	TYCAA
Mcerebralis	TT-----	CTCAA
Mlentisuturalis	-----	G
Mxiao	-----	
Myxo.	-----	G
Mportucalensis	TG-----	TATGG
Mmusculi	-----GTG-----CTTTC-----	TTTCG
Mcyprini	G-----GTG-----CCTTG-----	TT-CG
Mpseudodispar	-----GTG-----TAGTT-----	TTTCG
Mbibullatus	GCGAAGATTTATGTGGGGTAATTAGTTTTCCCTTCCATTTAGCAGTTTCT	
Melegans	-----GTA-----TTTGT-----	TTTTG

Mpendula	-----GTAG-----TATC-----TCATT
Mpellicides	-----GTAG-----TATC-----TCATT
Mhungaricus	-----GCCG-----TTTC-----CC--T
Myxiditruttae	-----TGAGC
Myxidium	-----TGAGA
Ethclone1AM	-----TAAAA
Ethclone3AM	-----TAAAA
Ethclone2AM	-----TAAAA
Mdiaphanus	-----TAAAA
M.lieberkuehni	CTAA-----GCACTGTGTAGCCTCAAA
Myxosp.	-----TAATA
Hennzschokkei	TAG-----TTAGA
Hennsalminicola	TAG-----TTATA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	CTG-----CCGGT
Hsp.	ACG-----TCGAA
Hdoori	ACG-----TCGAA
Hennictaluri	TTG-----TTGAA
Hennexilis	ATT-----CTGTA
Mspinacurvatura	-----
Michkeulensis	-----
Mosburni	GCG-----TCGTA
Muvuliferus	GCG-----TTGCA
Ceratomyxa	-----
Malgonquinensis	CG-----CTGTA
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	GTTAT-C-----ATCGAAGGC-----
Mdjragini	GTTAT-C-----ATCGAAGGC-----
Mbramae	GTTAT-C-----ATCGAAGGC-----
Mneurobius	GTTAT-C-----ATCGAAGGC-----
Marcticus	GTTAN-C-----ATCGAAGGC-----
Msandrae	GTTAT-CT-----ATNGAAGGC-----
Minsidiosus	GTTAT-C-----ATCGAAGGC-----
Mcerebralis	GTTATTCT-----ATCGTAGGC-----
Mlentisuturalis	CATGG-----TAAAA--C-----
Mxiao	-----
Myxo.	GGTGG-----CAACA--C-----
Mportucalensis	GGTAA-----AAATAGGC-----
Mmusculi	TGTCT-----CTGTAAGA-----
Mcyprini	TGTCT-----CTGTAAGA-----
Mpseudodispar	TGTCT-----CTTCAAGA-----
Mbibullatus	TACCT-----CGGTAGGACGCTGTCTTATGGAGAGA-----
Melegans	GATAC-----TTGTAGGT-----
Mpendula	AGCCT-----TTATAGGT-----
Mpellicides	AGCCT-----TTATAGGT-----
Mhungaricus	GGTCT-----TTGCAGGT-----
Myxiditruttae	AATC-----ATTGTGAG-----
Myxidium	AATCTCG-----GTTTTAAG-----
Ethclone1AM	GATTG-----ATTGTGA-----
Ethclone3AM	GATTG-----ATTGTGA-----

Ethclone2AM	GATTG-----ATTGTGA-----
Mdiaphanus	CAAAG-----ATGTAGA-----
M.lieberkuehni	AACTGCATAGTGCG--CTATGGAGAGACAACCGGGTATATCCAAAGCCGG
Myxosp.	TGTTCTG-----TCTGTAGC-----
Hennzschokkei	GGCTGTGTGGGTGTGCTGTTGCCGATGGTAATGT-----
Hennsalminicola	GGCTGTGTGAGTGTGCTGTTGTTGATGGTTGTAC-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	GGTGGTGGGCGT----TGTGTGCGGCT-----
Hsp.	GGTTTAATGAGA----CGGGGAGGTGCC-----
Hdoori	GGTTTAATGAGA----CGGGGAGGTGCC-----
Hennictaluri	GGCAT-----CGTATAGTTGCC-----
Hennexilis	GGTAC-----TGTGGGGAAGCT-----
Mspinacurvatura	-----CGTAGGG-----
Michkeulensis	-----CGTACGG-----
Mosburni	GGCAATGTCCG----TTTGGTGAGTTT-----
Muvuliferus	GGCAATGTCCG----TGTGGTGTGGTT-----
Ceratomyxa	-----TACTGA-----
Malgonquinensis	GGCAT-----AGTTGTGGAA-----
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----AGTGCTTGCGAGTTTATTGT--
Mdjragini	-----AGTGCTTGCGAGTTTATTGT--
Mbramae	-----AGTGCTTGCGAGTTTATTGT--
Mneurobius	-----AGTGCTTGCGAGTTTATTGT--
Marcticus	-----AGTGCTTGCGAGTTTATTGT--
Msandrae	-----AGTGCTNCGAGTTTATTGTG--
Minsidiosus	-----AGTGTTTTCGAGTTTATTGT--
Mcerebralis	-----AGTGTTTGTGAATTTAGTGT--
Mlentisuturalis	-----ATGTCGGTATATTGTTT--
Mxiao	-----
Myxo.	-----TTATCAG-AGGGTGTTT--
Mportucalensis	-----AGATTGGGCGCGCAATT--
Mmusculi	-----AAGAATATGCGAGTTC-----
Mcyprini	-----A-GAATATGCAAATTC-----
Mpseudodispar	-----A-GAATATTCAAGTTC-----
Mbibullatus	-----CAACGGGGAATATATAAGCTCGAGGAAG
Melegans	-----ATA-GCTGGTTAGC-----
Mpendula	-----AAAAT-ATCAAATTCCTG-----
Mpellicides	-----AAAAT-ATCAAATTCCTG-----
Mhungaricus	-----AAAATTATTGAATTTCA-----
Myxiditruttae	-----TTTGACTGCT-----
Myxidium	-----TTTGTGATTT-----
Ethclone1AM	-----TATGTATT-----
Ethclone3AM	-----TATGTATT-----
Ethclone2AM	-----TATGTATT-----
Mdiaphanus	-----TG-AAATT-----
M.lieberkuehni	AGGACGTATGGCAATAACAGGTCTGTGATGCCCTTAGATGTTGGGGCCG
Myxosp.	-----TGGTAGTT-----
Hennzschokkei	-----CAACTCGTTAAAAGGTTG-----
Hennsalminicola	-----CAATTGGTTTACTAGTTG-----
Ethclone2MA	-----

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Ethclone3MA -----
Ethclone1MA -----
Myxobprocerus -----
Mprocerus -----
Henneguya -----CAAG---GTTACCTTTTCGGG--
Hsp. -----GAGA---TGTTTGATGGGAAG--
Hdoori -----GAGA---TGTTTGATGGGAAG--
Hennictaluri -----AAGGGTGATTTGGTTGAGGTG--
Hennexilis -----CAGGGGGCATTTGGCTGAGATC--
Mspinacurvatura -----
Michkeulensis -----
Mosburni -----AAGAGTGATCT-CTGTTTCG--
Muvuliferus -----GAAAAATGTGAC--CTGGCTCG--
Ceratomyxa -----
Malgonquinensis -----GGAATTTGGTGCAGTGGGGTA--
Perchclone1AM -----
Perchclone2AM -----
Perchclone3AM -----

Melipsoides -----TGAAATATAAAAGAGTTGCGAGAACGGA-CTTAACCCCAT--
Mdjragini -----TGAAATATAAAAGAGTTGCGAGAACGGN-CTTAACCCCAT--
Mbramae -----TGAAATATAAAAGAGTTGCGAGAACGNTCTTAACCCCAT--
Mneurobius -----TGAAATATAAAAGAGTTGCGAGAACGGT-CTTAACCCCAT--
Marcticus -----TGAAATATAAAAGAGTTGCGAGAACGGA-CTTAACCCCAT--
Msandrae -----TGAAATATAAAAGAGTNGCGAGAACGTT-CTAAACCCCAT--
Minsidiosus -----TGAAATATAAAAGAGTTGCGAGAACGGT-CTTACCCCCATA-
Mcerebralis -----GAAATACAGTTTGTGCGAGGACGGG-ATAAACTCTT--
Mlentisuturalis -----CTTGAGAGGCAGAAACGTACC-TCTGCTGTGA---
Mxiaoi -----
Myxo. -----CTTGAGAAAGCAGAAAAGCGGC-TTGG--GTCG---
Mportucalensis -----GCTAGAGATTTGGTTTGCANA-CTTGTTTFTTGAG-
Mmusculi -----GCAGTGAT---CAGGGTGAT-C-GGGCAAC-CG-
Mcyprini -----TCAGAAAG---CTGGGCAAA-C-GGGCAAC-CG-
Mpseudodispar -----TCAATTGG---TTGGGTTTT-C-AGGCAAC-TG-
Mbibullatus AGTGGCTATAACAGGTCAGTGATGCCCTTCGATGTT-CAGGGCTGCACG-
Melegans -----TCAGTAGG---CTGTTGCAGG-AGGGGTAACCCG-
Mpendula -----CAGGGAA---AAGTATGAA-T-GGGCAACCAA-
Mpellicides -----CAGGGAA---AAGTATGAA-T-GGGCAACCAA-
Mhungaricus -----T-GGGTG---CGATGCGGT-G-GGGCAACTCG-
Myxiditruttae -----ATACTT---GCGG-----TAGTTGCACC-
Myxidium -----ACAATG---GTGGATGC---CATTTGCTCT-
Ethclone1AM -----TGCATTG-----TTGGTAA---AATGTG-----
Ethclone3AM -----TGCATTG-----TTGGTAA---AATGTG-----
Ethclone2AM -----TACATTG-----TTGGTAA---AATGTG-----
Mdiaphanus -----TACGTTG-----TGGGTAA---AGTGTG-----
M.lieberkuehni CACG----CGCGCTACAATGGTAACAACAGATAG---AGTCTGGTTC-
Myxosp. -----ACTG-----AATAACTGG---ATTTTTT-----
Hennzschokkei -----ATATTGCTTAAGGTGGCTGTT--TAGCTTGCACTC
Hennsalminicola -----GTGTGGCTTAAAGTGGCTGCT--TAGCTTGCACTC
Ethclone2MA -----
Ethclone3MA -----
Ethclone1MA -----
Myxobprocerus -----
Mprocerus -----
Henneguya -----GTGCCTTTCAGTCGTAAGGATGGCGTCGCATGCTGCT----
Hsp. -----GCAAC-----GACTT---GCGGACTACAAGGTAATGAC-
Hdoori -----GCAAC-----GACTT---GCGGACTACAAGGTAATGAC-

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Hennictaluri	-----GCAACATCAAGGCTG-----ATGCATT-TGAGGTAAT-----
Hennexilis	-----GCAA-GTCAAGGCTG-----GTGCACT-TTGAGTTAT-----
Mspinacurvatura	-----TT-----G-AAATC-----GGTAAA-----
Michkeulensis	-----TT-----GCGGATC-----GGTAAA-----
Mosburni	-----GCAGAGTGATATTTTGAGAGCGCATC--AAGATTGG-----
Muvuliferus	-----CCAG--TTAGCATCGTAGAACGCATC--AAGGCTGG-----
Ceratomyxa	-----GAATAGA-----
Malgonquinensis	-----ACTTGCTGTGCCTTTTTTTTGACACG--CGGCTCTCCCT--
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----TCTGGTAGCAAT-----TTGT
Mdjragini	-----TCTGGTAGCAAT-----TTGT
Mbramae	-----TCTGGTAGCAAT-----TTGT
Mneurobius	-----TCTGGTAGCAAT-----TTGT
Marcticus	-----TCTGGTAGCAAT-----TTGT
Msandrae	-----TCCGGNAGCAAT-----TGGT
Minsidiosus	-----TCTGGTAGCAAT-----TGGT
Mcerebralis	-----ACTTGTTGCAAA-----TTGT
Mlentisuturalis	-----TTGCTTCACGG-----TGGT
Mxiao	-----
Myxo.	-----TGAGTGGCAAC-----ATTT
Mportucalensis	-----CTGAATTGAAGG-----TGAT
Mmusculi	-----T-GAACT-CTGTGAT-----CTGA
Mcyprini	-----G-GAGCT-CAGTGAT-----TTGA
Mpseudodispar	-----G-CGACT-CAATCCG-----TTG-
Mbibullatus	-----C-GCGCTACAATGATGACGACAGCAAGTATCTGG
Melegans	-----A-ATGTGGCTGT-----CTGC
Mpendula	-----ACCTACT-TTAT-----CTCT
Mpellicides	-----ACCTACT-TTAT-----CTCT
Mhungaricus	-----CTGTGTT-GTAT-----CT--
Myxiditruttae	----TATGG----TGTG-CGTACAG-----C
Myxidium	----TTTGG----TGTG-CGTACAG-----A
Ethclone1AM	----AAAGA----TATA-ATCATAA-----T
Ethclone3AM	----AAAGA----TATA-ATCATAA-----T
Ethclone2AM	----AAAGA----TATA-ATCATAA-----T
Mdiaphanus	----GAGGA----GACA-AGGA-AA-----G
M.lieberkuehni	----GAAAGAATCAGGTA-ATCATAAATTG-----TTACCGT
Myxosp.	-----TCTTTT-ATTGTGA-----C
Hennzschokkei	CCTTGTACCAAACAGTATACAGTGTA-----
Hennsalminicola	CCTTGTACCAAACAGTATACAGTGTA-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-----GGCAAGCCTTCAGCCA-----GAC
Hsp.	-----TTGCGAGTTCCCTACGGTCG-----AAC
Hdoori	-----TTGCGAGTTCCCTACGGTCG-----AAC
Hennictaluri	-----CTGTGCGCTCCCTCCGGCTT-----AGC
Hennexilis	-----CTCCACAAATCCCTCCGGTTT-----AAC
Mspinacurvatura	-----ACGGTT-----A-C
Michkeulensis	-----ACGGTA-----AGC
Mosburni	-----CATTCCTTTGACGT-----TAC
Muvuliferus	-----CATTCCTCTGACGT-----TAC
Ceratomyxa	-----GAGACAAC-----T

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Malgonquinensis -----TTCATCACACAGAGACTG-----GCCTCG
Perchclone1AM -----
Perchclone2AM -----
Perchclone3AM -----

Melipsoides      ACTAAATGTA----AATTTGTTGGCATAAC---CCTTC-----
Mdjragini        ACTAAATGTA----AATTTGTTGGCATTTC---CCTTC-----
Mbaramae         ACTAAATGTA----AATTTGTTGGCATTTC---CCTTC-----
Mneurobius       ACTAAATGTA----AATTTGTTGGCATTTC---CCTTC-----
Marcticus        ACTAAATGTA----AATTTGTTGGCATNC---CCTTC-----
Msandrae         ACTAAANGNA----AATTGGTGGGCATTTC---CCTTC-----
Minsidiosus      ACTAAATGTA----AATTTGNNGGCATTTC---CCTTC-----
Mcerebralis      ACTACACCTG----AGTTTGTTGGCATTTC---CCTTC-----
Mlentisuturalis TGCAATA-----GAGGGCACCGATCTC---CATG-----
Mxiao            -----
Myxo.            CATGGCC-----CANGCTGCTGATCTC---TATG-----
Mportucalensis  TGTGCTT-----AGC-----TTGTTCTC---CCTT-----
Mmusculi         --TGGAA-----GAA--GTGTATTTTC---TCTT-----
Mcyprini         --TGGAG-----GA--GTGTATTTTC---TCTT-----
Mpseudodispar   --TGGAA-----TGC--GGGTATTTTC---TCTT-----
Mbibullatus      GTTGAAA-----AACTTGGGTAATCTCGAATCGTCATC-----GTGA
Melegans         --TGGGT-----CA--GCCAATTCTC---CCTT-----
Mpendula         TGTGAA-----GGA--GATTGTTTTTC---CCTA-----
Mpellicides      TGTGAA-----GGA--GATTGTTTTTC---CCTA-----
Mhungaricus      -GTGAAA-----GGA--TATGGTTTTTC---CCTA-----
Myxiditruttae    AATGTGCGTC--CCTATGGAGAGACAGCCGGATTGTAAG-----
Myxidium         AATGTATGTC--CCTATGGAGAGACAGCCGGATTGTAAG-----
Ethclone1AM      GATGAA-ATA--TAGG----GTAAAACTATATTGAAT-----
Ethclone3AM      GATGAA-ATA--TAGG----GTAAAACTATATTGAAT-----
Ethclone2AM      GATGAA-ATA--TAGG----GTA-----
Mdiaphanus       GAGAGG-GAG--TGTG----GCAACATGCAACTTCGAA-----
M.lieberkuehni   AATGGG-GAC--TGTGCTTTGTAATTATCGCACACGAAAG-----AGGA
Myxosp.          GTTGGA-G-----GCAACTTTGAC---CGAT-----
Hennzschokkei    ACTGTATGCTGTCTCATGGAGAGACGGGTGGATATAATCA-----AA--
Hennsalminicola  ACTGTATGCTGTCTCATGGAGAGACGGGTGGATATAATCA-----AA--
Ethclone2MA      -----
Ethclone3MA      -----
Ethclone1MA      -----
Myxobprocerus    -----
Mprocerus        -----
Henneguya        AGTGCTTCGTTTGGAGTG-----CTGTATCATGGAG-AGACAACGG
Hsp.             AGT-CGTGAATCCGTGAGAGTTCCGTGCTGTATCATGGAGGAGACAACGG
Hdoori           AGT-CGTGAATCCGTGAGAGTTCCGTGCTGTATCATGGAGGAGACA-CGG
Hennictaluri     AGTGCAATGT-CTTGCAAGAGTCGTGTATTGTATCATGGAG-AGACAACGG
Hennexilis       AGTGCGTG--CTCGCAAGAGTGG-GTATTGTATCATGGAG-AGACAACGG
Mspinacurvatura  AGTGCGTACGTGTGCAGG-----CAGTAT-ATG-----
Michkeulensis    AAGTTGTGCGTTTGCAGG-----CAGTAT-ACG-----
Mosburni         AGAGTAT-TGTTTCGCAAGAGCGATGCTTTGTTTCATGGAG-AGACGTCGG
Muvuliferus      AGAGTAT-TGTTTCGTAAGAGTGGTGTCTTTGT-----
Ceratomyxa       AGTTCAAG-----
Malgonquinensis  GTTGGTCGCTGTCTCATGGGGAGACAACGAGGTATAANCA-----
Perchclone1AM    -----
Perchclone2AM    -----
Perchclone3AM    -----

Melipsoides      -----C-GTTATACGCTGTTCTACTTACCAAAGT-----GG
Mdjragini        -----C-GTTATACGCTGTTCTACTTACCAAAGT-----GG

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Mbramae	-----CCGTTATACGCTGTTCTACTTACCAAAGT-----GG
Mneurobius	-----C-GTTATACGCTGTTCTACTTACCAAAGT-----GG
Marcticus	-----C-GTTATACGCTGTNCTACTTACCAAAGT-----GG
Msandrae	-----C-GTAAAACGCTGTTTAACTACCCAAAGT-----GG
Minsidiosus	-----C--TTAATAACTGTTCTACTTRCCAAAGT-----GG
Mcerebralis	-----C-GTTATACGCTGTTCAACTACCCA--GT-----TG
Mlentisuturalis	-----CTGCCA--AGCAGTGCAAGGCTGAGAAGT-----TG
Mxiao	-----
Myxo.	-----TTGCCG--AGCAGTGGGAGACCGAAAGGT-----TG
Mportucalensis	-----TTGTGA--AGTAGTGTGTAATCTCGTTAT-----TG
Mmusculi	-----TTGTCA--AGCAGTCTTG---GGGTAACC-----TT
Mcyprini	-----TTGTCA--AGCAGTCATG---GGGCAACC-----TA
Mpseudodispar	-----TTGTCA--AGCAGTCATT---GGGCAACC-----TT
Mbibullatus	TGGGGATTGACCGTTGTAAATTATCGGTCATGAAAGAGGAATC-----CC
Melegans	-----TCGTTG--AGCAGTATTT-----GGATTC-----TC
Mpendula	-----CTGTTA--AGCAGATTAG---GGTCTAC-----CT
Mpellicides	-----CTGTTA--AGCAGATTAG---GGTCTAC-----CT
Mhungaricus	-----CCGTTG--TGCAGTGGTG---GGCAAAA-----CC
Myxiditruttae	-----CCGGAGGAAGCGTGGCAATAACAGG-----TC
Myxidium	-----CCGGAGGAAGCGTGGCAATAACAGG-----TC
Ethclone1AM	-----TAGAATGAGTATGGAAGT-----TA
Ethclone3AM	-----TAGAATGAGTATGGAAGT-----TA
Ethclone2AM	-----
Mdiaphanus	-----TGGAAAGGTT---GAATC-----TA
M.lieberkuehni	A-----TTCCTAGTAAGTGCTCGGAATCAACGAGTGCTGATTG
Myxosp.	-----CAGTAGGAGATTAAATTTGG-----A
Hennzschokkei	-----CCACAGGACGGCTGGCAAAAACAGG-----TC
Hennsalminicola	-----CCACAGGACGGCTGGCAAAAACAGG-----TC
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	AACATATTACAAAATCCGAGG--AAGTGTGGCTATAACAGGT-----C
Hsp.	AAAAATATAAGAAAAATCCGAGGAAAGTGTGGCTATAACAGGT-----C
Hdori	AAAA---TAACCAAATCNA---GAAGTGTG-CTATAACAGGT-----C
Hennictaluri	AAAAATATTTTAAAAATCCGAG-GAAGTGTGGCTATAACAGGT-----C
Hennexilis	AATATACTAAAAAAATCCGAG-GAAGTGTGGCTATAACAGGT-----C
Mspinacurvatura	-----TTG---AACTGCT-----
Michkeulensis	-----CTG---AGTTGTT-----
Mosburni	AATATA--TNCAAAATCCGAG-AAAGTGGCGCCATAACAGGT-----C
Muvuliferus	-----
Ceratomyxa	-----CTAGGGGAAGCGTGGCAATAACAGGT-----C
Malgonquinensis	-----AGCTCGAGGAAGAGTGGCTATAACAGGT-----C
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	AGC-----AGTGTGTCATGGA
Mdjrugini	AGC-----AGTGTGTCATGGA
Mbramae	AGC-----AGTGTGTCATGGA
Mneurobius	AGC-----AGTGTGTCATGGA
Marcticus	AGC-----AGTGTGTCATGGA
Msandrae	ACC-----AGTGTGTCATGGA
Minsidiosus	AGC-----AGTGTGTCATGGA
Mcerebralis	AGC-----AGTGTGTCATGGA
Mlentisuturalis	AAC-----ACTGTATCATGAA

Mxiao	-----
Myxo.	AAC-----ACTGTATCATGAA
Mportucalensis	CAC-----AGTACACCATGGA
Mmusculi	TGG-----CTGACTTATGGA
Mcyprini	TGG-----CTGACTTATGGA
Mpseudodispar	TGG-----CTGACTTATGGA
Mbibullatus	TAGTATATGTATTTTATTAGAAATACATAGACTTAGTCCCTGCCCTTTGT
Melegans	CGAG-----TGCTGTCTTATGGG
Mpendula	TA-----ACGCTGCCCTTATGGA
Mpellicides	TA-----ACGCTGCCCTTATGGA
Mhungaricus	TAC-----ACGCTGCCCTTATGGA
Myxiditruttae	TGT-----GATGCCCTTTCGAT
Myxidium	TGT-----GATGCCCTTTCGAT
Ethclone1AM	TAT-----TTTAC-CTCCAGT
Ethclone3AM	TAT-----TTTAC-CTCCAGT
Ethclone2AM	-----
Mdiaphanus	CAT-----TTTGC-CTCCAGT
M.lieberkuehni	CGT-----CCCGC-CCTTTGT
Myxosp.	GAT-----GCTGTACTCCCTTT
Hennzschokkei	TGT-----TATGCCCTTAGAT
Hennsalminicola	TGT-----TATGCCCTTAGAT
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	TGT-----GATGCCCTTAGAT
Hsp.	TGT-----GATGCCCTTAGAT
Hdoori	TGT-----GAT-CCCTTAGAT
Hennictaluri	TGT-----GATGCCCTTAGAT
Hennexilis	TGT-----GATGCCCTTAGAT
Mspinacurvatura	--T-----GGTGTC-----T
Michkeulensis	--T-----GGTGTC-----T
Mosburni	TGT-----GATGCCCTTAGAT
Muvuliferus	-----
Ceratomyxa	TGT-----GATGCCCTTTCGAT
Malgonquinensis	AGT-----GATGCCCTTTCGAT
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	GAGACTGT-----GAGG-----
Mdfragini	GAGACTGT-----GAGG-----
Mbramae	GAGACTGT-----GAGG-----
Mneurobius	GAGACTGT-----GAGG-----
Marcticus	GAGACTGT-----GAGG-----
Msandrae	GAGACTGT-----GAGG-----
Minsidiosus	GAGACTGT-----GAGGAATATATCCAAGC---TC
Mcerebralis	GAGACTGT-----GAGGTATATATCCAAGC---TC
Mlentisuturalis	GAGACAACT-----GTGTATATT-CAAAGAC----AG
Mxiao	-----
Myxo.	GAGACAACT-----GTGGATATAACAAAGAC----AG
Mportucalensis	GAGACTGTT-----GGGTATATC-CAAAGCC----AA
Mmusculi	GGGACTGC-----TGGTGTTACAAAACC---AGA
Mcyprini	GGGACTGC-----TGGTGTTACAAAACC---AGA
Mpseudodispar	GGGACTGC-----TGGTGTTACAAAACC---AGA
Mbibullatus	CACACCGCCCGTCGCTACTACCGAGTGAATGGTGTCATGATGCCTTGGGA

Melegans	GAGACAAC-----GAGGTATATTTAAGCTC---GC
Mpendula	GAGACAAC-----AGGTATATAAAAACCT---GA
Mpellicides	GAGACAAC-----AGGTATATAAAAACCT---GA
Mhungaricus	GAGACAAC-----AGGTATATAAAAGCCT-----
Myxiditruttae	GTTCAGG-----CCGCACG-----
Myxidium	GTCAGGGT-----CCGCACGCGCGCTACA-ATGA
Ethclone1AM	-TAAAC-----AGTGTATAGT-----TAAC
Ethclone3AM	-TAAAC-----AGTGTATAGT-----TAAC
Ethclone2AM	-----
Mdiaphanus	-CAAAC-----ATTGTGTAGT-----TAGC
M.lieberkuehni	ACACACCGCC----CGTCGCTACTACCGAGTGAATGGTGTTCATGA-TGCC
Myxosp.	GCAA-----TAAACGGT-----GGC
Hennzschokkei	ATCCGAG-----GCGGCACGCGCGCTACA-ATGA
Hennsalminicola	ATCCGAG-----GCGGCACGCGCGCTACA-ATGA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	GTTC-AGG-----GCTGCACGCGCGCTACAATG
Hsp.	GNC--AGG-----GCTGCACGCGCGCTACAATG
Hdoori	GTTCGAGG-----GCTGCACGCGCGCTACAATG
Hennictaluri	GTTC-AGG-----GCTGCACGCGCGCTACAATG
Hennexilis	GTTC-AGG-----GCTGCACGCGCGCTACAATG
Mspinacurvatura	GTTC-AGG-----GCAACCTG---GCTGAAGGA
Michkeulensis	GTTC-AGG-----GCAACCTG---GCGGAAGGA
Mosburni	GTTC-AGG-----GCTGCACGCGCGCTACAATG
Muvuliferus	-----
Ceratomyxa	GTTCCTGG-----GCTGCACGCGCGCTACAATG
Malgonquinensis	GTTCAGG-----GCTGCACGCGCGCTACAATG
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdfragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	AATGAAGCTAGGCCATAA---CAGGTCTGTGATGCCCTAAGATGTCCTGG
Mcerebralis	AATGAAGCAAGGCCATAA---CAGGTCTGTGATGCCCTAAGATGTCCTGG
Mlentisuturalis	AGGAAG-TG-----
Mxiao	-----
Myxo.	AGGAAG-TG-----
Mportucalensis	ACGAAGCTG-----
Mmusculi	--GGAAGTG-----
Mcyprini	--GGAAGTG-----
Mpseudodispar	--GGAAGTG-----
Mbibullatus	CTGGACGTGGGATGGATT---CTCGGATCTGTTCTACGCTGGGATCAATG
Melegans	--GGAAGAG-----
Mpendula	--GGAAGTGTGGCTATAA---CAGGTCAGTGATGCCCTTCGATGTTTCGAG
Mpellicides	--GGAAGTGTGGCTATAA---CAGGTCAGTGATGCCCTTCGATGTTTCGAG
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	TAACGACAGCGAGAGTCT---GGAAGTGTGGTCTGGGTAATCTTATGA
Ethclone1AM	TTTTAACTGTGTGCTGTA-----TCATGGAGAGA-----

Ethclone3AM	TTTTAACTGTGTGCTGTA-----TCATGGAGAGA-----
Ethclone2AM	-----
Mdiaphanus	TTTTAGCTGTCATGGTGTA-----TCATGGAGAGA-----
M.lieberkuehni	TTGTGACCGGACGTCGTA---GAGGATGAAAGTCTTCGAACGATGCTGGA
Myxosp.	GGGAAACTGTCCACTGTC-----TTATGGAGAGA-----
Hennzschokkei	TAACGACAGC--GAGTTTC---TAGGTTGAAAGACCTGGGTAATCTT-TGA
Hennsalminicola	TAACGACAGCAGAGTTTC---TAGGTTGAAAGACCTGGGTAATCTT-TGA
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	GCAGCGACAGCGAGTGTT---TGAATCGAAAGATTCGGGTAATCTTGTA
Hsp.	GCGGCGACAGCGAGTGTC---TGGATCGAAAGATTTGGGTAACCTTGTA
Hdoori	G---CGACAGCGAGTTTT---TGGATCGAAAGATTCGAGTAATCTTGTA
Hennictaluri	ACTACGACAGCGAGTCTC---TGGATCGAAAGATTCGGGTAATCTTGTA
Hennexilis	ACTACGACAGCGAGTCTC---TGGATCGAAAGATTCGGGTAATCTTGTA
Mspinacurvatura	A----GGCAGTGAGAC-----GAT-----ACTC-----CCTT-TGG
Michkeulensis	A----GACGATAAGGC-----GAT-----ACTC-----CCTT-TGG
Mosburni	GCAACGACAGCGAGTATC---TGCATCGAAAGATGTGGGTAATCTTGTA
Muvuliferus	-----
Ceratomyxa	GCAGCGACAAAAAGCATCACCTGCTCTGAGAAGAGTGGGAAATCTTTAA
Malgonquinensis	ATAACGTCAGCGAGTGTC---TGCGTCGAAAGACGTGGGTAATCTTT--A
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	GCTGCACGCGCGCTACAATGATGGTGACAGCAAGTTTCTAGGTCGAGAGA
Mcerebralis	GCTGCACGCGCGCTACAATGATGGTGACAGCGAGTTTCTAGGTCGAGAGA
Mlentisuturalis	-----TGGCA--ATAACAGGTCTGTGATGCCCTTAGATGTTTCGGGGCTGC
Mxiao	-----
Myxo.	-----TGGCA--ATAACAG-----
Mportucalensis	-----CGGCA--ATAACAG-----
Mmusculi	-----TGGCG--ATAACAAG-----
Mcyprini	-----TGGCG--ATAACAG-----
Mpseudodispar	-----TGGCG--ATAAC-----
Mbibullatus	TAAAATGGCGCAATTTTCGAGGAAGTAAAA-----
Melegans	-----TGGCT--ATAAAC-----
Mpendula	GCAGCACGCGCGCTACAATGATAACAACAACAAGTTTCTGGGTCGAGAGA
Mpellicides	GCAGCACGCGCGCTACAATGATAACAACAACAAGTTTCTGGGTCGAGAGA
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	ATCGTTATCGTGATGGGGATTGAGCATTGTAATTGTTGCTCATGAAATAG
Ethclone1AM	-----CAACGGAA-ATA-ATAAAAAATC-GAGGAA-----
Ethclone3AM	-----CAACGGAA-ATA-ATAAAAAATC-GAGGAA-----
Ethclone2AM	-----
Mdiaphanus	-----CAACGGAAACATA-ATCAAAATCCGAGGAAGTG-TG-GCTATAACAG
M.lieberkuehni	AT---CAATGTAAAAATG-GCGCAATTTTCGAGGAAGTAAAAAGTCGTAACAA
Myxosp.	-----CAACGGTGAAATACGTCAAAGACCGAGGACGTGTG-GCGATAACAG
Hennzschokkei	ATCGTTATCGTGATGAGGATTGACGGTTGTAATTTTCCGTCATGAAATAG
Hennsalminicola	ATCGTTATCGTGATGAGGATTGACGGTTGTAATTTTCCGTCATGAAATAG

Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	TCG-TTGCCGTGATGGGGATTGACGTTTGTA---ACTCCTCATGAAAGAG
Hsp.	TCG-CCGCCGTGATGGGGATCGACGTTTGTA--AATACGTCGTGAAAGAG
Hdoori	TCGGCCGCCGTGATGGGGATCGACGTTTGTA--AATACGTCGTGAAAGAG
Hennictaluri	TCG-TAGTCGTGATGGGGATCGACGTTTGTA--ATTACGTCGTGAAAGAG
Hennexilis	TCG-TAGTCGTGATGGGGATCGACGTTTGTA--ATTACGTCGTGAAAGAG
Mspinacurvatura	CTA-----CACAAACG-----TATTTATAC-GTTGCATCATGGA-GAG
Michkeulensis	TTA-----CACAAACG-----TATTTATAC-GTTGCATCATGGA-GAG
Mosburni	TCG-TTGCCGTGAAGGGGATTGATGTTTGTA--AATACGTCATGAAAGAG
Muvuliferus	-----
Ceratomyxa	ATCGCTGTCGTGATTGGGGATTGAGCCTTGTAATAATTGCTCATGAAATAG
Malgonquinensis	ATCGTTATCGTGATGGGGATTGACCGTTGTAATTGTCGGTCATGAAAGAG
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	CCCCGGCAATCTTGTAATCACCATCGTGATGGGGATTGACCATTGTAATT
Mcerebralis	CCTGGGCAATCTTGTAATCACCATCGTGATGGGGATTGACCATTGTAATT
Mlentisuturalis	ACGCGCGCTACAATGATGACTACAACAAGCTTCTGGGTCGAGAGACTCGG
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-----
Melegans	-----
Mpendula	CTTGGGTAATCGCTAATTGTTATCGTGATGGGGATTGACGATTGTAATTT
Mpellicides	CTTGGGTAATCGCTAATTGTTATCGTGATGGGGATTGACGATTGTAATTT
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	GAATCCCTTGTATACGTGT-CTCACTTAGGACACGTAGAATGCGTCTCTG
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	GTCTGTGATGCCTAAGATG-TCCTGGAAGGG-----
M.lieberkuehni	GGTT-----TCCGTAGGTGA-ACCTGCAGAAGGATCAAGCTT-----
Myxosp.	G-----
Hennzschokkei	GAATTCCTCGTATGTGTACCGTCATTAGCCGTGTACAGAATAAGTCCCTG
Hennsalminicola	GAATTCCTCGTATGTGTAC-GTCATTAGC-GTGTACAGAATAAGTCCCTG
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	GAATCCCTAGTATGTGCGATTCA--TGAGATCCCCAAATTACGTCCCTGC
Hsp.	GAATNCCTANTACGTGCGATTCA-TGAAATCGCGGGATTTAGTCCCTGC

Hdoori	GAATCCCTAGTACGTGCGATTCAATGAAATCGCGCGGATTTAGTCCCTGC
Hennictaluri	GAATCCCTAGTACGTGCGATTCA-TGAAATCGCGCGGATTTAGTCCCTGC
Hennexilis	GAATCCCTAGTACGTGCGATTCA-TTAAATCGCGCGGATTTAGTCCCTGC
Mspinacurvatura	ACAGC---GGTATAT---ATTCA---AAAAC-CGAGGA---AGT-----
Michkeulensis	ACGGC---GGTATAT---ATTCA---AAAAC-CGAGGA---AGT-----
Mosburni	GAATCCCTAGTAGATGCGATTCA-TGAGATTGCATCGATTAAGTCCCTGC
Muvuliferus	-----
Ceratomyxa	GAATTCCCTCGTAAGCGTGAGTCA-CCAACATCATGTTGAATACGTCTCTGC
Malgonquinensis	GAATCCCTAGTATGCACATTTTA-TTAGAATGTGCAGATTGAGTCCCTGC
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	TGGTCATGAAATAGGAATTCCTTGTAGGCACACTTTTATTAGAGTGTGCC
Mcerebralis	TGGTCATGAAATAGGAATTCCTTGTAGGCACACTTTTATTAGAGTGTGCC
Mlentisuturalis	GTAATCAGTAATAGTCATCGTGATGGGGATTGACCGTTGTAATTATCGGC
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-----
Melegans	-----
Mpendula	CGTCATGAAATAGGAATCCCTAGTATGTGTAATTTATTAAATTGCACAGA
Mpellicides	CGTCATGAAATAGGAATCCCTAGTATGTGTAATTTATTAAATTGCACAGA
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	CCCTTTGTA-----
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	-----
Myxosp.	-----
Hennzschokkei	CCCTTTGTA-----
Hennsalminicola	CCCTTTGTA-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	CCTTTGACACACCCCCGT-----
Hsp.	CCTTTGTA-----
Hdoori	CCTTTGTACACACCGCCCGTCGCTACTACCGAGTGAATGGTGTCATAATG
Hennictaluri	CCTTTGTACACACCGCCCGTCGCTACTACCGAGCGAATGGTGTCATAATA
Hennexilis	CCTTTGTACACACCGCCCGTCGCTACTACCGAGCGAATGGTGTCATAATA
Mspinacurvatura	-----GTA-----GCTATAACAGG-----
Michkeulensis	-----GTA-----GCTATAACAGG-----
Mosburni	CCTTTGTACACACCGCCCGTCGCTACTACCGAGTGAATAGTGTCATAATG
Muvuliferus	-----

Ceratomyxa	CCTTTGTACACACCGCCCGTCGCTAGTACCG-----
Malgonquinensis	CCTTTGTACACACCGCCCGTCGCTACTACCGAGTGAATGGTGTGTCATGATG
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	GAACGAGTCCCTGCCCTCTGTA-----
Mcerebralis	GAACGAGTCCCTGCCCTTTGTA-----
Mlentisuturalis	CATGAAAGAGGAATCCCTAGTATGTGCACGTCATAAGCGTGCGCGGACTG
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-----
Melegans	-----
Mpendula	TTTAGTCCCTGCCCTTTGTACACACCGCCCGTCGCTACTACCGAGTGAAT
Mpellicides	TTTAGTCCCTGCCCTTTGTACACACCGCCCGTCGCTACTACCGAGTGAAT
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	-----
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	-----
Myxosp.	-----
Hennzschokkei	-----
Hennsalminicola	-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-----
Hsp.	-----
Hdoori	CCTTGGAACGGACGTTG-GCTGGGTCTTGTATCCGGA-CAATGCTGCAA
Hennictaluri	CCTTGGAAGCGGACGTCGCGCTGGG-CCTGTGCTCGGAGCGATGCTGCGA
Hennexilis	CCTTGGAAGCGGACGTCGCGCTGGG-CTTATGCTCGGAGCGATGCTGCGA
Mspinacurvatura	-----
Michkeulensis	-----
Mosburni	CCTTGAGAGTGGACGTTTTTCCAG--CGTTCTGTTGGGAAAACGCTACAA
Muvuliferus	-----
Ceratomyxa	-----
Malgonquinensis	CGCTGGGACTGGACGTTA-GATGGGTGTAACAGCTCGTCCGACGCTGGGA
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----

Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	-----
Mcerebralis	-----
Mlentisuturalis	AGTCCCTGCCCTTTGTACACACCGCCCGTCGCTACTACCGAGTGAATGGT
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-----
Melegans	-----
Mpendula	TATGTCATGATGCTCTGGGACTGGACTTTGAAGGGGCTTAACCGTTTTTT
Mpellicides	TATGTCATGATGCTCTGGGACTGGACTTTGAAGGGGCTTAACCGTTTTTT
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	-----
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	-----
Myxosp.	-----
Hennzschokkei	-----
Hennsalminicola	-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-----
Hsp.	-----
Hdoori	TCAGCGTAGAATGGCGCAATTTTCGAGGAAGTAAAAGTCGTAACAAGGTTT
Hennictaluri	TCAGCGTAGAATGGCGCAATTTTCGAGGAAGTAAAAGTCGTAACAAGGTTT
Hennexilis	TCAGCGTAAATGGCGCAATTTTCGAGGAAGTAAAAGTCGTAACAAGGTTT
Mspinacurvatura	-----
Michkeulensis	-----
Mosburni	CCAGCGTAGAATGGCGCAATTTTCGAGGAAGTAAAAGTCGTAACAAGGTTT
Muvuliferus	-----
Ceratomyxa	-----
Malgonquinensis	TCAGCGTAAATGGCGACATTTTCGAGGAAGTAAAAGTCGTAACAAGGTTT
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	-----
Mcerebralis	-----

Mlentisuturalis	ATCGTGATGCCCTTGGGACTGGACGCCGTGCGAGCTTCGGTTCGCAATGGC
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-----
Melegans	-----
Mpendula	CGAAGCTAGAATCAGCGTAAAAATGGCGCAATTTCGAGGAAGTAAAAGTCG
Mpellicides	CGAAGCTAGAATCAGCGTAAAAATGGCGCAATTTCGAGGAAGTAAAAGTCG
Mhungaricus	-----
Myxiditruttae	-----
Myxidium	-----
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	-----
Myxosp.	-----
Hennzschokkei	-----
Hennsalminicola	-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-----
Hsp.	-----
Hdoori	CCGTAGGTGAACCTGCCGAAG-----
Hennictaluri	CCGTAGGTGAACCTGCCGAAGGATC-----
Hennexilis	CCGTAGGTGAACCTGCCGAAGGATC-----
Mspinacurvatura	-----
Michkeulensis	-----
Mosburni	CCGT-----
Muvuliferus	-----
Ceratomyxa	-----
Malgonquinensis	CCGT-----
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdjragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	-----
Mcerebralis	-----
Mlentisuturalis	GCCGGGATCAATGTAAAACGGTGCAATTTCGAGGAAGTAAAAGTCGTAAC
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----

Mbibullatus	-----
Melegans	-----
Mpendula	TAACAAGGTTTCCG-----
Mpellicides	TAACAAGGTTTCCGT-----
Mhungaricus	-----
Myxiditrutiae	-----
Myxidium	-----
Ethclone1AM	-----
Ethclone3AM	-----
Ethclone2AM	-----
Mdiaphanus	-----
M.lieberkuehni	-----
Myxosp.	-----
Hennzschokkei	-----
Hennsalminicola	-----
Ethclone2MA	-----
Ethclone3MA	-----
Ethclone1MA	-----
Myxobprocerus	-----
Mprocerus	-----
Henneguya	-----
Hsp.	-----
Hdoori	-----
Hennictaluri	-----
Hennexilis	-----
Mspinacurvatura	-----
Michkeulensis	-----
Mosburni	-----
Muvuliferus	-----
Ceratomyxa	-----
Malgonquinensis	-----
Perchclone1AM	-----
Perchclone2AM	-----
Perchclone3AM	-----
Melipsoides	-----
Mdfragini	-----
Mbramae	-----
Mneurobius	-----
Marcticus	-----
Msandrae	-----
Minsidiosus	-----
Mcerebralis	-----
Mlentisuturalis	AAGGTTTCCGTAGGTGAACCAGCGGAAG
Mxiao	-----
Myxo.	-----
Mportucalensis	-----
Mmusculi	-----
Mcyprini	-----
Mpseudodispar	-----
Mbibullatus	-----
Melegans	-----
Mpendula	-----
Mpellicides	-----
Mhungaricus	-----
Myxiditrutiae	-----
Myxidium	-----

```

Ethclone1AM -----
Ethclone3AM -----
Ethclone2AM -----
Mdiaphanus -----
M.lieberkuehni -----
Myxosp. -----
Hennzschokkei -----
Hennsalminicola -----
Ethclone2MA -----
Ethclone3MA -----
Ethclone1MA -----
Myxobprocerus -----
Mprocerus -----
Henneguya -----
Hsp. -----
Hdoori -----
Hennictaluri -----
Hennexilis -----
Mspinacurvatura -----
Michkeulensis -----
Mosburni -----
Muvuliferus -----
Ceratomyxa -----
Malgonquinensis -----
Perchclone1AM -----
Perchclone2AM -----
Perchclone3AM -----
;
end;

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Letters of Permission for Articles



15 January 2004

Our ref: HW/ MR/ JAN 04 J165

Dr Russell H Easy
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St. Mary's University

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Council